

GLM as a unified framework for data analysis

Francisco Rodríguez-Sánchez

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So many questions

- **Why** should we really use analysis Y over Z?

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- What if my data are **not Normal**?

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- What even is a **p-value**?

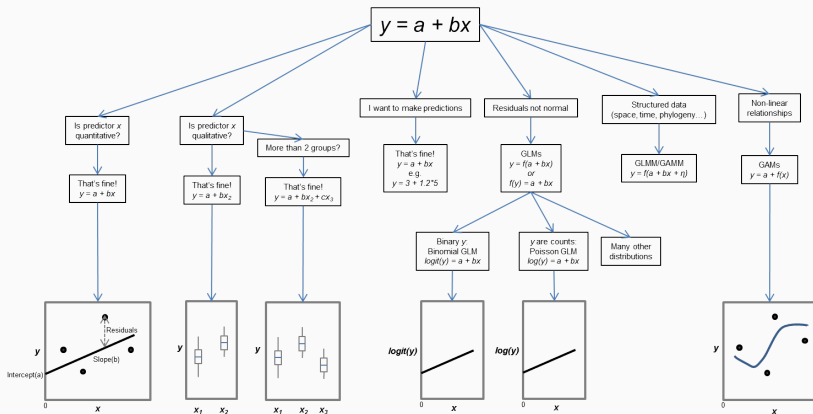
So many questions

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So many questions

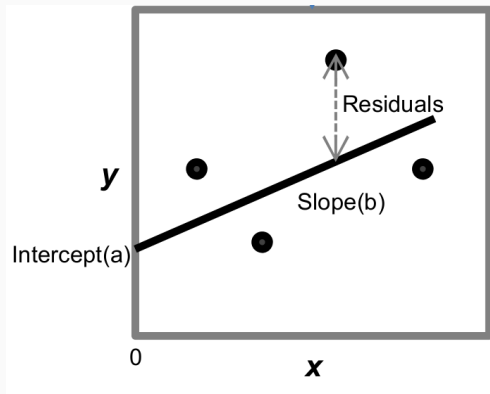
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- How can I take **different factors** into account?
- Can I make **predictions**?

A unified framework



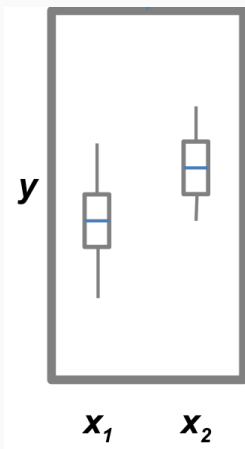
Linear regression

$$y = a + bx$$



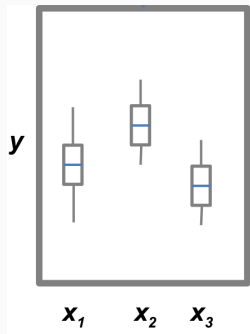
Is predictor X qualitative?

$$y = a + bx_2$$



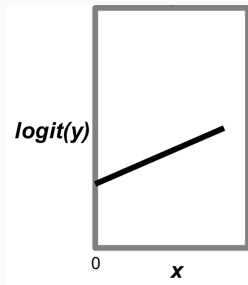
More than 2 groups?

$$y = a + bx_2 + cx_3$$



My data (residuals) are not Normal

$$y = f(a + bx)$$

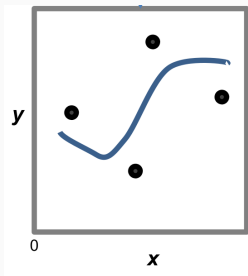


My data are structured (space, time, phylogeny)

$$y = f(a + bx + \eta)$$

Relationships are not linear

$$y = a + f(x)$$



t-tests

ANOVA

regression

.

are special cases of GLM

With GLM we can analyse
many different types of data
using many predictors
(quantitative & qualitative)

Unified, coherent framework for data analysis with many extensions:

- **GLMM** (mixed models): accomodate data structure & variation (space, time, phylogeny)

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Unified, coherent framework for data analysis with many extensions:

- **GLMM** (mixed models): accomodate data structure & variation (space, time, phylogeny)
- **GAMM** (generalised additive models): non-linear relationships
- **Model-based multivariate** statistics

Unified, coherent framework for data analysis with many extensions:

- **GLMM** (mixed models): accomodate data structure & variation (space, time, phylogeny)
- **GAMM** (generalised additive models): non-linear relationships
- **Model-based multivariate** statistics
- **Bayesian** modelling

The Generalised Linear Model (GLM) is a particularly reasonable vantage point on statistical analyses, as **many tests and procedures are special cases** of the GLM. The downside of that (and any other) vantage point is that **we first have to climb it**. There are the morass of unfamiliar terminology, the scree slopes of probability and the cliffs of distributions. **The vista, however, is magnificent.** From the GLM, t-test, ANOVA and regression neatly arrange themselves into regular patterns, and we can see the paths leading towards the horizon: to time series analyses, Bayesian statistics, spatial statistics and so forth.

[Dormann 2020](#)

Introduction to linear models

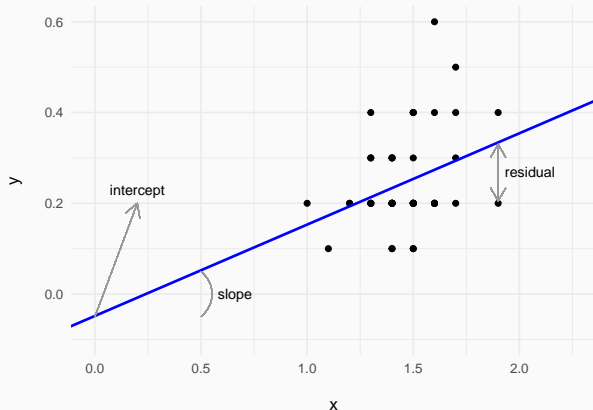
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Our unified regression framework (GLM)

$$y_i = a + bx_i + \varepsilon_i$$

$$\varepsilon_i \sim N(0, \sigma^2)$$



Data

y = response variable

x = predictor

Parameters

a = intercept

b = slope

σ = residual variation

ε = residuals

What's the intercept?

Expected value of y when predictors (x) = 0

If $x = 0$:

- $y = a + b \cdot 0$

What's the intercept?

Expected value of y when predictors $(x) = 0$

If $x = 0$:

- $y = a + b \cdot 0$
- $y = a$

What's the slope?

How much y increases (or decreases) when x increases in 1 unit

If we have model

$$y = 0.5 + 2 * x$$

- If $x = 10 \rightarrow y = 0.5 + 2 * 10 = 20.5$

If x increases 1 unit, y increases 2 units

What's the slope?

How much y increases (or decreases) when x increases in 1 unit

If we have model

$$y = 0.5 + 2 * x$$

- If $x = 10 \rightarrow y = 0.5 + 2 * 10 = 20.5$
- If $x = 11 \rightarrow y = 0.5 + 2 * 11 = 22.5$

If x increases 1 unit, y increases 2 units

Slopes can be negative

If we have model

$$y = 0.5 - 2 * x$$

- If $x = 10 \rightarrow y = 0.5 - 2 * 10 = -19.5$

If x increases 1 unit, y decreases 2 units

Slopes can be negative

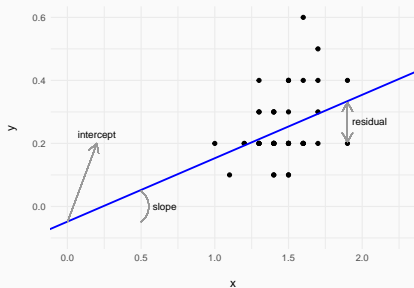
If we have model

$$y = 0.5 - 2 * x$$

- If $x = 10 \rightarrow y = 0.5 - 2 * 10 = -19.5$
- If $x = 11 \rightarrow y = 0.5 - 2 * 11 = -21.5$

If x increases 1 unit, y decreases 2 units

What are residuals?



How far points fall from the regression line

Difference between observed values and values predicted by model
(regression line)

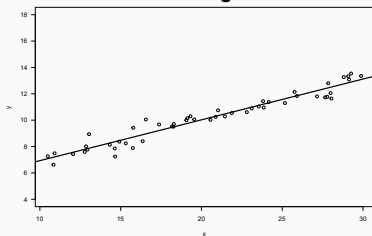
If sigma is large, residuals are larger

$$\varepsilon_i \sim N(0, \sigma^2)$$

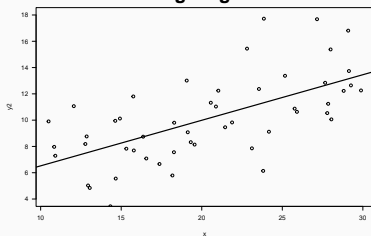
If sigma is larger:

- points farther from regression line
- larger difference of observed - predicted values

small sigma



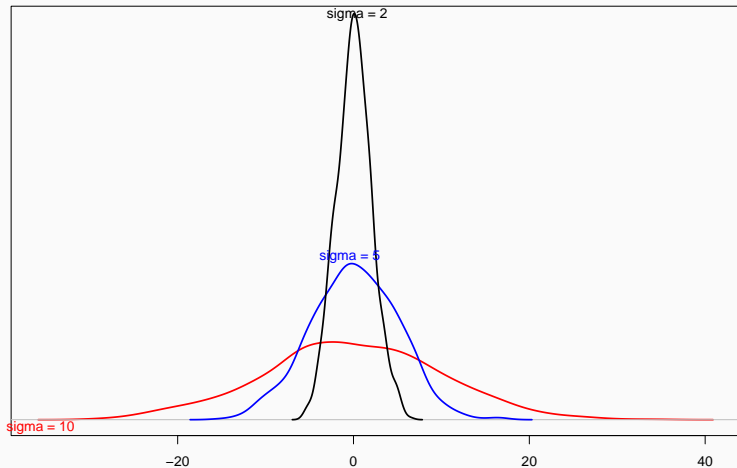
large sigma



Residual variation (sigma) is the Std. Dev. of residuals

$$\varepsilon_i \sim N(0, \sigma^2)$$

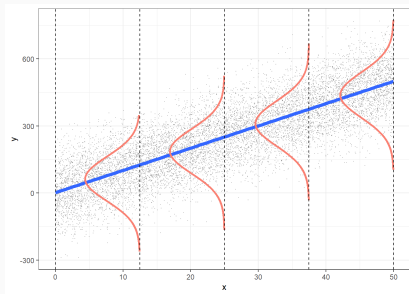
Distribution of residuals



In a general linear model we assume residuals are

$$\varepsilon_i \sim N(0, \sigma^2)$$

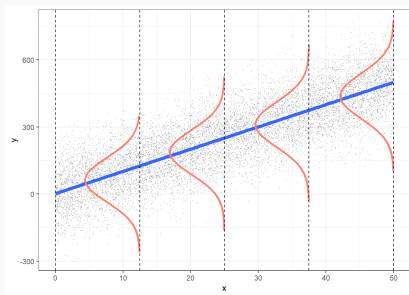
- Normal



In a general linear model we assume residuals are

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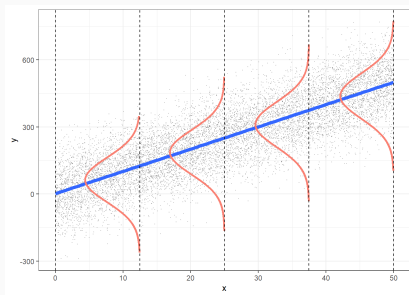
- Normal
- Centred on 0 (no bias)



In a general linear model we assume residuals are

$$\varepsilon_i \sim N(0, \sigma^2)$$

- Normal
- Centred on 0 (no bias)
- Homogeneous variance (*homoscedasticity*)



Different ways to write same model

$$y_i = a + bx_i + \varepsilon_i$$
$$\varepsilon_i \sim N(0, \sigma^2)$$

$$y_i \sim N(\mu_i, \sigma^2)$$
$$\mu_i = a + bx_i$$
$$\varepsilon_i \sim N(0, \sigma^2)$$

<https://pollev.com/franciscorod726>

Linear models

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Example dataset: forest trees

- Download [this dataset](#) (or the entire [zip file](#))

```
trees <- read.csv("data/trees.csv")  
head(trees)
```

	site	dbh	height	sex	dead
1	4	29.68	36.1	male	0
2	5	33.29	42.3	male	0
3	2	28.03	41.9	female	0
4	5	39.86	46.5	female	0
5	1	47.94	43.9	female	0
6	1	10.82	26.2	male	0

Example dataset: forest trees

- Download [this dataset](#) (or the entire [zip file](#))
- Import:

```
trees <- read.csv("data/trees.csv")  
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	site	dbh	height	sex	dead
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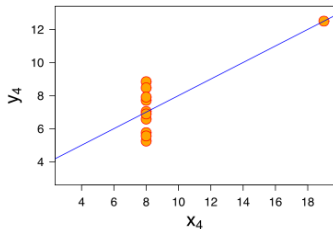
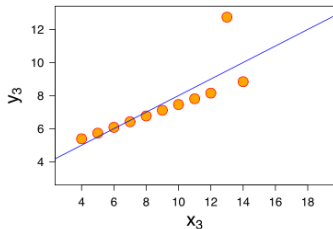
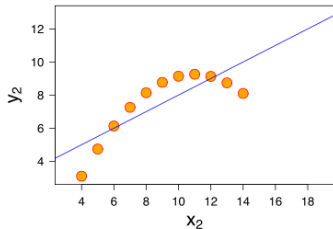
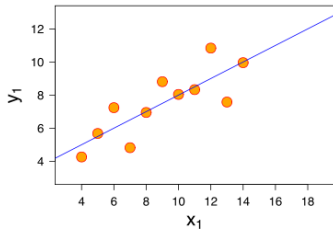
- What is the relationship between DBH and height?

- What is the relationship between DBH and height?
- Do taller trees have bigger trunks?

- What is the relationship between DBH and height?
- Do taller trees have bigger trunks?
- Can we predict height from DBH? How well?

Always plot your data first!

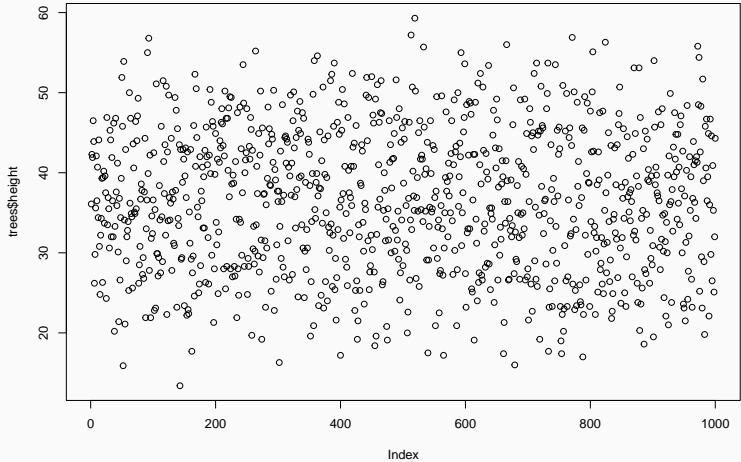
Always plot your data first!



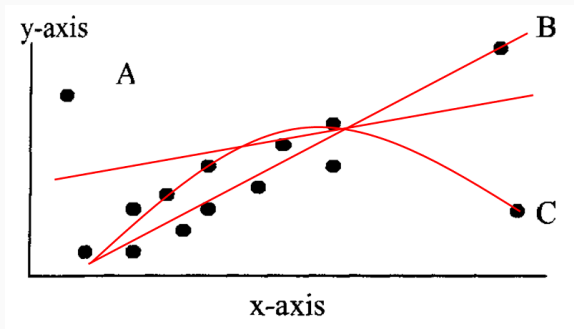
Exploratory Data Analysis (EDA)

Outliers

```
plot(trees$height)
```



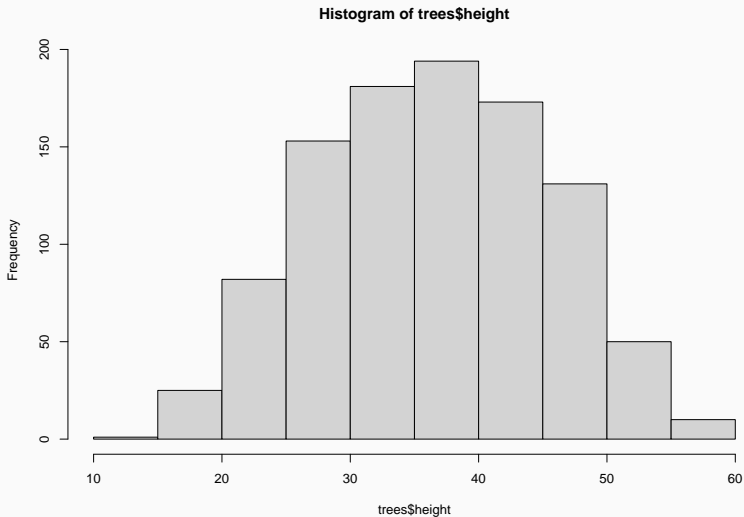
Outliers impact on regression



See <http://rpsychologist.com/d3/correlation/>

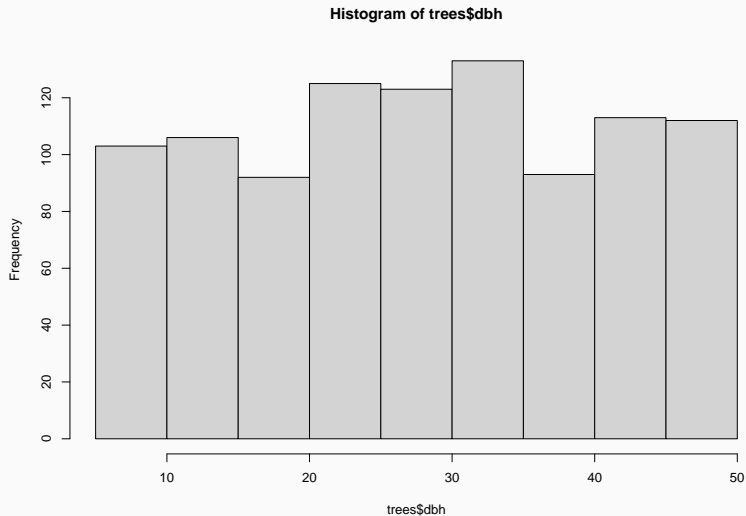
Histogram of response variable

```
hist(trees$height)
```



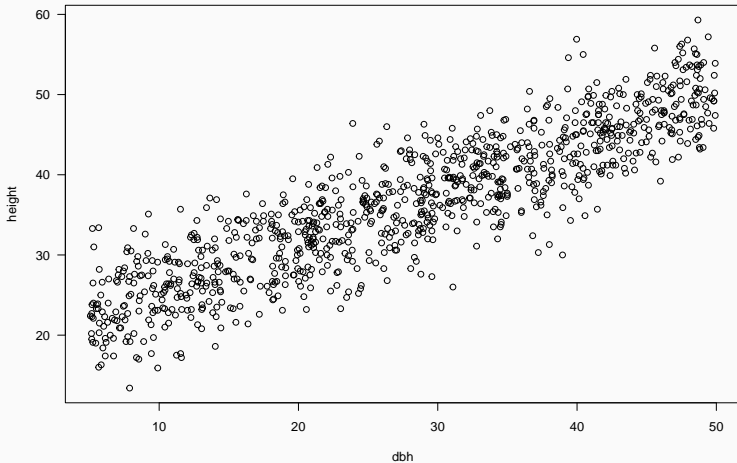
Histogram of predictor variable

```
hist(trees$dbh)
```



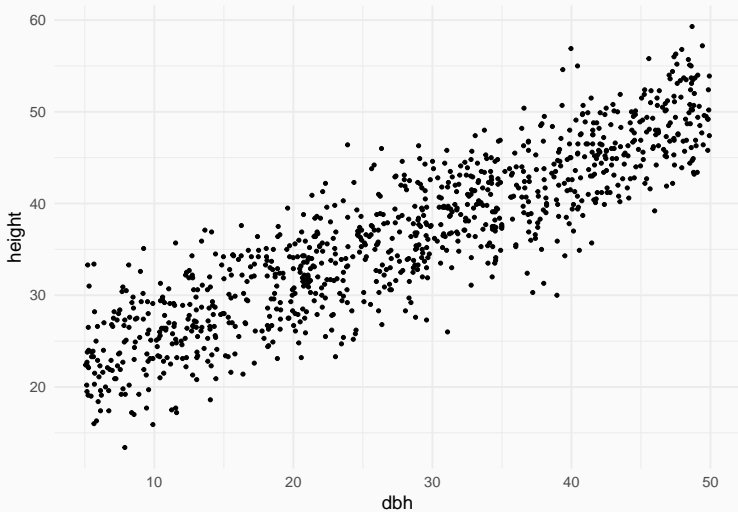
Scatterplot

```
plot(height ~ dbh, data = trees, las = 1)
```



Scatterplot

```
ggplot(trees) +  
  geom_point(aes(dbh, height))
```



Model fitting

Now fit model

Hint: `lm`

Now fit model

Hint: `lm`

```
m1 <- lm(height ~ dbh, data = trees)
```

which corresponds to

$$\text{Height}_i = a + b \cdot \text{DBH}_i + \varepsilon_i$$

$$\varepsilon_i \sim N(0, \sigma^2)$$

Package `equatiomatic` returns model structure

```
library("equatiomatic")  
m1 <- lm(height ~ dbh, data = trees)  
equatiomatic::extract_eq(m1)
```

$$\text{height} = \alpha + \beta_1(\text{dbh}) + \epsilon \quad (1)$$

```
equatiomatic::extract_eq(m1, use_coefs = TRUE)
```

$$\widehat{\text{height}} = 19.34 + 0.62(\text{dbh}) \quad (2)$$

Model interpretation

What does this mean?

```
summary(m1)
```

Call:

```
lm(formula = height ~ dbh, data = trees)
```

Residuals:

Min	1Q	Median	3Q	Max
-13.3270	-2.8978	0.1057	2.7924	12.9511

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	19.33920	0.31064	62.26	<2e-16 ***
dbh	0.61570	0.01013	60.79	<2e-16 ***

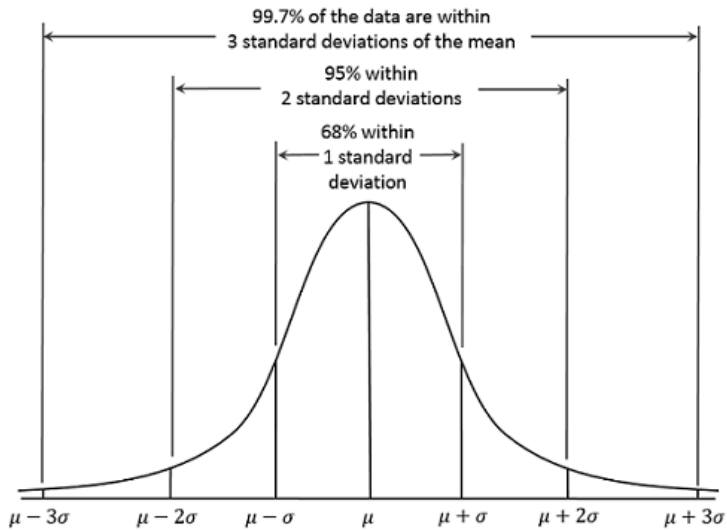
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Residual standard error: 4.093 on 998 degrees of freedom

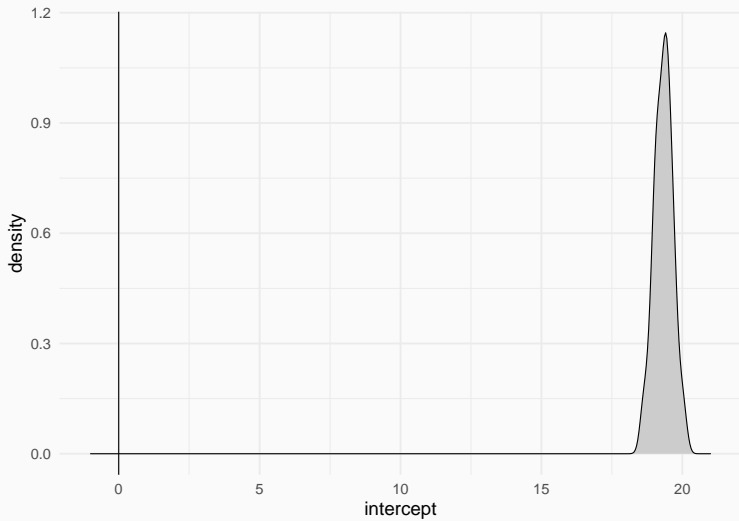
Multiple R-squared: 0.7874, Adjusted R-squared: 0.7871

F-statistic: 3695 on 1 and 998 DF, p-value: < 2.2e-16

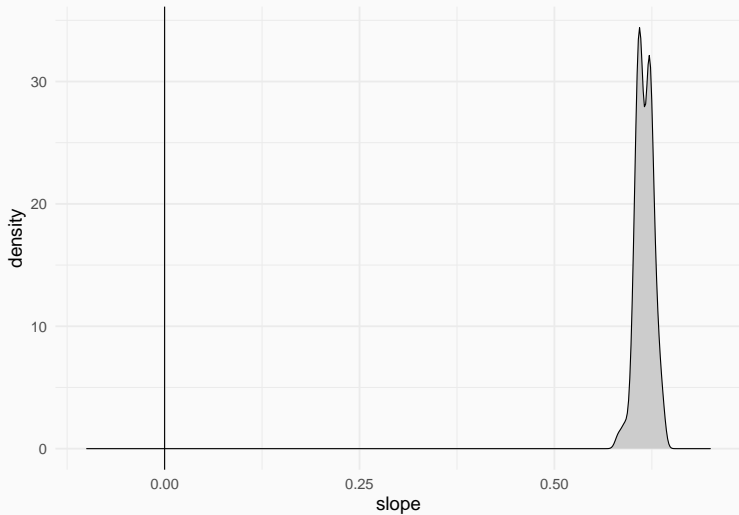
Remember that in a Normal distribution



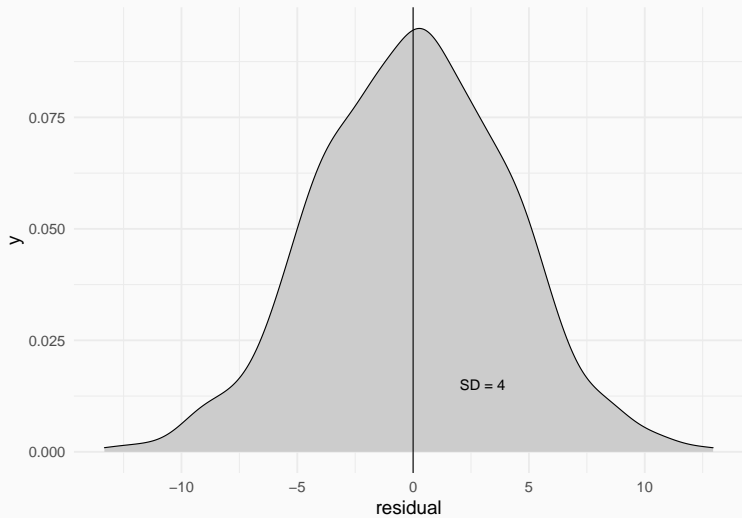
Estimated distribution of the intercept parameter



Estimated distribution of the slope parameter



Distribution of residuals



$$DF = n - p$$

n = sample size

p = number of estimated parameters

Proportion of 'explained' variance

$$R^2 = 1 - \frac{\textit{ResidualVariation}}{\textit{TotalVariation}}$$

Accounts for model complexity (number of parameters)

$$R_{adj}^2 = 1 - (1 - R^2) \frac{n-1}{n-p-1}$$

<https://pollev.com/franciscorod726>

Retrieving model coefficients

```
coef(m1)
```

```
(Intercept)      dbh  
 19.3391968    0.6157036
```


Confidence intervals for parameters

```
confint(m1)
```

```
                2.5 %    97.5 %  
(Intercept) 18.7296053 19.948788  
dbh           0.5958282  0.635579
```

Tidy up model coefficients with broom

```
library("broom")  
tidy(m1)
```

```
# A tibble: 2 x 5  
  term      estimate std.error statistic p.value  
  <chr>      <dbl>    <dbl>    <dbl>    <dbl>  
1 (Intercept) 19.3      0.311     62.3      0  
2 dbh         0.616    0.0101    60.8      0
```

```
glance(m1)
```

```
# A tibble: 1 x 12  
  r.squ~1 adj.r~2 sigma stati~3 p.value    df logLik   AIC   BIC devia~4 df.re~5  
  <dbl>   <dbl> <dbl>   <dbl>   <dbl> <dbl> <dbl> <dbl> <dbl>   <dbl>   <int>  
1  0.787  0.787  4.09  3695.     0     1 -2827. 5660. 5675. 16716.    998  
# ... with 1 more variable: nobs <int>, and abbreviated variable names  
# 1: r.squared, 2: adj.r.squared, 3: statistic, 4: deviance, 5: df.residual
```

<https://broom.tidymodels.org/>

Retrieving model parameters with parameters package

```
library("parameters")  
parameters(m1)
```

Parameter	Coefficient	SE	95% CI	t(998)	p
(Intercept)	19.34	0.31	[18.73, 19.95]	62.26	< .001
dbh	0.62	0.01	[0.60, 0.64]	60.79	< .001

<https://easystats.github.io/parameters/>

Understanding the fitted effects with effects package

```
library("effects")  
summary(allEffects(m1))
```

model: height ~ dbh

dbh effect

dbh	5	20	30	40	50
	22.41771	31.65327	37.81030	43.96734	50.12438

Lower 95 Percent Confidence Limits

dbh	5	20	30	40	50
	21.89682	31.35487	37.55287	43.61733	49.61669

Upper 95 Percent Confidence Limits

dbh	5	20	30	40	50
	22.93861	31.95167	38.06774	44.31735	50.63207

Communicating results



The image shows the top portion of a web page from the journal Nature. At the top left, there is a dark red navigation bar containing a 'MENU' button with a downward arrow and the 'nature' logo with the tagline 'international journal of science'. To the right of this bar is a blue button labeled 'Subs'. Below the navigation bar, the text 'EDITORIAL • 20 MARCH 2019' is displayed. The main title of the article, 'It's time to talk about ditching statistical significance', is written in a large, dark serif font.

- “Never conclude there is ‘no difference’ or ‘no association’ just because $p > 0.05$ or CI includes zero”



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MENU ▾ nature
empirical journal of science Subs

EDITORIAL • 20 MARCH 2019

It's time to talk about ditching statistical significance

- “Never conclude there is ‘no difference’ or ‘no association’ just because $p > 0.05$ or CI includes zero”
- Estimate and communicate **effect sizes and their uncertainty**



The image shows the top portion of a web page from the journal Nature. At the top left, there is a dark red navigation bar containing a 'MENU' button with a downward arrow and the 'nature' logo with the tagline 'international journal of science'. To the right of this bar is a blue button labeled 'Subs'. Below the navigation bar, the text 'EDITORIAL • 20 MARCH 2019' is displayed. The main title of the article, 'It's time to talk about ditching statistical significance', is written in a large, black, serif font.

- “Never conclude there is ‘no difference’ or ‘no association’ just because $p > 0.05$ or CI includes zero”
- Estimate and communicate effect sizes and their uncertainty
- <https://doi.org/10.1038/d41586-019-00857-9>

We found a **significant relationship** between DBH and Height ($p < 0.05$).

We found a significant positive relationship between DBH and Height ($p < 0.05$) ($b = 0.61$, $SE = 0.01$).

Models that describe themselves

```
library("report")  
report(m1)
```

We fitted a linear model (estimated using OLS) to predict height with dbh (formula: $\text{height} \sim \text{dbh}$). The model explains a statistically significant and substantial proportion of variance ($R^2 = 0.79$, $F(1, 998) = 3695.40$, $p < .001$, adj. $R^2 = 0.79$). The model's intercept, corresponding to $\text{dbh} = 0$, is at 19.34 (95% CI [18.73, 19.95], $t(998) = 62.26$, $p < .001$). Within this model:

- The effect of dbh is statistically significant and positive ($\beta = 0.62$, 95% CI [0.60, 0.64], $t(998) = 60.79$, $p < .001$; Std. $\beta = 0.89$, 95% CI [0.86, 0.92])

Standardized parameters were obtained by fitting the model on a standardized version of the dataset. 95% Confidence Intervals (CIs) and p-values were computed using a Wald t-distribution approximation.

<https://easystats.github.io/report/>

Generating table with model results: xtable

```
library("xtable")  
xtable(m1, digits = 2)
```

% latex table generated in R 4.2.1 by xtable 1.8-4 package % Sat Sep 17 21:12:52 2022

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	19.34	0.31	62.26	0.00
dbh	0.62	0.01	60.79	0.00

Generating table with model results: `texreg`

```
library("texreg")
texreg(m1, single.row = TRUE)
```

	Model 1
(Intercept)	19.34 (0.31)***
dbh	0.62 (0.01)***
R ²	0.79
Adj. R ²	0.79
Num. obs.	1000

*** $p < 0.001$; ** $p < 0.01$; * $p < 0.05$

Table 1: Statistical models

Generating table with model results: modelsummary

```
library("modelsummary")  
modelsummary(m1, output = "markdown")
```

	Model 1
(Intercept)	19.339 (0.311)
dbh	0.616 (0.010)
Num.Obs.	1000
R2	0.787
R2 Adj.	0.787
AIC	5660.3
BIC	5675.0
Log.Lik.	-2827.125
F	3695.395
RMSE	4.09

Generating table with model results: gtsummary

```
library("gtsummary")  
tbl_regression(m1, intercept = TRUE)
```

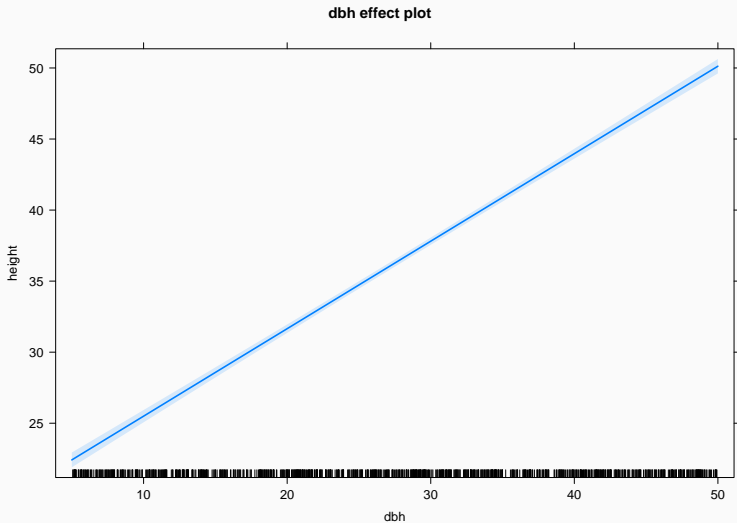
Characteristic	**Beta**	**95% CI**	**p-value**
(Intercept)	19	19, 20	<0.001
dbh	0.62	0.60, 0.64	<0.001

<https://www.danieljsjoberg.com/gtsummary>

Visualising fitted model

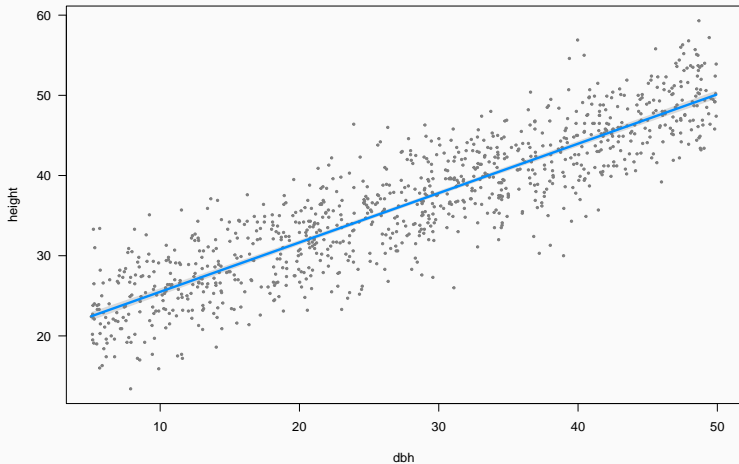
Plot model: effects package

```
library("effects")  
plot(allEffects(m1))
```



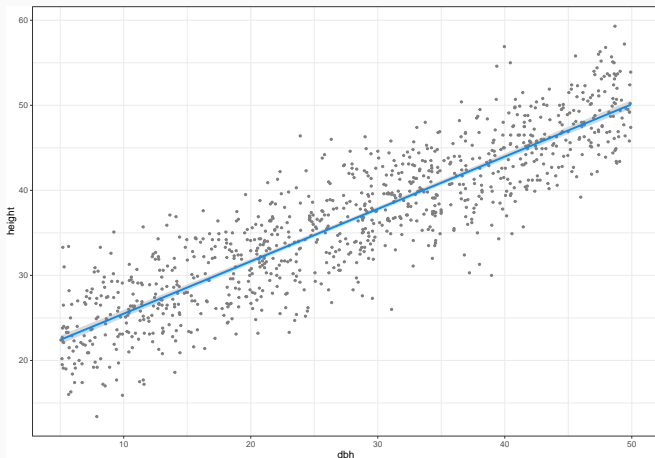
Plot model: visreg

```
library("visreg")  
visreg(m1)
```



visreg can use ggplot2 too

```
visreg(m1, gg = TRUE) + theme_bw()
```

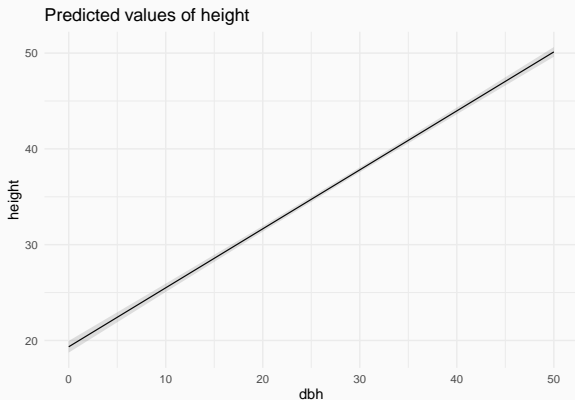


<https://pbreheny.github.io/visreg>

Plot model: sjPlot

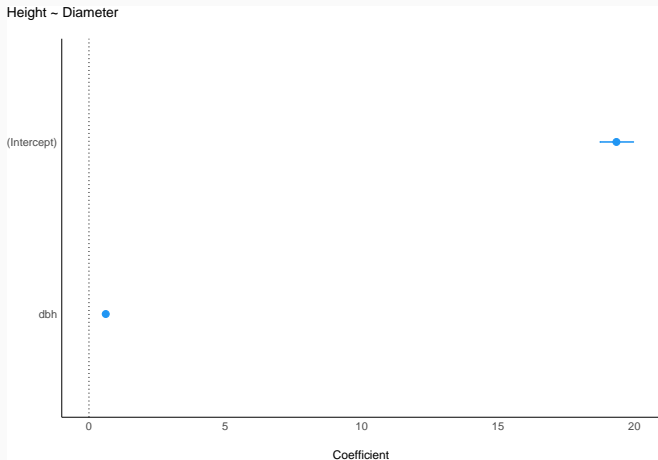
```
library("sjPlot")  
plot_model(m1, type = "eff")
```

\$dbh



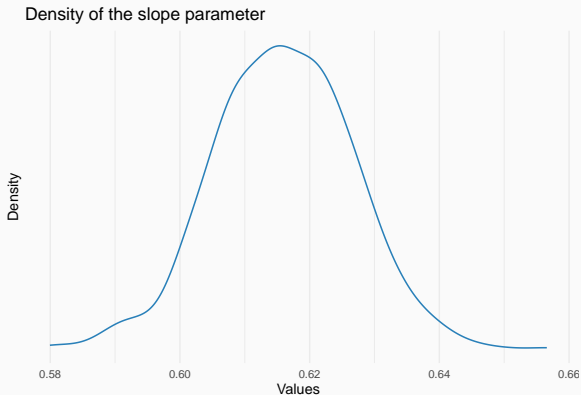
Plot model: see

```
library("see")  
plot(parameters(m1), show_intercept = TRUE) +  
  labs(title = "Height ~ Diameter") # ggplot2
```



Plot parameters' estimated distribution: see

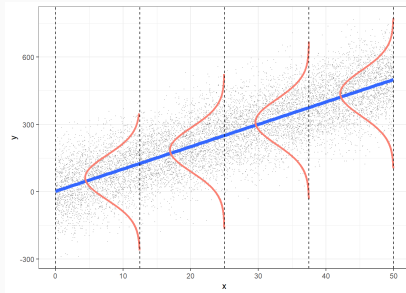
```
plot(simulate_parameters(m1)) +  
  labs(title = "Density of the slope parameter")
```



Model checking

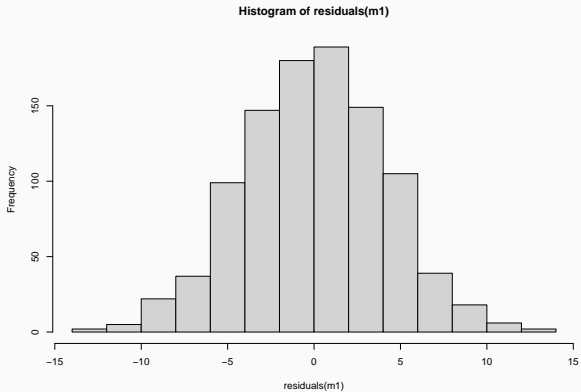
Linear model assumptions

- Linearity (transformations, GAM...)
- Residuals:
 - Independent
 - Equal variance
 - Normal
- Negligible measurement error in predictors



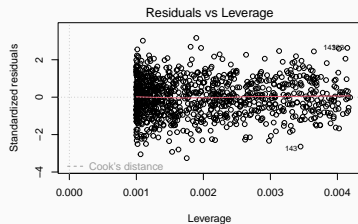
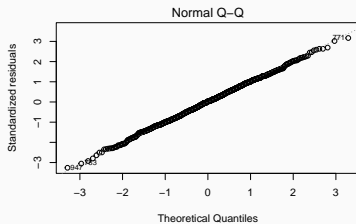
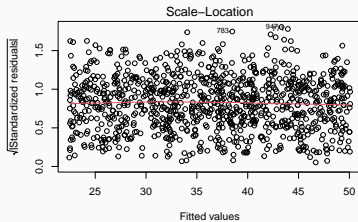
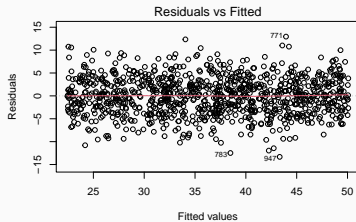
Are residuals normal?

```
hist(residuals(m1))
```



SD = 4.09

Model checking: `plot(model)`

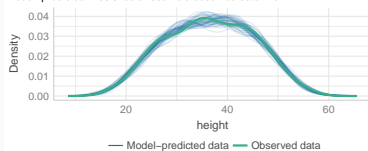


Model checking with performance package

```
library("performance")  
check_model(m1)
```

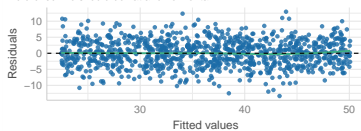
Posterior Predictive Check

Model-predicted lines should resemble observed data line



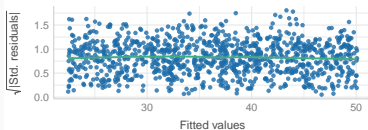
Linearity

Reference line should be flat and horizontal



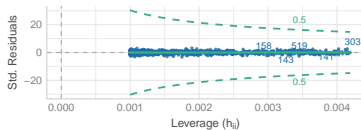
Homogeneity of Variance

Reference line should be flat and horizontal



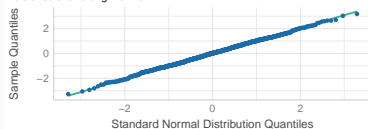
Influential Observations

Points should be inside the contour lines



Normality of Residuals

Dots should fall along the line



A dashboard to explore the full model

```
library("easystats")  
model_dashboard(m1)
```

Using model for prediction

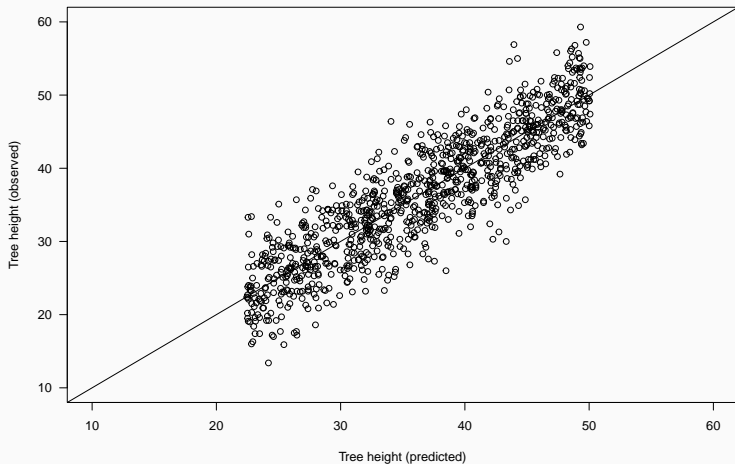
How good is the model in predicting tree height?

`fitted` gives expected value for each observation

```
trees$height.pred <- fitted(m1)
trees$resid <- residuals(m1)
head(trees)
```

	site	dbh	height	sex	dead	height.pred	resid
1	4	29.68	36.1	male	0	37.61328	-1.5132797
2	5	33.29	42.3	male	0	39.83597	2.4640303
3	2	28.03	41.9	female	0	36.59737	5.3026313
4	5	39.86	46.5	female	0	43.88114	2.6188577
5	1	47.94	43.9	female	0	48.85603	-4.9560274
6	1	10.82	26.2	male	0	26.00111	0.1988903

Calibration plot: Observed vs Predicted values



Making predictions for new data

Q: Expected tree height if DBH = 39 cm?

```
new.dbh <- data.frame(dbh = c(39))  
predict(m1, new.dbh, se.fit = TRUE)
```

```
$fit
```

```
1
```

```
43.35164
```

```
$se.fit
```

```
[1] 0.1715514
```

```
$df
```

```
[1] 998
```

```
$residual.scale
```

```
[1] 4.092629
```

Confidence vs Prediction Intervals

Q: Expected tree height if DBH = 39 cm?

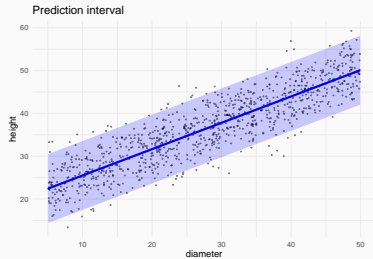
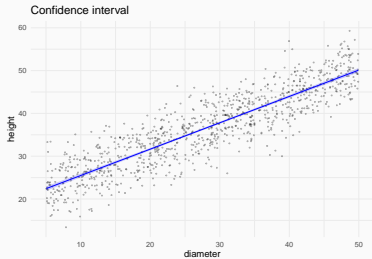
```
predict(m1, new.dbh, interval = "confidence")
```

```
      fit      lwr      upr
1 43.35164 43.01499 43.68828
```

```
predict(m1, new.dbh, interval = "prediction")
```

```
      fit      lwr      upr
1 43.35164 35.31344 51.38983
```


Confidence vs Prediction Intervals



- Visualise data

- Visualise data
- Understand fitted model (summary, allEffects...)

- Visualise data
- Understand fitted model (`summary`, `allEffects`...)
- Visualise model (`plot(allEffects)`, `visreg`, `see`, `plot_model`...)

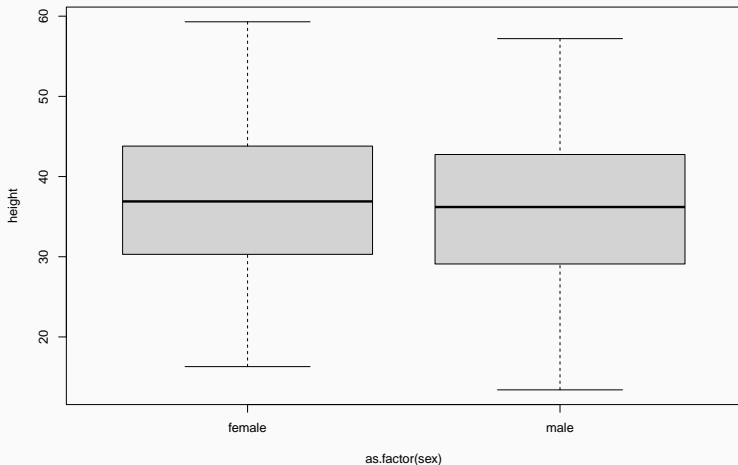
- Visualise data
- Understand fitted model (summary, allEffects...)
- Visualise model (plot(allEffects), visreg, see, plot_model...)
- Check model (plot, check_model, calibration plot...)

- Visualise data
- Understand fitted model (summary, allEffects...)
- Visualise model (plot(allEffects), visreg, see, plot_model...)
- Check model (plot, check_model, calibration plot...)
- Predict (fitted, predict)

Categorical predictors (factors)

Q: Does tree height vary with sex?

```
plot(height ~ as.factor(sex), data = trees)
```



Model height ~ sex

```
m2 <- lm(height ~ sex, data = trees)
```

Call:

```
lm(formula = height ~ sex, data = trees)
```

Residuals:

Min	1Q	Median	3Q	Max
-22.6881	-6.7881	-0.0097	6.7261	22.3687

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	36.9312	0.3981	92.778	<2e-16 ***
sexmale	-0.8432	0.5607	-1.504	0.133

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Residual standard error: 8.865 on 998 degrees of freedom

Multiple R-squared: 0.002261, Adjusted R-squared: 0.001261

F-statistic: 2.261 on 1 and 998 DF, p-value: 0.133

Linear model with categorical predictors

```
m2 <- lm(height ~ sex, data = trees)
```

corresponds to

$$\begin{aligned} \text{Height}_i &= a + b_{\text{male}} + \varepsilon_i \\ \varepsilon_i &\sim N(0, \sigma^2) \end{aligned}$$

Model height ~ sex

```
m2 <- lm(height ~ sex, data = trees)
```

Call:

```
lm(formula = height ~ sex, data = trees)
```

Residuals:

	Min	1Q	Median	3Q	Max
	-22.6881	-6.7881	-0.0097	6.7261	22.3687

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	36.9312	0.3981	92.778	<2e-16 ***
sexmale	-0.8432	0.5607	-1.504	0.133

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

<https://pollev.com/franciscorod726>

Let's read the model report...

`report(m2)`

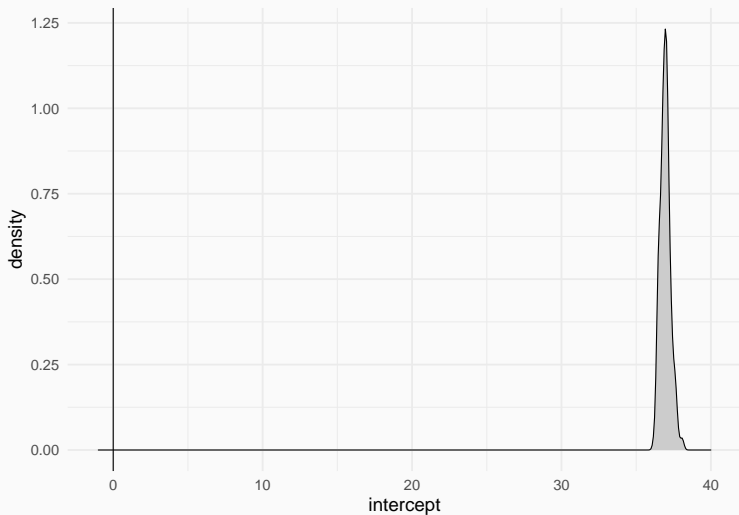
We fitted a linear model (estimated using OLS) to predict height with sex (formula: `height ~ sex`). The model explains a statistically not significant and very weak proportion of variance ($R^2 = 2.26e-03$, $F(1, 998) = 2.26$, $p = 0.133$, adj. $R^2 = 1.26e-03$). The model's intercept, corresponding to sex = female, is at 36.93 (95% CI [36.15, 37.71], $t(998) = 92.78$, $p < .001$). Within this model:

- The effect of sex [male] is statistically non-significant and negative (beta = -0.84, 95% CI [-1.94, 0.26], $t(998) = -1.50$, $p = 0.133$; Std. beta = -0.10, 95% CI [-0.22, 0.03])

Standardized parameters were obtained by fitting the model on a standardized version of the dataset. 95% Confidence Intervals (CIs) and p-values were computed using a Wald t-distribution approximation.

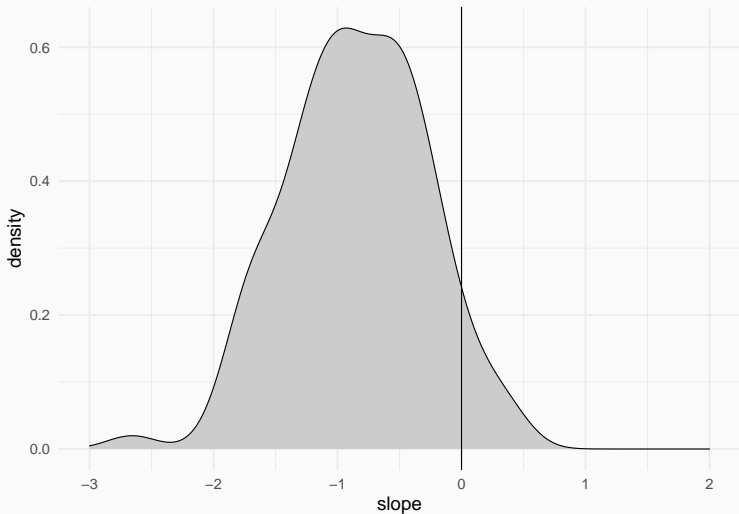
Estimated distribution of the intercept parameter

Intercept = Height of females



Estimated distribution of the *beta* parameter

beta = height difference of males vs females



Analysing differences among factor levels

```
library("modelbased")  
estimate_means(m2)
```

Estimated Marginal Means

sex	Mean	SE	95% CI
male	36.09	0.39	[35.31, 36.86]
female	36.93	0.40	[36.15, 37.71]

Marginal means estimated at sex

Analysing differences among factor levels

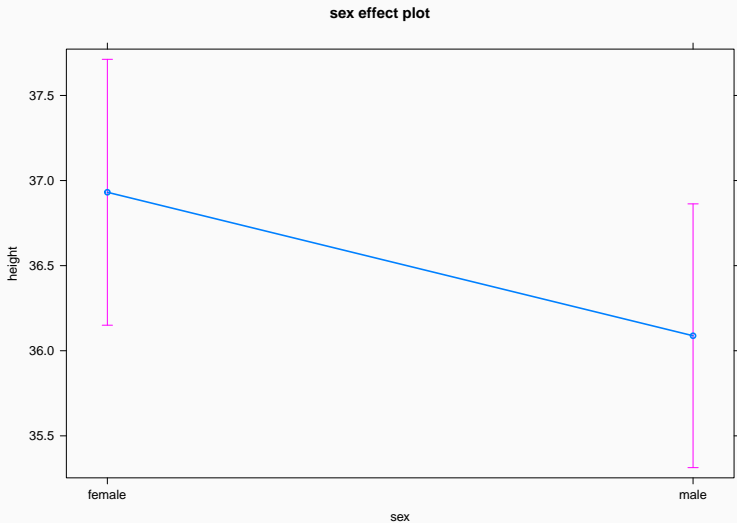
```
estimate_contrasts(m2)
```

Marginal Contrasts Analysis

Level1	Level2	Difference	95% CI	SE	t(998)	p
male	female	-0.84	[-1.94, 0.26]	0.56	-1.50	0.133

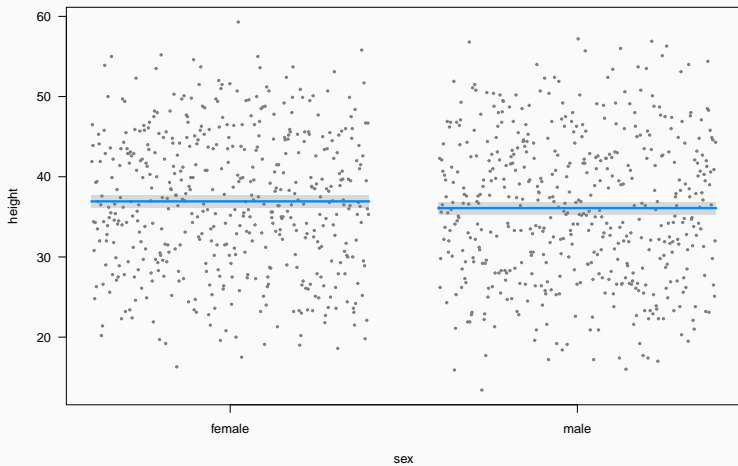
Marginal contrasts estimated at sex
p-value adjustment method: Holm (1979)

```
plot(allEffects(m2))
```



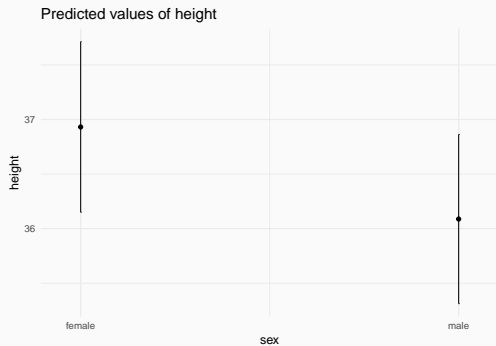
Plot (visreg)

`visreg(m2)`

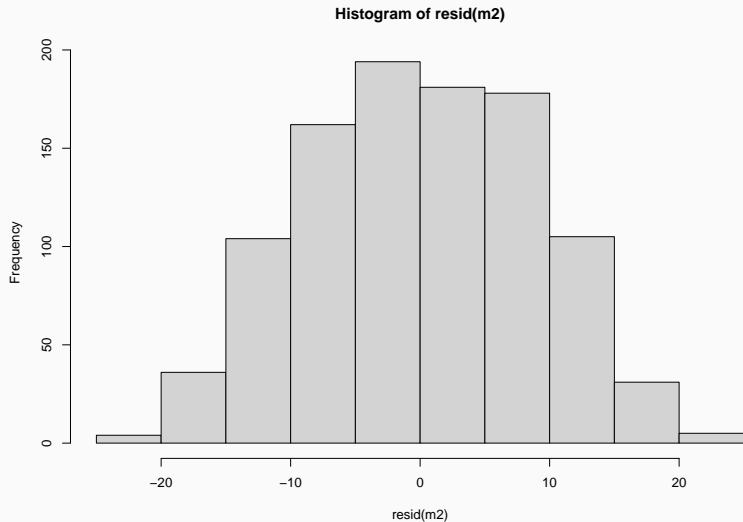


```
plot_model(m2, type = "eff")
```

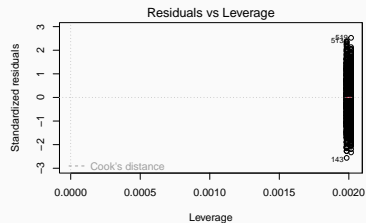
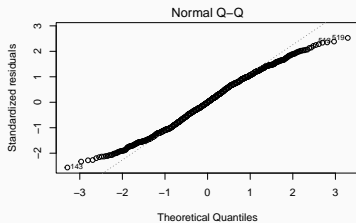
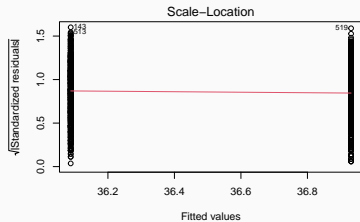
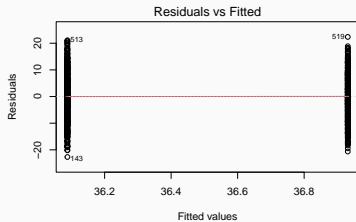
`$sex`



```
hist(resid(m2))
```



Model checking: residuals

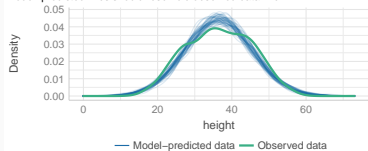


Model checking

```
library("performance")  
check_model(m2)
```

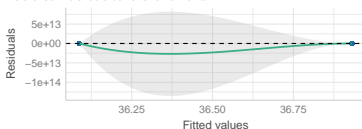
Posterior Predictive Check

Model-predicted lines should resemble observed data line



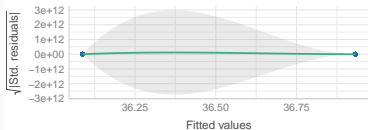
Linearity

Reference line should be flat and horizontal



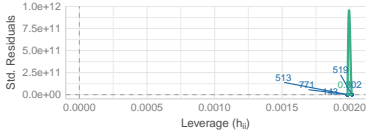
Homogeneity of Variance

Reference line should be flat and horizontal



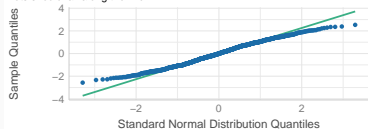
Influential Observations

Points should be inside the contour lines



Normality of Residuals

Dots should fall along the line

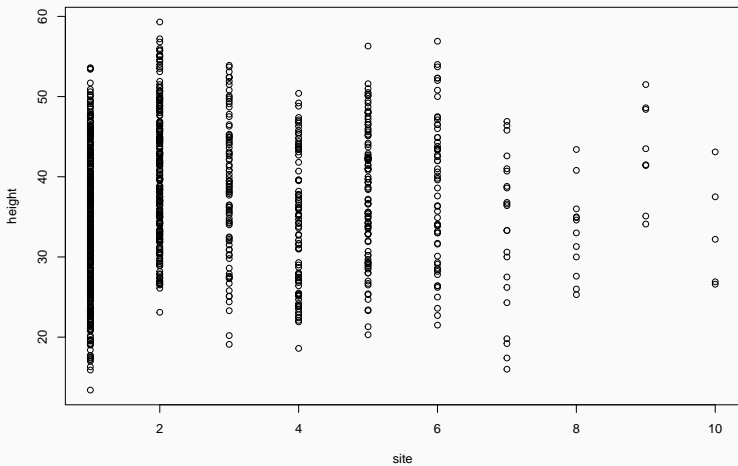


```
model_dashboard(m2)
```


Q: Does height differ among field sites?

Plot data first

```
plot(height ~ site, data = trees)
```



Linear model with categorical predictors

```
m3 <- lm(height ~ site, data = trees)
```

$$y_i = a + b_{site2} + c_{site3} + d_{site4} + e_{site5} + \dots + \varepsilon_i$$
$$\varepsilon_i \sim N(0, \sigma^2)$$

All right here?

```
m3 <- lm(height ~ site, data = trees)
```

Call:

```
lm(formula = height ~ site, data = trees)
```

Residuals:

Min	1Q	Median	3Q	Max
-22.4498	-6.7049	0.0709	6.7537	23.0640

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	35.4636	0.4730	74.975	< 2e-16 ***
site	0.3862	0.1413	2.733	0.00639 **

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Residual standard error: 8.842 on 998 degrees of freedom

Multiple R-squared: 0.007429, Adjusted R-squared: 0.006435

F-statistic: 7.47 on 1 and 998 DF, p-value: 0.006385

```
extract_eq(m3)
```

$$\text{height} = \alpha + \beta_1(\text{site}) + \epsilon \quad (3)$$

site is a factor!

```
trees$site <- as.factor(trees$site)
```

Let's check model structure with `equatiomatic`

```
m3 <- lm(height ~ site, data = trees)
extract_eq(m3)
```

$$\text{height} = \alpha + \beta_1(\text{site}_2) + \beta_2(\text{site}_3) + \beta_3(\text{site}_4) + \beta_4(\text{site}_5) + \beta_5(\text{site}_6) + \beta_6(\text{site}_7) + \beta_7(\text{site}_8) \quad (4)$$

Model Height ~ site

Call:

```
lm(formula = height ~ site, data = trees)
```

Residuals:

Min	1Q	Median	3Q	Max
-20.4416	-6.9004	0.0379	6.3051	19.7584

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)	
(Intercept)	33.8416	0.4266	79.329	< 2e-16	***
site2	6.3411	0.7126	8.899	< 2e-16	***
site3	4.9991	0.9828	5.086	4.36e-07	***
site4	0.5329	0.9872	0.540	0.58949	
site5	4.3723	0.9425	4.639	3.97e-06	***
site6	4.7601	1.1709	4.065	5.18e-05	***
site7	-0.7416	1.8506	-0.401	0.68871	
site8	-0.6832	2.4753	-0.276	0.78258	
site9	9.1709	3.0165	3.040	0.00243	**
site10	-0.5816	3.8013	-0.153	0.87843	

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

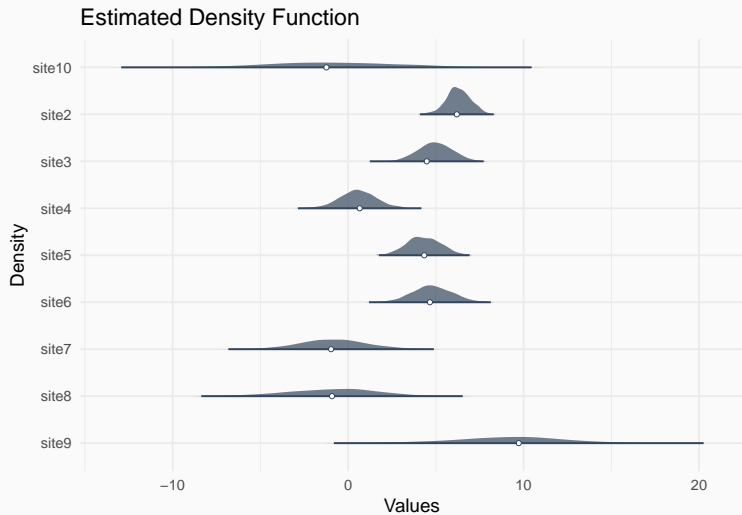
Residual standard error: 8.446 on 990 degrees of freedom

Multiple R-squared: 0.1016, Adjusted R-squared: 0.09344

F-statistic: 12.44 on 9 and 990 DF, p-value: < 2.2e-16

Estimated parameter distributions

```
plot(simulate_parameters(m3), stack = FALSE)
```



Analysing differences among factor levels

```
library("modelbased")  
estimate_means(m3)
```

Estimated Marginal Means

site	Mean	SE	95% CI
1	33.84	0.43	[33.00, 34.68]
2	40.18	0.57	[39.06, 41.30]
3	38.84	0.89	[37.10, 40.58]
4	34.37	0.89	[32.63, 36.12]
5	38.21	0.84	[36.56, 39.86]
6	38.60	1.09	[36.46, 40.74]
7	33.10	1.80	[29.57, 36.63]
8	33.16	2.44	[28.37, 37.94]
9	43.01	2.99	[37.15, 48.87]
10	33.26	3.78	[25.85, 40.67]

Analysing differences among factor levels

For finer control see `emmeans` package

```
estimate_contrasts(m3)
```

Marginal Contrasts Analysis

Level1	Level2	Difference	95% CI	SE	t(990)	p
site1	site10	0.58	[-11.85, 13.01]	3.80	0.15	> .999
site1	site2	-6.34	[-8.67, -4.01]	0.71	-8.90	< .001
site1	site3	-5.00	[-8.21, -1.78]	0.98	-5.09	< .001
site1	site4	-0.53	[-3.76, 2.70]	0.99	-0.54	> .999
site1	site5	-4.37	[-7.45, -1.29]	0.94	-4.64	< .001
site1	site6	-4.76	[-8.59, -0.93]	1.17	-4.07	0.002
site1	site7	0.74	[-5.31, 6.79]	1.85	0.40	> .999
site1	site8	0.68	[-7.41, 8.78]	2.48	0.28	> .999
site1	site9	-9.17	[-19.04, 0.69]	3.02	-3.04	0.073
site2	site10	6.92	[-5.57, 19.42]	3.82	1.81	0.728
site2	site3	1.34	[-2.10, 4.79]	1.05	1.27	0.959
site2	site4	5.81	[2.35, 9.27]	1.06	5.49	< .001
site2	site5	1.97	[-1.35, 5.29]	1.02	1.94	0.643
site2	site6	1.58	[-2.44, 5.61]	1.23	1.28	0.957
site2	site7	7.08	[0.90, 13.26]	1.89	3.75	0.007
site2	site8	7.02	[-1.17, 15.21]	2.50	2.81	0.136
site2	site9	-2.83	[-12.77, 7.11]	3.04	-0.93	0.995
site3	site10	5.58	[-7.11, 18.27]	3.88	1.44	0.915
site3	site4	4.47	[0.36, 8.57]	1.26	3.56	0.014
site3	site5	0.63	[-3.37, 4.62]	1.22	0.51	> .999
site3	site6	0.24	[-4.35, 4.83]	1.40	0.17	> .999
site3	site7	5.74	[-0.82, 12.30]	2.01	2.86	0.118
site3	site8	5.68	[-2.80, 14.17]	2.59	2.19	0.464
site3	site9	-4.17	[-14.36, 6.01]	3.11	-1.34	0.944
site4	site10	1.11	[-11.58, 13.81]	3.88	0.29	> .999
site4	site5	-3.84	[-7.84, 0.16]	1.22	-3.14	0.055
site4	site6	-4.23	[-8.83, 0.38]	1.41	-3.00	0.081

Presenting model results

```
kable(xtable::xtable(m3), digits = 2)
```

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	33.84	0.43	79.33	0.00
site2	6.34	0.71	8.90	0.00
site3	5.00	0.98	5.09	0.00
site4	0.53	0.99	0.54	0.59
site5	4.37	0.94	4.64	0.00
site6	4.76	1.17	4.07	0.00
site7	-0.74	1.85	-0.40	0.69
site8	-0.68	2.48	-0.28	0.78
site9	9.17	3.02	3.04	0.00
site10	-0.58	3.80	-0.15	0.88

Estimated tree heights for each site

```
summary(allEffects(m3))
```

```
model: height ~ site
```

```
site effect
```

```
site
```

	1	2	3	4	5	6	7	8
	33.84158	40.18265	38.84066	34.37444	38.21386	38.60167	33.10000	33.15833
	9	10						
	43.01250	33.26000						

```
Lower 95 Percent Confidence Limits
```

```
site
```

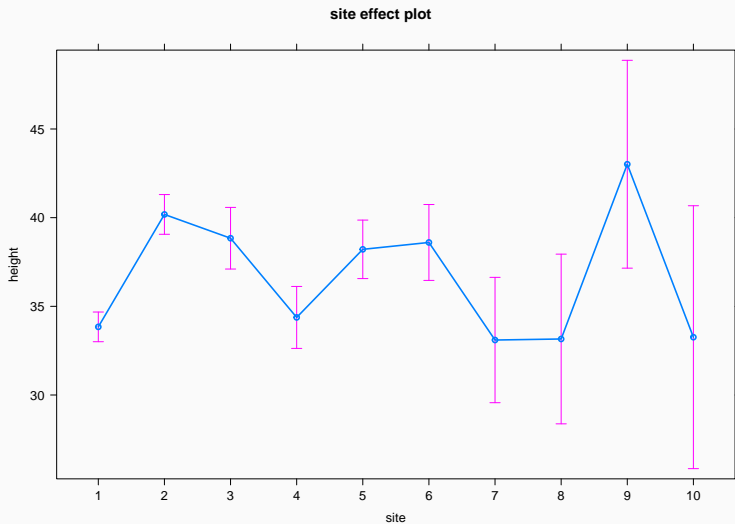
	1	2	3	4	5	6	7	8
	33.00444	39.06264	37.10317	32.62733	36.56463	36.46190	29.56629	28.37367
	9	10						
	37.15251	25.84764						

```
Upper 95 Percent Confidence Limits
```

```
site
```

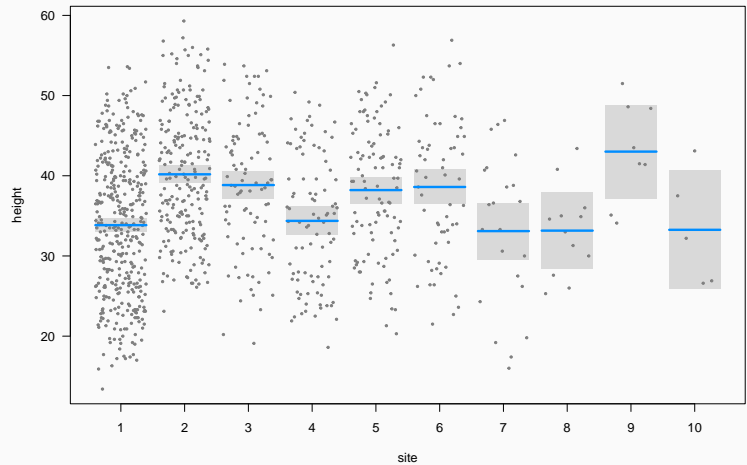
	1	2	3	4	5	6	7	8
	34.67872	41.30265	40.57814	36.12156	39.86309	40.74143	36.63371	37.94299
	9	10						
	48.87249	40.67236						

```
plot(allEffects(m3))
```



Plot (visreg)

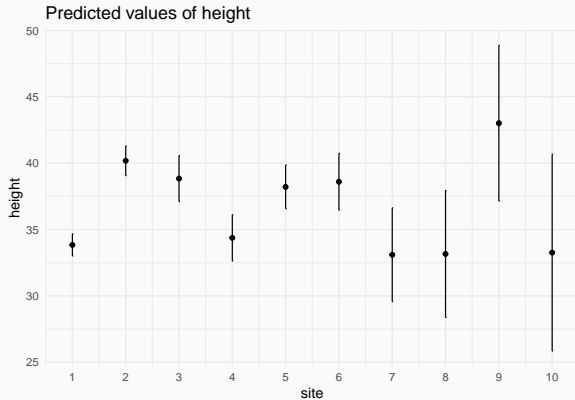
visreg(m3)



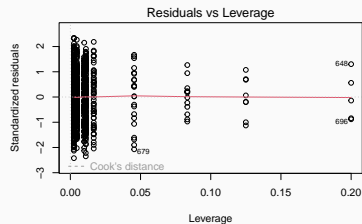
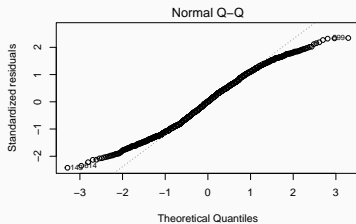
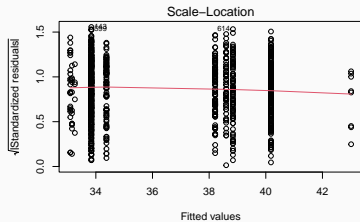
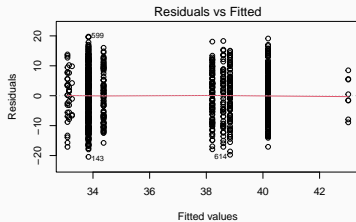
Plot model (sjPlot)

```
plot_model(m3, type = "eff")
```

`$site`



Model checking: residuals

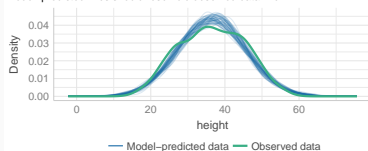


Model checking: residuals

check_model(m3)

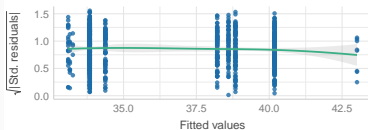
Posterior Predictive Check

Model-predicted lines should resemble observed data line



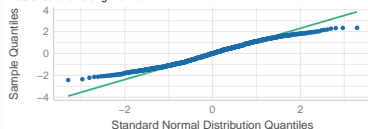
Homogeneity of Variance

Reference line should be flat and horizontal



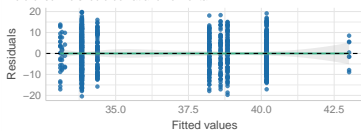
Normality of Residuals

Dots should fall along the line



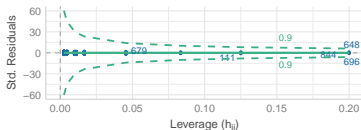
Linearity

Reference line should be flat and horizontal



Influential Observations

Points should be inside the contour lines



Combining continuous and categorical predictors

Predicting tree height based on dbh and site

```
lm(height ~ site + dbh, data = trees)
```

corresponds to

$$y_i = a + b_{site2} + c_{site3} + d_{site4} + e_{site5} + \dots + k \cdot DBH_i + \varepsilon_i$$
$$\varepsilon_i \sim N(0, \sigma^2)$$

Predicting tree height based on dbh and site

Call:

```
lm(formula = height ~ site + dbh, data = trees)
```

Residuals:

Min	1Q	Median	3Q	Max
-10.1130	-1.9885	0.0582	2.0314	11.3320

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)	
(Intercept)	16.699037	0.260565	64.088	< 2e-16	***
site2	6.504303	0.256730	25.335	< 2e-16	***
site3	4.357457	0.354181	12.303	< 2e-16	***
site4	1.934650	0.356102	5.433	6.98e-08	***
site5	3.637432	0.339688	10.708	< 2e-16	***
site6	4.204511	0.421906	9.966	< 2e-16	***
site7	-0.176193	0.666772	-0.264	0.7916	
site8	-5.312648	0.893603	-5.945	3.82e-09	***
site9	5.437049	1.087766	4.998	6.84e-07	***
site10	2.263338	1.369986	1.652	0.0988	.
dbh	0.617075	0.007574	81.473	< 2e-16	***

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Residual standard error: 3.043 on 989 degrees of freedom

Multiple R-squared: 0.8835, Adjusted R-squared: 0.8823

Presenting model results

```
parameters(m4)
```

Parameter	Coefficient	SE	95% CI	t(989)	p
(Intercept)	16.70	0.26	[16.19, 17.21]	64.09	< .001
site [2]	6.50	0.26	[6.00, 7.01]	25.34	< .001
site [3]	4.36	0.35	[3.66, 5.05]	12.30	< .001
site [4]	1.93	0.36	[1.24, 2.63]	5.43	< .001
site [5]	3.64	0.34	[2.97, 4.30]	10.71	< .001
site [6]	4.20	0.42	[3.38, 5.03]	9.97	< .001
site [7]	-0.18	0.67	[-1.48, 1.13]	-0.26	0.792
site [8]	-5.31	0.89	[-7.07, -3.56]	-5.95	< .001
site [9]	5.44	1.09	[3.30, 7.57]	5.00	< .001
site [10]	2.26	1.37	[-0.43, 4.95]	1.65	0.099
dbh	0.62	7.57e-03	[0.60, 0.63]	81.47	< .001

Estimated tree heights for each site

```
summary(allEffects(m4))
```

```
model: height ~ site + dbh
```

```
site effect
```

```
site
```

	1	2	3	4	5	6	7	8
	33.90437	40.40868	38.26183	35.83902	37.54181	38.10889	33.72818	28.59173
	9	10						
	39.34142	36.16771						

```
Lower 95 Percent Confidence Limits
```

```
site
```

	1	2	3	4	5	6	7	8
	33.60276	40.00512	37.63569	35.20858	36.94739	37.33787	32.45495	26.86438
	9	10						
	37.22831	33.49623						

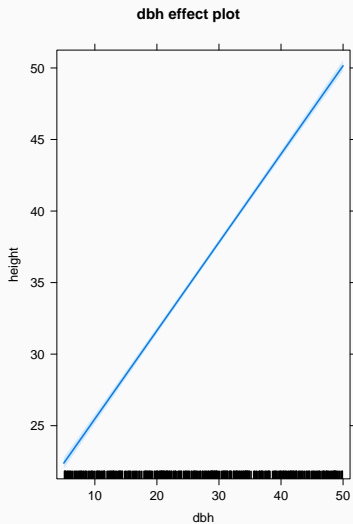
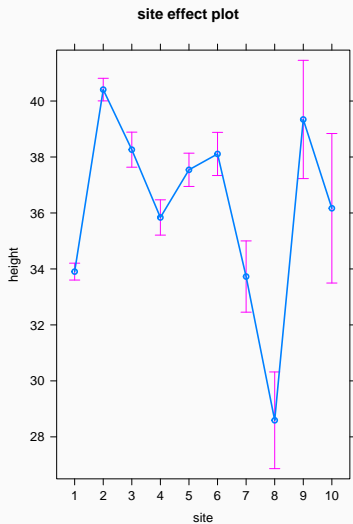
```
Upper 95 Percent Confidence Limits
```

```
site
```

	1	2	3	4	5	6	7	8
	34.20599	40.81223	38.88798	36.46947	38.13622	38.87990	35.00141	30.31907
	9	10						
	41.45454	38.83919						

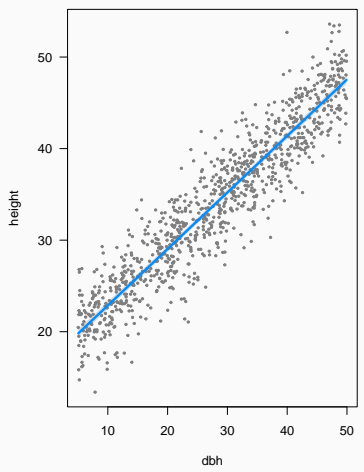
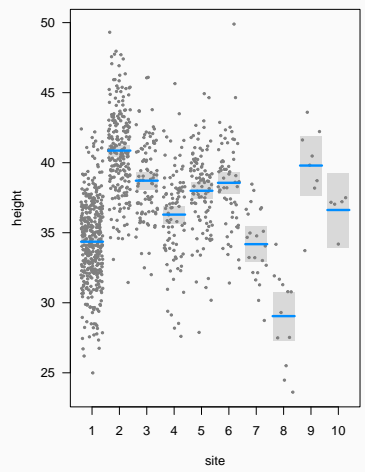
```
dbh effect
```

```
plot(allEffects(m4))
```



Plot (visreg)

```
visreg(m4)
```

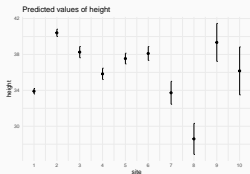


null device
1

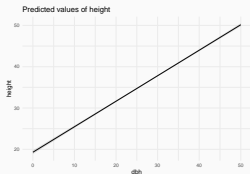
Plot model (sjPlot)

```
plot_model(m4, type = "eff")
```

`$site`

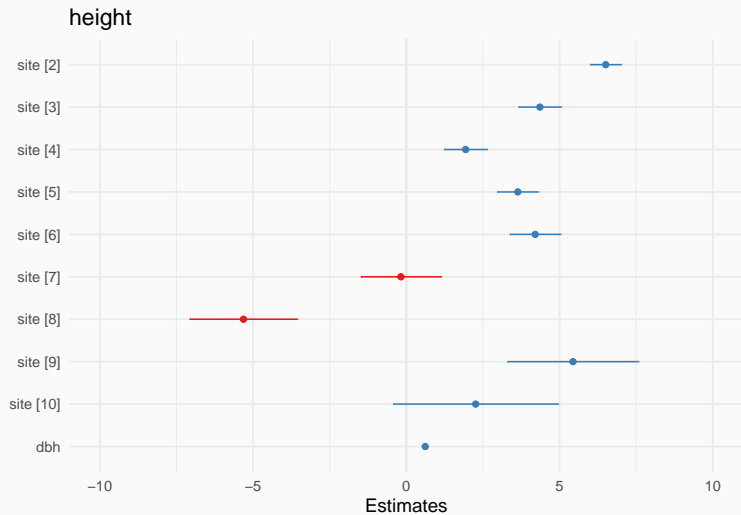


`$dbh`



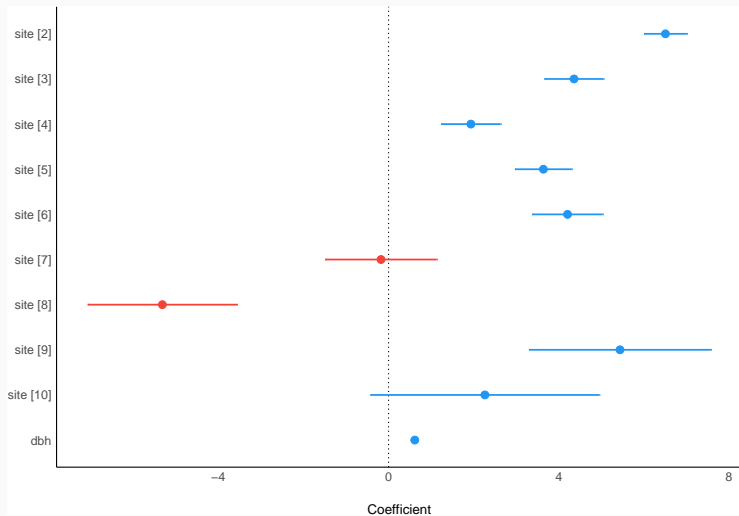
Plot model (sjPlot)

```
plot_model(m4, type = "est")
```

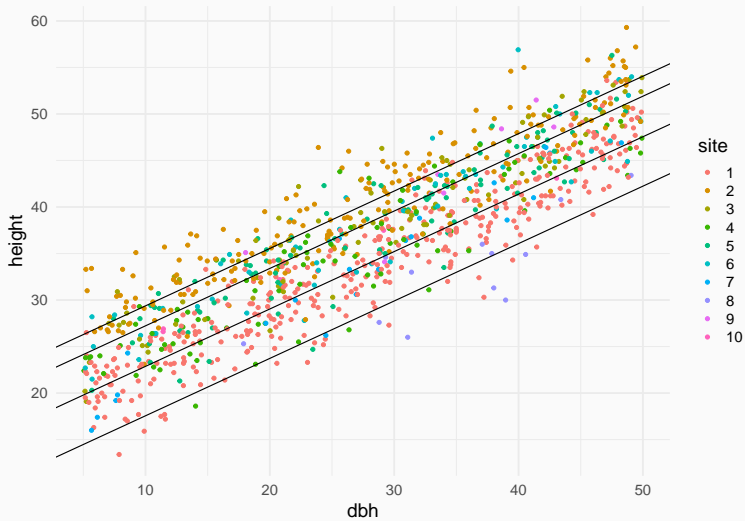


Plot model (see)

```
plot(parameters(m4))
```



We have fitted model w/ many intercepts and single slope



Slope is the same for all sites

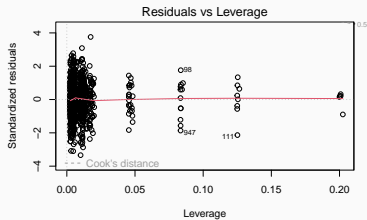
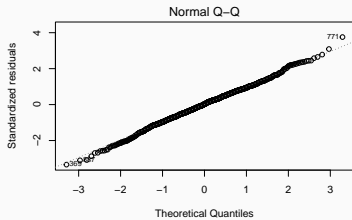
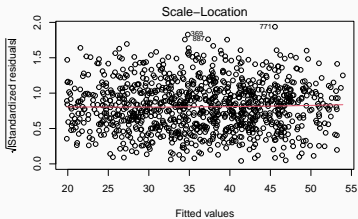
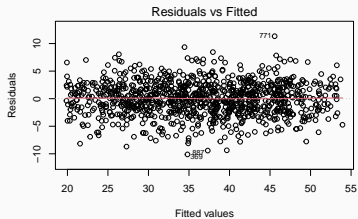
```
estimate_slopes(m4)
```

Estimated Marginal Effects

Coefficient	SE	95% CI	t(989)	p
0.62	7.57e-03	[0.60, 0.63]	81.47	< .001

Marginal effects estimated for dbh

Model checking: residuals

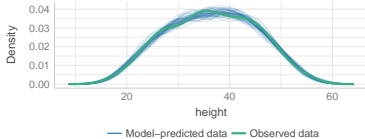


Model checking: residuals

check_model(m4)

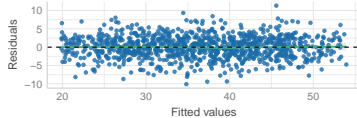
Posterior Predictive Check

Model-predicted lines should resemble observed data line



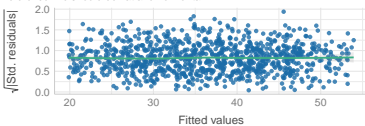
Linearity

Reference line should be flat and horizontal



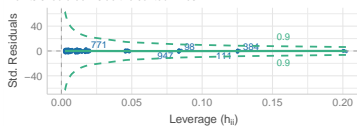
Homogeneity of Variance

Reference line should be flat and horizontal



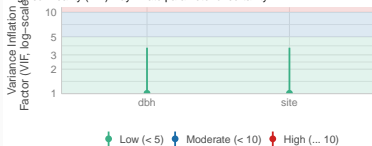
Influential Observations

Points should be inside the contour lines



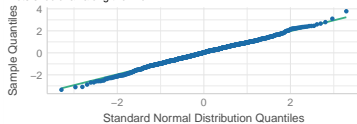
Collinearity

High collinearity (VIF) may inflate parameter uncertainty



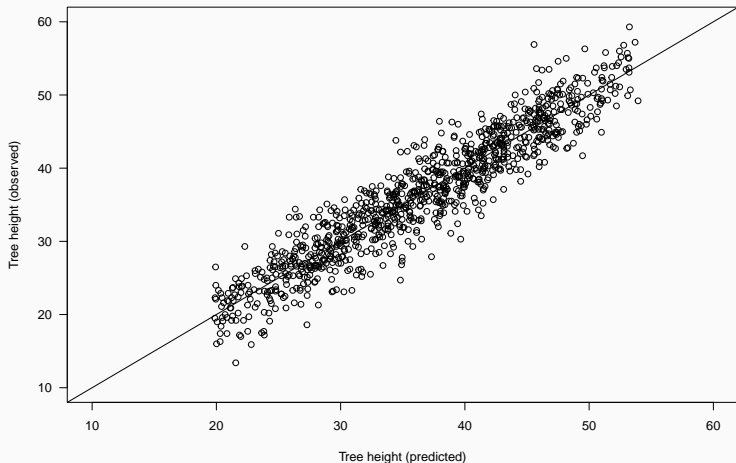
Normality of Residuals

Dots should fall along the line



How good is this model? Calibration plot

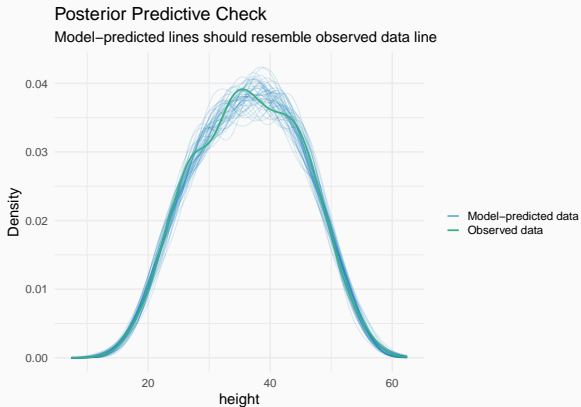
```
trees$height.pred <- fitted(m4)
plot(trees$height.pred, trees$height, xlab = "Tree height (predicted)",
      abline(a = 0, b = 1))
```



Posterior predictive checking

Simulating response data from fitted model (y_{rep})
and comparing with observed response (y)

```
performance::check_predictions(m4)
```



Using model for prediction

Expected height of 10-cm diameter tree in each site?

```
trees.10cm <- data.frame(site = as.factor(1:10),  
                          dbh = 10)
```

```
trees.10cm
```

	site	dbh
1	1	10
2	2	10
3	3	10
4	4	10
5	5	10
6	6	10
7	7	10
8	8	10
9	9	10
10	10	10

Using model for prediction

Confidence interval

```
predict(m4, newdata = trees.10cm, interval = "confidence")
```

	fit	lwr	upr
1	22.86979	22.46878	23.27079
2	29.37409	28.89388	29.85430
3	27.22724	26.54160	27.91289
4	24.80444	24.13410	25.47477
5	26.50722	25.84952	27.16492
6	27.07430	26.25490	27.89370
7	22.69359	21.39601	23.99117
8	17.55714	15.79282	19.32146
9	28.30683	26.16606	30.44761
10	25.13312	22.45540	27.81085

Using model for prediction

Prediction interval (accounting for residual variance)

```
predict(m4, newdata = trees.10cm, interval = "prediction")
```

	fit	lwr	upr
1	22.86979	16.88478	28.85480
2	29.37409	23.38325	35.36493
3	27.22724	21.21645	33.23804
4	24.80444	18.79537	30.81350
5	26.50722	20.49955	32.51489
6	27.07430	21.04678	33.10181
7	22.69359	16.58268	28.80451
8	17.55714	11.33039	23.78388
9	28.30683	21.96314	34.65053
10	25.13312	18.58868	31.67757

Using model for prediction

Prediction interval (99%)

```
predict(m4, newdata = trees.10cm, interval = "prediction",  
        level = 0.99)
```

	fit	lwr	upr
1	22.86979	14.998587	30.74098
2	29.37409	21.495225	37.25295
3	27.22724	19.322133	35.13235
4	24.80444	16.901598	32.70727
5	26.50722	18.606216	34.40822
6	27.07430	19.147195	35.00140
7	22.69359	14.656813	30.73037
8	17.55714	9.368019	25.74626
9	28.30683	19.963913	36.64976
10	25.13312	16.526183	33.74007

Q: Does allometric relationship
between Height and Diameter
vary among sites?

Model with interactions

Call:

```
lm(formula = height ~ site * dbh, data = trees)
```

Residuals:

	Min	1Q	Median	3Q	Max
	-10.1017	-1.9839	0.0645	2.0486	11.1789

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	16.359437	0.360054	45.436	< 2e-16 ***
site2	7.684781	0.609657	12.605	< 2e-16 ***
site3	4.518568	0.867008	5.212	2.28e-07 ***
site4	2.769336	0.813259	3.405	0.000688 ***
site5	3.917607	0.870983	4.498	7.68e-06 ***
site6	4.155161	1.009379	4.117	4.17e-05 ***
site7	-2.306799	1.551303	-1.487	0.137334
site8	-2.616095	4.090671	-0.640	0.522630
site9	2.621560	5.073794	0.517	0.605492
site10	4.662340	2.991072	1.559	0.119378
dbh	0.629299	0.011722	53.685	< 2e-16 ***
site2:dbh	-0.042784	0.020033	-2.136	0.032950 *
site3:dbh	-0.006031	0.027640	-0.218	0.827312
site4:dbh	-0.031633	0.028225	-1.121	0.262677
site5:dbh	-0.010173	0.027887	-0.365	0.715334
site6:dbh	0.001337	0.032109	0.042	0.966797
site7:dbh	0.079728	0.052056	1.532	0.125951
site8:dbh	-0.079027	0.113386	-0.697	0.485984
site9:dbh	0.081035	0.146649	0.553	0.580679
site10:dbh	-0.101107	0.114520	-0.883	0.377522

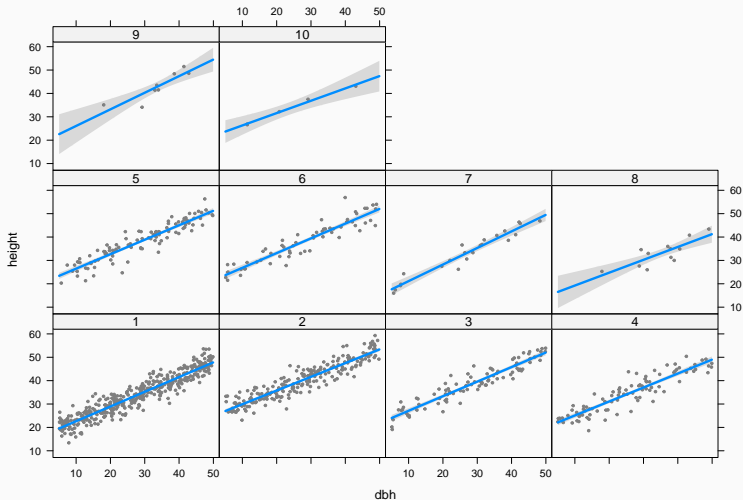
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Residual standard error: 3.041 on 980 degrees of freedom

Multiple R-squared: 0.8847 Adjusted R-squared: 0.8825

Does slope vary among sites?

```
visreg(m5, xvar = "dbh", by = "site")
```



Examining fitted model with {modelStudio}

```
library("modelStudio")  
m5.explain <- DALEX::explain(m5, data = trees, y = trees$height)  
modelStudio(m5.explain)
```

- [paperplanes](#): How does flight distance differ with age, gender or paper type?

- [paperplanes](#): How does flight distance differ with age, gender or paper type?
- [mammal sleep](#): Are sleep patterns related to diet?

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- [Penguins data](#): Body mass ~ Flipper length, Bill length ~ Bill depth, differences across sites...
- [racing pigeons](#): is speed related to sex?

Variable and model selection

Francisco Rodríguez-Sánchez

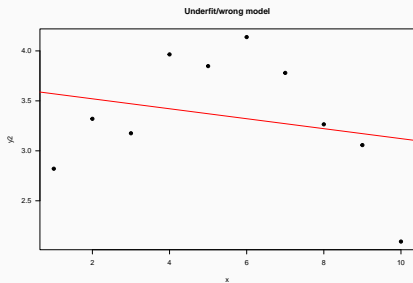
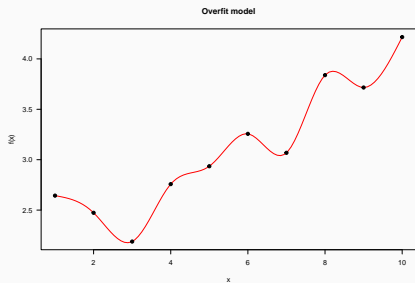
<https://frodriguezsanchez.net>

- On one hand, we want to **maximise fit**.

Overfitting and balanced model complexity

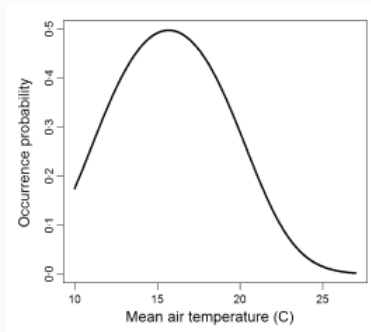
- On one hand, we want to **maximise fit**.
- On the other hand, we want to **avoid overfitting** and overly complex models.

Overfitting and balanced model complexity

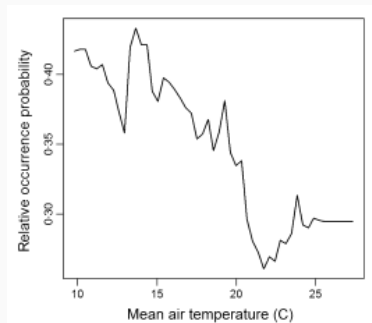


Overfitting and balanced model complexity

GLMM



Random forests



Wenger & Olden (2012)

Overfitted models will work badly on new data



- Cross-validation (k-fold, leave one out...)

- Cross-validation (k-fold, leave one out...)
- Information Criteria:

- Cross-validation (k-fold, leave one out...)
- Information Criteria:
 - AIC

- Cross-validation (k-fold, leave one out...)
- Information Criteria:
 - AIC
 - BIC

- Cross-validation (k-fold, leave one out...)
- Information Criteria:
 - AIC
 - BIC
 - DIC

- Cross-validation (k-fold, leave one out...)
- Information Criteria:
 - AIC
 - BIC
 - DIC
 - WAIC...

- Cross-validation (k-fold, leave one out...)
- **Information Criteria:**
 - AIC
 - BIC
 - DIC
 - WAIC...
- All these methods have flaws!

$$AIC = -2 * \text{LogLikelihood} + 2K$$

- First term: **model fit**

$$AIC = -2 * \text{LogLikelihood} + 2K$$

- First term: **model fit**
- **K = number of parameters** (penalisation for model complexity)

AIC (Akaike Information Criteria)

$$AIC = -2 * \text{LogLikelihood} + 2K$$

- First term: **model fit**
- **K = number of parameters** (penalisation for model complexity)
- Lower is better

AIC (Akaike Information Criteria)

$$AIC = -2 * \text{LogLikelihood} + 2K$$

- First term: **model fit**
- **K = number of parameters** (penalisation for model complexity)
- Lower is better
- AIC biased towards complex models.

$$AIC = -2 * \text{LogLikelihood} + 2K$$

- First term: **model fit**
- **K = number of parameters** (penalisation for model complexity)
- Lower is better
- AIC biased towards complex models.
- AICc recommended with 'small' sample sizes ($n/p < 40$). But see [Richards 2005](#)

- No information criteria is panacea: all have problems.

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- No information criteria is panacea: all have problems.
- They estimate *average* out-of-sample prediction error. But errors can differ substantially within dataset.
- Sometimes better models rank poorly (e.g. see [Gelman et al. 2013](#)). Combine with **thorough model checks**.

So which variables should enter
my model?

Choosing predictors

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Choosing predictors

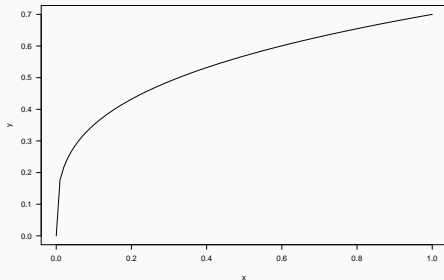
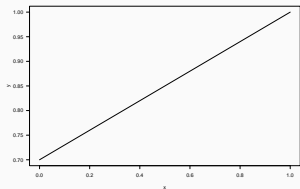
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 - Or combine 2 or more in a synthetic variable (e.g. water deficit ~ Temp + Precip).
 - Many methods available, e.g. sequential, ridge regression...
 - Measurement error can seriously complicate things (Biggs et al 2009; Freckleton 2011)
- For predictors with large effects, **consider interactions**.

Think about the shape of relationships

$$y \sim x + z$$

Really? Not everything has to be linear! Actually, it often is not.

Think about shape of relationship.



Removing predictors

Stepwise regression has many problems

- Whittingham et al. (2006) Why do we still use stepwise modelling in ecology and behaviour? *J. Animal Ecology*.

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- Mundry & Nunn (2009) Stepwise Model Fitting and Statistical Inference: Turning Noise into Signal Pollution. *Am Nat*.
- This includes **stepAIC** (e.g. Dahlgren 2010; Burnham et al 2011; Hegyi & Garamszegi 2011).

- Testing bivariate relationships before building multivariable model

Heinze & Dunkler 2016

- Testing bivariate relationships before building multivariable model
- Removing non-significant predictors

Heinze & Dunkler 2016

- Always keep **'core' predictors** (based on previous knowledge)

Heinze et al 2018

Removing predictors?

- Always **keep 'core' predictors** (based on previous knowledge)
- If ratio sample size/number of predictors is low (<10 EPP), avoid variable selection (too unstable)

Heinze et al 2018

Removing predictors?

- Always **keep 'core' predictors** (based on previous knowledge)
- If ratio sample size/number of predictors is low (<10 EPP), avoid variable selection (too unstable)
- If performing variable selection, always **assess stability** (bootstrap, etc)

Heinze et al 2018

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4. Always check fitted models thoroughly
5. Always report effect sizes

Model comparison

Francisco Rodríguez-Sánchez

<https://frodriguezsanchez.net>

Trees dataset

```
trees <- read.csv("data/trees.csv")  
head(trees)
```

	site	dbh	height	sex	dead
1	4	29.68	36.1	male	0
2	5	33.29	42.3	male	0
3	2	28.03	41.9	female	0
4	5	39.86	46.5	female	0
5	1	47.94	43.9	female	0
6	1	10.82	26.2	male	0

Four models

```
m1 <- lm(height ~ dbh, data = trees)
```

```
m2 <- lm(height ~ sex, data = trees)
```

```
m3 <- lm(height ~ site, data = trees)
```

```
m4 <- lm(height ~ site*dbh, data = trees)
```

Compare model performance

```
library("performance")  
compare_performance(m1, m2, m3, m4)
```

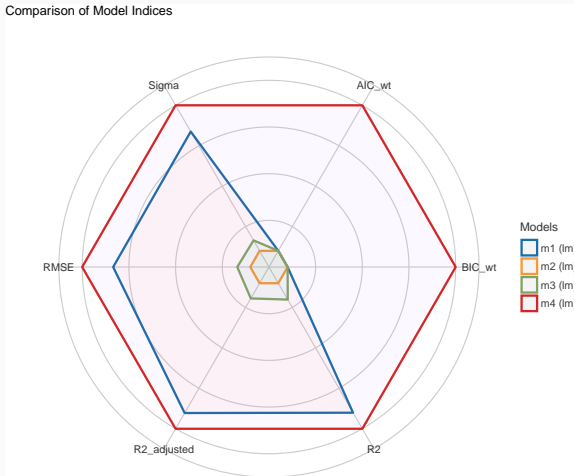
```
# Comparison of Model Performance Indices
```

Name	Model	AIC	AIC weights	BIC	BIC weights	R2	R2 (adj.)
m1	lm	5660.250	8.39e-126	5674.973	1.28e-106	0.787	0.787
m2	lm	7206.145	0.00e+00	7220.868	0.00e+00	0.002	0.001
m3	lm	7117.264	0.00e+00	7171.250	0.00e+00	0.102	0.093
m4	lm	5084.253	1.00	5187.316	1.00	0.885	0.882

Compare model performance

```
library("see")  
plot(compare_performance(m1, m2, m3, m4))
```

Comparison of Model Indices



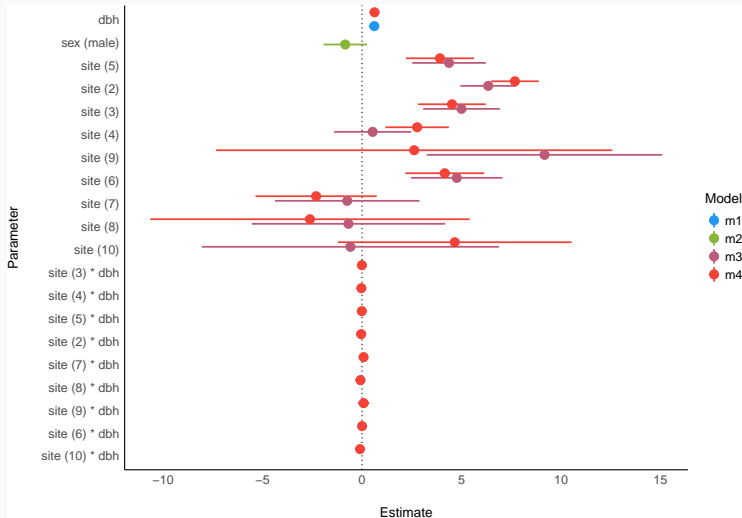
Compare parameters

```
library("parameters")
compare_parameters(m1, m2, m3, m4)
```

Parameter	m1	m2	m3	m4
(Intercept)	19.34 (18.73, 19.95)	36.93 (36.15, 37.71)	33.84 (33.00, 34.68)	16.36 (15.65, 17.07)
dbh	0.62 (0.60, 0.64)			0.63 (0.61, 0.65)
sex (male)	-0.84 (-1.94, 0.26)			
site (5)		4.37 (2.52, 6.22)		3.92 (2.21, 5.63)
site (2)		6.34 (4.94, 7.74)		7.68 (6.49, 8.88)
site (3)		5.00 (3.07, 6.93)		4.52 (2.82, 6.22)
site (4)		0.53 (-1.40, 2.47)		2.77 (1.17, 4.37)
site (9)		9.17 (3.25, 15.09)		2.62 (-7.34, 12.58)
site (6)		4.76 (2.46, 7.06)		4.16 (2.17, 6.14)
site (7)		-0.74 (-4.37, 2.89)		-2.31 (-5.35, 0.74)
site (8)		-0.68 (-5.54, 4.17)		-2.62 (-10.64, 5.41)
site (10)		-0.58 (-8.04, 6.88)		4.66 (-1.21, 10.53)
site (3) * dbh			-6.03e-03 (-0.06, 0.05)	
site (4) * dbh			-0.03 (-0.09, 0.02)	
site (5) * dbh			-0.01 (-0.06, 0.04)	
site (2) * dbh			-0.04 (-0.08, 0.00)	
site (7) * dbh			0.08 (-0.02, 0.18)	
site (8) * dbh			-0.08 (-0.30, 0.14)	
site (9) * dbh			0.08 (-0.21, 0.37)	
site (6) * dbh			1.34e-03 (-0.06, 0.06)	
site (10) * dbh			-0.10 (-0.33, 0.12)	
Observations	1000	1000	1000	1000

Compare parameters

```
library("parameters")  
plot(compare_parameters(m1, m2, m3, m4))
```



Generalised Linear Models

Logistic regression

Francisco Rodríguez-Sánchez

<https://frodriguezsanchez.net>

Q: Survival of passengers on the Titanic ~ Class

Read `titanic_long.csv` dataset and fit linear model (survival ~ class).

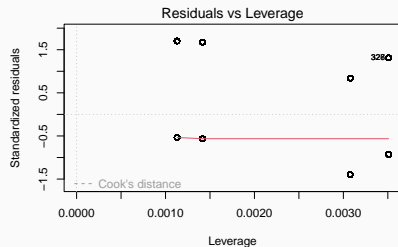
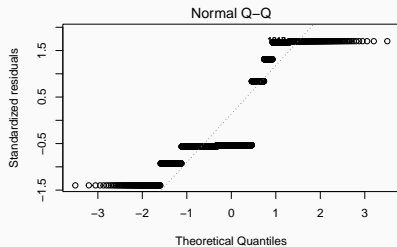
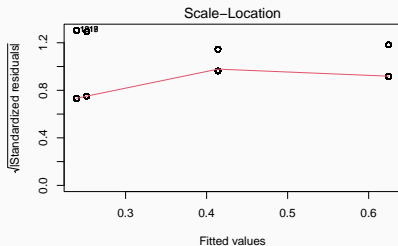
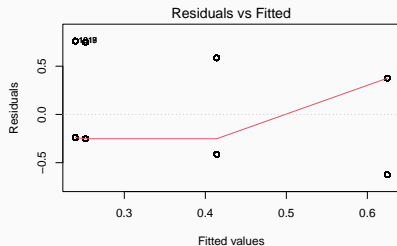
```
class age sex survived
1 first adult male      1
2 first adult male      1
3 first adult male      1
4 first adult male      1
5 first adult male      1
6 first adult male      1
```

Quiz: Did passenger class influence survival?

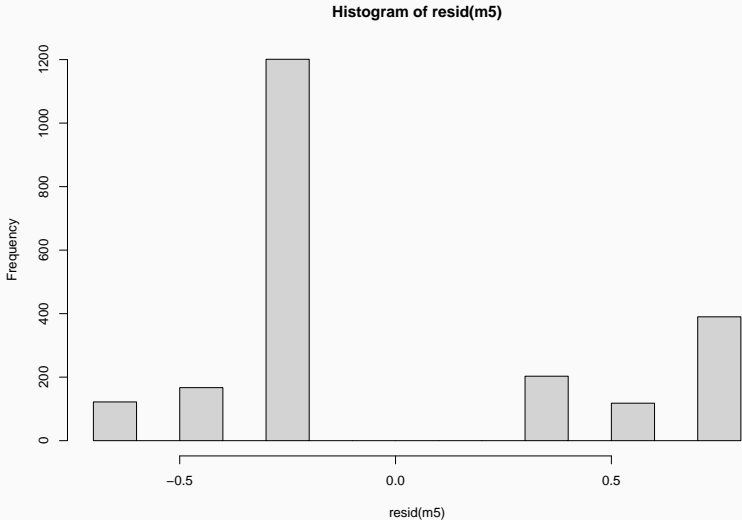
<https://pollev.com/franciscorod726>

Let's check linear model:

```
m5 <- lm(survived ~ class, data = titanic)
```



Weird residuals!



What if your residuals are clearly non-normal
or variance not constant (heteroscedasticity)?

Binary variables (0/1)

Counts (0, 1, 2, 3, ...)

Categories (“small”, “medium”, “large”...)

Generalised Linear Models to the rescue!

1. Response variable - distribution family

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- Bernoulli - Binomial

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- Bernoulli - Binomial
- Poisson

1. Response variable - distribution family

- Bernoulli - Binomial
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- Gamma

1. Response variable - distribution family

- Bernoulli - Binomial
- Poisson
- Gamma
- etc

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- etc

2. Predictors (continuous or categorical)

1. Response variable - distribution family

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- etc

2. Predictors (continuous or categorical)

3. Link function

1. Response variable - distribution family

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- Gaussian: identity

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- Gaussian: identity
- Binomial: logit, probit

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- Poisson: log..

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- Bernoulli - Binomial
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- Gamma
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- Gaussian: identity
- Binomial: logit, probit
- Poisson: log..
- See **family**.

The modelling process

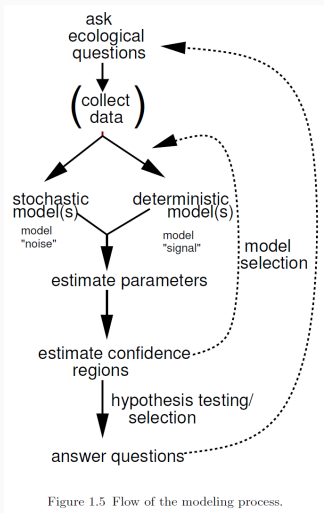


Figure 1.5 Flow of the modeling process.

Bernoulli - Binomial distribution (Logistic regression)

Response variable: **Yes/No** (e.g. survival, sex, presence/absence)

Canonical link function: **logit** (*log odds*), but others possible (see **family**)

$$\text{logit}(p) = \log\left(\frac{p}{1-p}\right)$$

Then

$$\text{logit}(P(\text{alive})) = a + bx$$

$$P(\text{alive}) = \text{invlogit}(a + bx) = \frac{e^{a+bx}}{1 + e^{a+bx}}$$

Where is the variance?

In a Gaussian GLM

$$y \sim \text{Normal}(\mu, \sigma)$$

In a Binomial GLM

$$y \sim \text{Binomial}(n, p)$$

n = number of trials

p = probability of success

$$\text{Var}(y) = np(1 - p)$$

(maximum variance when **p** around 0.5)

Back to survival of Titanic passengers

How many survived in each class?

```
table(titanic$class, titanic$survived)
```

	0	1
crew	673	212
first	122	203
second	167	118
third	528	178

How many survived in each class? (*dplyr*)

```
titanic %>%  
  group_by(class, survived) %>%  
  summarise(count = n())
```

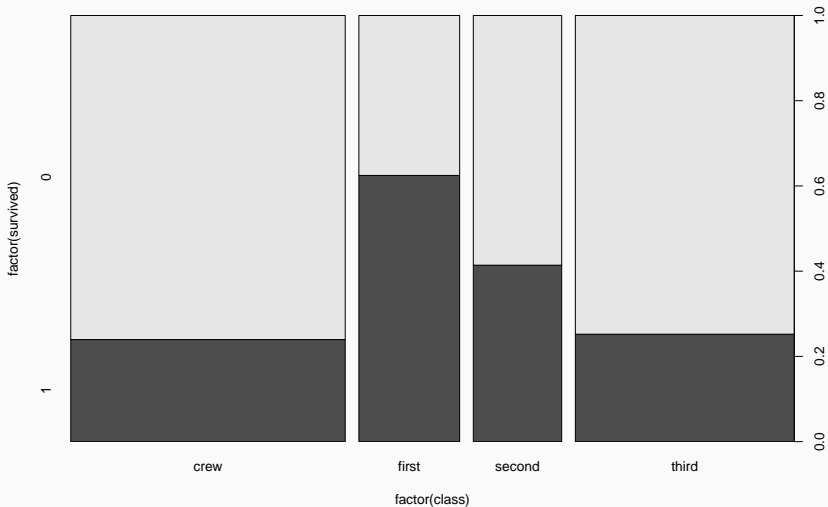
```
# A tibble: 8 x 3
```

```
# Groups:   class [4]
```

	class	survived	count
	<chr>	<int>	<int>
1	crew	0	673
2	crew	1	212
3	first	0	122
4	first	1	203
5	second	0	167
6	second	1	118
7	third	0	528
8	third	1	178

Data visualisation (mosaic plot)

```
plot(factor(survived) ~ factor(class), data = titanic)
```



Mosaic plots (ggplot2)

```
ggplot(titanic) +  
  geom_mosaic(aes(x = product(survived, class))) +  
  labs(x = "", y = "Survived")
```



```
tit.glm <- glm(survived ~ class,  
              data = titanic,  
              family = binomial)
```

which corresponds to

$$\text{logit}(P(\text{survival})_i) = a + b \cdot \text{class}_i$$

$$\text{logit}(P(\text{survival})_i) = a + b_{\text{first}} + c_{\text{second}} + d_{\text{third}}$$

Interpreting binomial GLM

```
tit.glm <- glm(survived ~ class, data = titanic, family = binomial)
```

Call:

```
glm(formula = survived ~ class, family = binomial, data = titanic)
```

Deviance Residuals:

Min	1Q	Median	3Q	Max
-1.3999	-0.7623	-0.7401	0.9702	1.6906

Coefficients:

	Estimate	Std. Error	z value	Pr(> z)
(Intercept)	-1.15516	0.07876	-14.667	< 2e-16 ***
classfirst	1.66434	0.13902	11.972	< 2e-16 ***
classecond	0.80785	0.14375	5.620	1.91e-08 ***
classtthird	0.06785	0.11711	0.579	0.562

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

(Dispersion parameter for binomial family taken to be 1)

Null deviance: 2769.5 on 2200 degrees of freedom
Residual deviance: 2588.6 on 2197 degrees of freedom
AIC: 2596.6

Number of Fisher Scoring iterations: 4

Binomial GLM estimates are in `logit` scale!

We need to **back-transform** (apply *inverse logit*):

- Manually: `plogis`

Binomial GLM estimates are in `logit` scale!

We need to **back-transform** (apply *inverse logit*):

- Manually: `plogis`
- Automatically: `effects`, `modelbased`, etc.

Interpreting logistic regression output (effects pkg)

```
library("effects")  
allEffects(tit.glm)
```

```
model: survived ~ class
```

```
class effect
```

```
class
```

	crew	first	second	third
	0.2395480	0.6246154	0.4140351	0.2521246

Interpreting logistic regression output (effects pkg)

Including confidence intervals:

```
summary(allEffects(tit.glm))
```

```
model: survived ~ class
```

```
class effect
```

```
class
```

```
      crew      first      second      third  
0.2395480 0.6246154 0.4140351 0.2521246
```

```
Lower 95 Percent Confidence Limits
```

```
class
```

```
      crew      first      second      third  
0.2125668 0.5706887 0.3582390 0.2214588
```

```
Upper 95 Percent Confidence Limits
```

```
class
```

```
      crew      first      second      third  
0.2687850 0.6756185 0.4721282 0.2854798
```


Interpreting logistic regression output (modelbased)

```
library("modelbased")  
estimate_means(tit.glm)
```

Estimated Marginal Means

class	Probability	SE	95% CI
first	0.62	0.03	[0.57, 0.68]
second	0.41	0.03	[0.36, 0.47]
third	0.25	0.02	[0.22, 0.29]
crew	0.24	0.01	[0.21, 0.27]

Marginal means estimated at class

Analysing differences among factor levels (class)

```
library("modelbased")  
estimate_contrasts(tit.glm)
```

Marginal Contrasts Analysis

Level1	Level2	Difference	95% CI	SE	df	z
first	crew	1.66	[1.30, 2.03]	0.14	Inf	11.97
first	second	0.86	[0.42, 1.29]	0.17	Inf	5.16
first	third	1.60	[1.22, 1.98]	0.14	Inf	11.11
second	crew	0.81	[0.43, 1.19]	0.14	Inf	5.62
second	third	0.74	[0.35, 1.13]	0.15	Inf	4.99
third	crew	0.07	[-0.24, 0.38]	0.12	Inf	0.58

Marginal contrasts estimated at class
p-value adjustment method: Holm (1979)

```
library("performance")  
r2(tit.glm)
```

```
# R2 for Logistic Regression  
Tjur's R2: 0.087
```

But there are caveats (e.g. see [here](#) and [here](#))

```
kable(xtable::xtable(tit.glm), digits = 2)
```

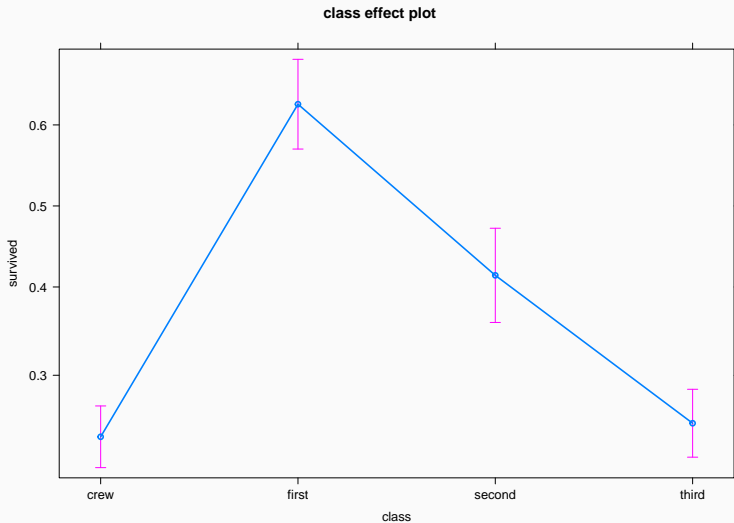
	Estimate	Std. Error	z value	Pr(> z)
(Intercept)	-1.16	0.08	-14.67	0.00
classfirst	1.66	0.14	11.97	0.00
classecond	0.81	0.14	5.62	0.00
classthir	0.07	0.12	0.58	0.56

Presenting model results

```
library("modelsummary")  
modelsummary(tit.glm)
```

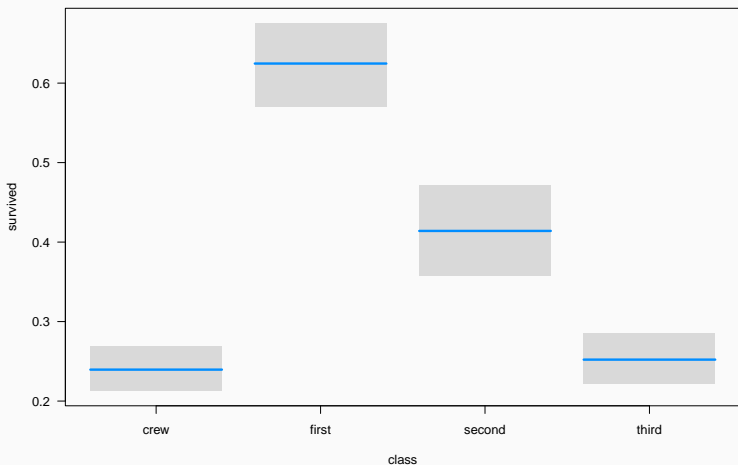
Visualising model: effects package

```
plot(allEffects(tit.glm))
```



Visualising model: visreg package

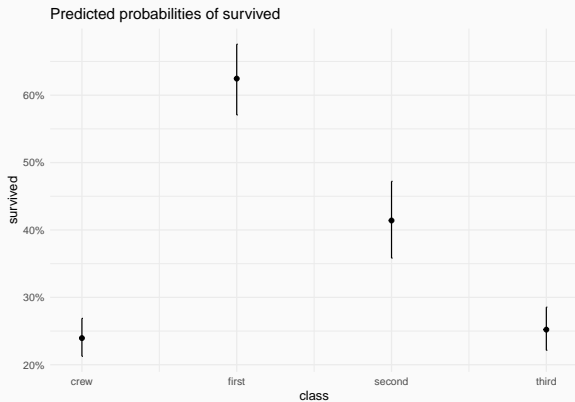
```
visreg(tit.glm, scale = "response", rug = FALSE)
```



Visualising model: sjPlot package

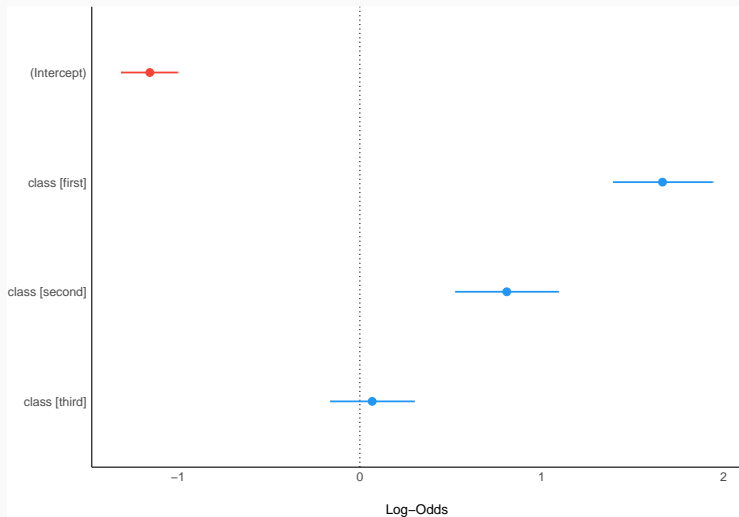
```
sjPlot::plot_model(tit.glm, type = "eff")
```

`$class`



Visualising model: see package

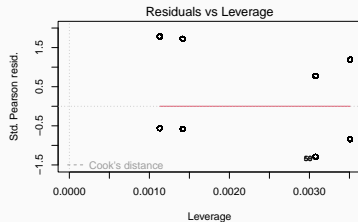
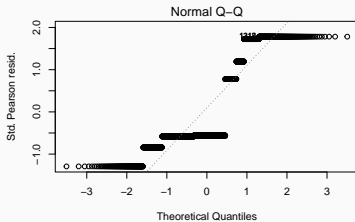
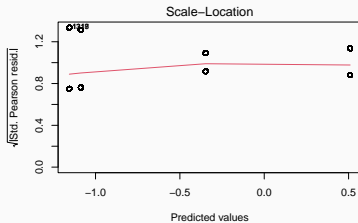
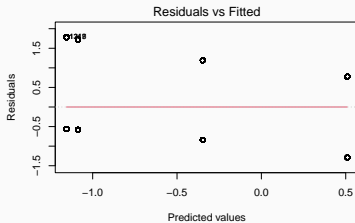
```
plot(parameters(tit.glm), show_intercept = TRUE)
```



Model checking

plot(model) not very useful with binomial GLM

```
plot(tit.glm)
```



```
null device
```

```
1
```

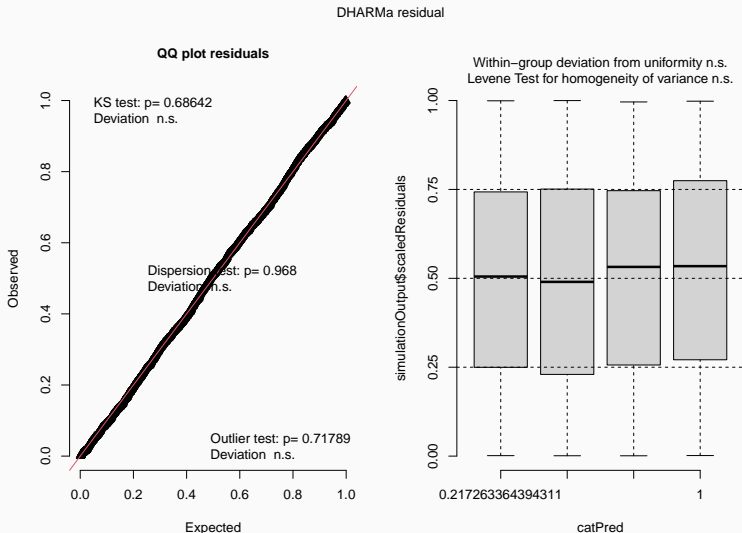
Binned residual plots for logistic regression

```
binned_residuals(tit.glm)
```

Ok: About 100% of the residuals are inside the error bounds.

Residual diagnostics with DHARMa

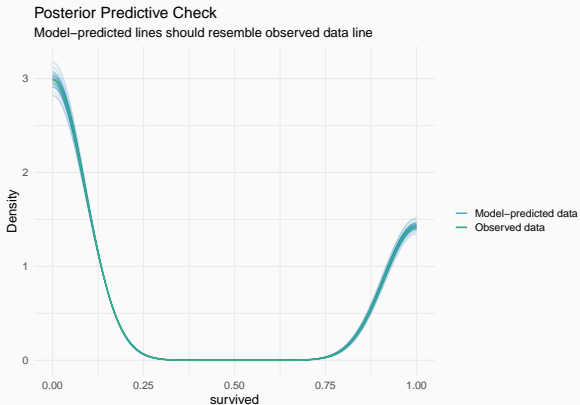
```
library("DHARMa")  
simulateResiduals(tit.glm, plot = TRUE)
```



Posterior predictive checking

Simulate data from fitted model (**yrep**) and compare with observed data (**y**)

```
check_predictions(tit.glm)
```

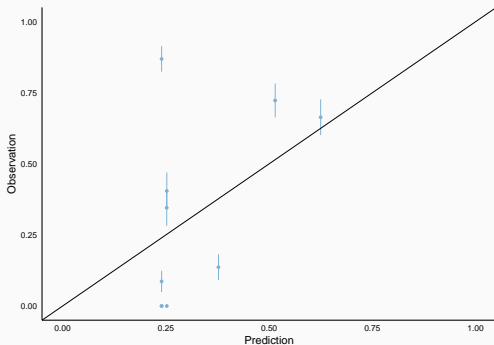


Calibration plot

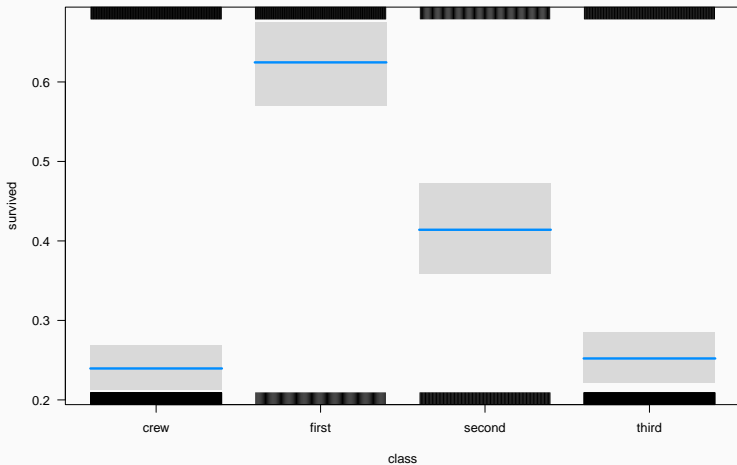
Compares predicted vs observed probabilities (grouped by quantiles)

```
library("predtools")
titanic$surv.pred <- predict(tit.glm, type = "response")
calibration_plot(data = titanic, obs = "survived", pred = "surv.pred",
                 x_lim = c(0,1), y_lim = c(0,1))
```

`$calibration_plot`



Passenger class was important, but lots of unexplained variation



The goal is not to test whether the model's assumptions are "true", because all models are false.

Rather, the goal is to assess exactly **how the model fails to describe the data**, as a path towards **model comprehension, revision, and improvement**.

Richard McElreath. *Statistical Rethinking*

1. Visualise data

Recapitulating

1. Visualise data
2. Fit model: `glm`. Don't forget to specify `family`!

Recapitulating

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3. Examine model: `summary`

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4. Back-transform parameters from *logit* into probability scale (e.g. `allEffects`)

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(e.g. `allEffects`)
5. Plot model: `plot(allEffects(model))`, `visreg`, `plot_model`,
calibration plot...

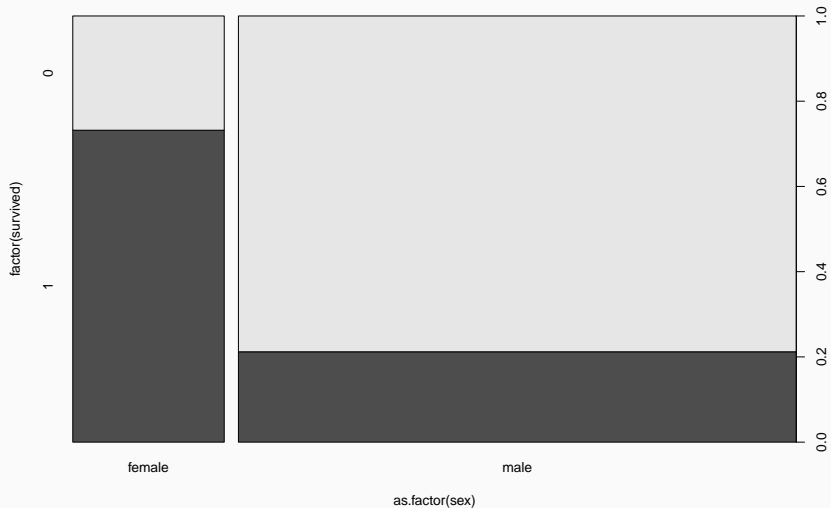
Recapitulating

1. Visualise data
2. Fit model: `glm`. Don't forget to specify `family`!
3. Examine model: `summary`
4. Back-transform parameters from *logit* into probability scale
(e.g. `allEffects`)
5. Plot model: `plot(allEffects(model))`, `visreg`, `plot_model`,
calibration plot...
6. Examine residuals: `DHARMA::simulateResiduals`.

Q: Did men have higher survival
than women?

<https://pollev.com/franciscorod726>

First, visualise data



Fit model

Call:

```
glm(formula = survived ~ sex, family = binomial, data = titanic)
```

Deviance Residuals:

Min	1Q	Median	3Q	Max
-1.6226	-0.6903	-0.6903	0.7901	1.7613

Coefficients:

	Estimate	Std. Error	z value	Pr(> z)
(Intercept)	1.0044	0.1041	9.645	<2e-16 ***
sexmale	-2.3172	0.1196	-19.376	<2e-16 ***

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

(Dispersion parameter for binomial family taken to be 1)

Null deviance: 2769.5 on 2200 degrees of freedom
Residual deviance: 2335.0 on 2199 degrees of freedom
AIC: 2339

Number of Fisher Scoring iterations: 4

Model interpretation

```
model: survived ~ sex
```

```
sex effect
```

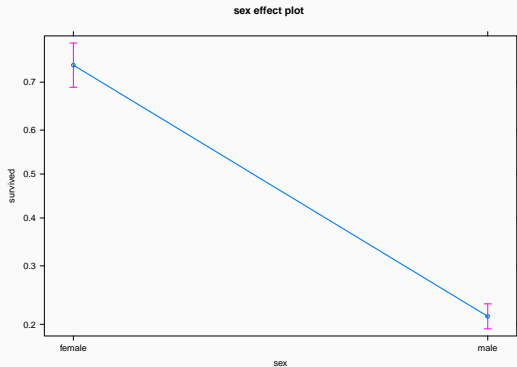
```
sex
```

```
female
```

```
male
```

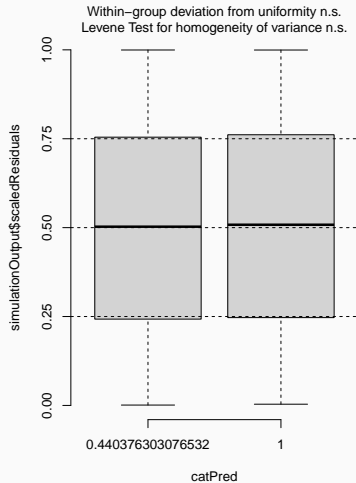
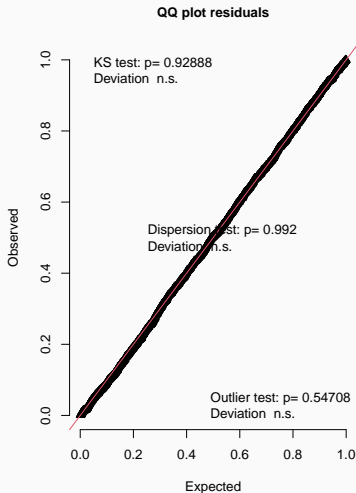
```
0.7319149
```

```
0.2120162
```



```
simulateResiduals(tit.sex, plot = TRUE)
```

DHARMA residual



Q: Did women have higher survival because they travelled more in first class?

Did women have higher survival because they travelled more in first class?



Let's look at the data

```
table(titanic$class, titanic$survived, titanic$sex)
```

```
, , = female
```

	0	1
crew	3	20
first	4	141
second	13	93
third	106	90

```
, , = male
```

	0	1
crew	670	192
first	118	62
second	154	25
third	422	88

<https://pollev.com/franciscorod726>

Fit additive model with both factors

Call:

```
glm(formula = survived ~ class + sex, family = binomial, data = titanic)
```

Deviance Residuals:

Min	1Q	Median	3Q	Max
-2.0915	-0.7149	-0.5012	0.7297	2.0673

Coefficients:

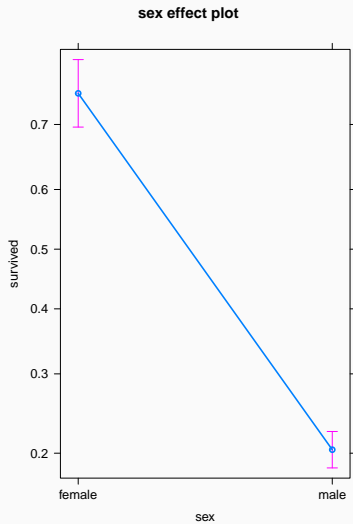
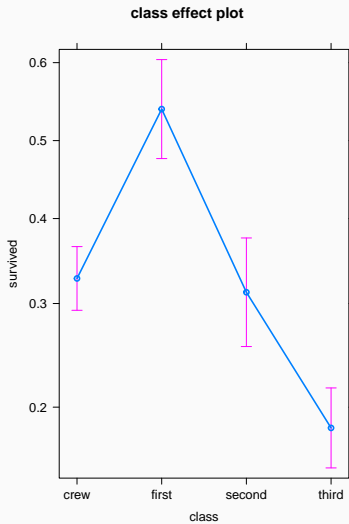
	Estimate	Std. Error	z value	Pr(> z)	
(Intercept)	1.18740	0.15747	7.541	4.68e-14	***
classfirst	0.88081	0.15697	5.611	2.01e-08	***
classecond	-0.07178	0.17093	-0.420	0.675	
classtthird	-0.77742	0.14231	-5.463	4.69e-08	***
sexmale	-2.42133	0.13909	-17.408	< 2e-16	***

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

(Dispersion parameter for binomial family taken to be 1)

Null deviance: 2769.5 on 2200 degrees of freedom
Residual deviance: 2228.9 on 2196 degrees of freedom

Plot additive model



Fit model with the interaction of both factors

Call:

```
glm(formula = survived ~ class * sex, family = binomial, data = titanic)
```

Deviance Residuals:

Min	1Q	Median	3Q	Max
-2.6797	-0.7099	-0.6155	0.5115	1.9842

Coefficients:

	Estimate	Std. Error	z value	Pr(> z)	
(Intercept)	1.89712	0.61914	3.064	0.00218	**
classfirst	1.66535	0.80026	2.081	0.03743	*
classecond	0.07053	0.68630	0.103	0.91815	
classthird	-2.06075	0.63551	-3.243	0.00118	**
sexmale	-3.14690	0.62453	-5.039	4.68e-07	***
classfirst:sexmale	-1.05911	0.81959	-1.292	0.19627	
classecond:sexmale	-0.63882	0.72402	-0.882	0.37760	
classthird:sexmale	1.74286	0.65139	2.676	0.00746	**

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

(Dispersion parameter for binomial family taken to be 1)

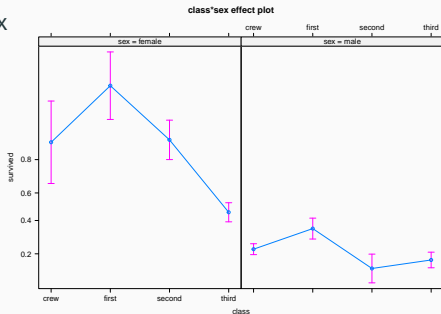
Women had higher survival than men, even within the same class

```
model: survived ~ class * sex
```

```
class*sex effect
```

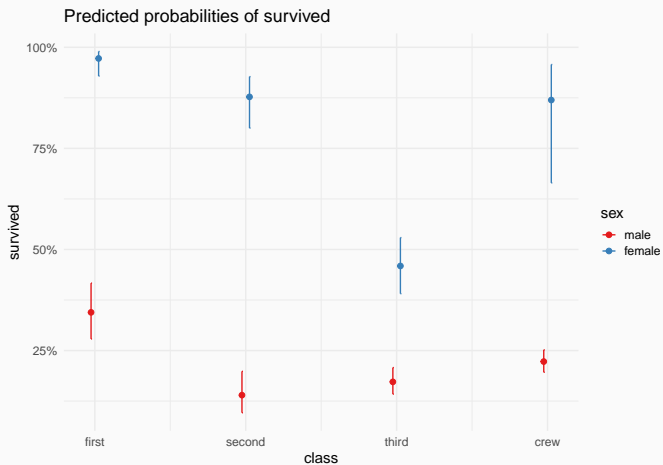
```
sex
```

class	female	male
crew	0.8695652	0.2227378
first	0.9724138	0.3444444
second	0.8773585	0.1396648
third	0.4591837	0.1725490



Visualising model (sjPlot)

```
plot_model(tit.sex.class.int, type = "int")
```



Comparing models

```
library("performance")
compare_performance(tit.sex.class.add, tit.sex.class.int)
```

```
# Comparison of Model Performance Indices
```

Name	Model	AIC	AIC weights	BIC	BIC weights	Tjur'
tit.sex.class.add	glm	2238.913	1.41e-13	2267.396	7.25e-10	0
tit.sex.class.int	glm	2179.733	1.000	2225.306	1.000	0

Comparing parameters

```
compare_parameters(tit.sex.class.add, tit.sex.class.int)
```

Parameter	tit.sex.class.add	tit.sex.class.int
(Intercept)	1.19 (0.88, 1.50)	1.90 (0.68, 3.11)
class (first)	0.88 (0.57, 1.19)	1.67 (0.10, 3.23)
class (second)	-0.07 (-0.41, 0.26)	0.07 (-1.27, 1.42)
class (third)	-0.78 (-1.06, -0.50)	-2.06 (-3.31, -0.82)
sex (male)	-2.42 (-2.69, -2.15)	-3.15 (-4.37, -1.92)
class (first) * sex (male)		-1.06 (-2.67, 0.55)
class (second) * sex (male)		-0.64 (-2.06, 0.78)
class (third) * sex (male)		1.74 (0.47, 3.02)
Observations	2201	2201

Is survival related to age?

Are age effects dependent on sex?

Logistic regression for proportion data

Read Titanic data in different format

Read `titanic_prop.csv` data.

	X	Class	Sex	Age	No	Yes
1	1	1st	Female	Adult	4	140
2	2	1st	Female	Child	0	1
3	3	1st	Male	Adult	118	57
4	4	1st	Male	Child	0	5
5	5	2nd	Female	Adult	13	80
6	6	2nd	Female	Child	0	13

These are the same data, but summarized (see `Freq` variable).

Use `cbind(n.success, n.failures)` as response

```
prop.glm <- glm(cbind(Yes, No) ~ Class, data = tit.prop, family = binomial)
```

Call:

```
glm(formula = cbind(Yes, No) ~ Class, family = binomial, data = tit.prop)
```

Deviance Residuals:

Min	1Q	Median	3Q	Max
-9.6404	-0.2915	1.5698	5.0366	10.1516

Coefficients:

	Estimate	Std. Error	z value	Pr(> z)	
(Intercept)	0.5092	0.1146	4.445	8.79e-06	***
Class2nd	-0.8565	0.1661	-5.157	2.51e-07	***
Class3rd	-1.5965	0.1436	-11.114	< 2e-16	***
ClassCrew	-1.6643	0.1390	-11.972	< 2e-16	***

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

(Dispersion parameter for binomial family taken to be 1)

Null deviance: 671.96 on 13 degrees of freedom

Effects

```
model: cbind(Yes, No) ~ Class
```

```
Class effect
```

```
Class
```

	1st	2nd	3rd	Crew
	0.6246154	0.4140351	0.2521246	0.2395480

Compare with former model based on binary data:

```
model: survived ~ class
```

```
class effect
```

```
class
```

	crew	first	second	third
	0.2395480	0.6246154	0.4140351	0.2521246

Logistic regression with continuous predictors

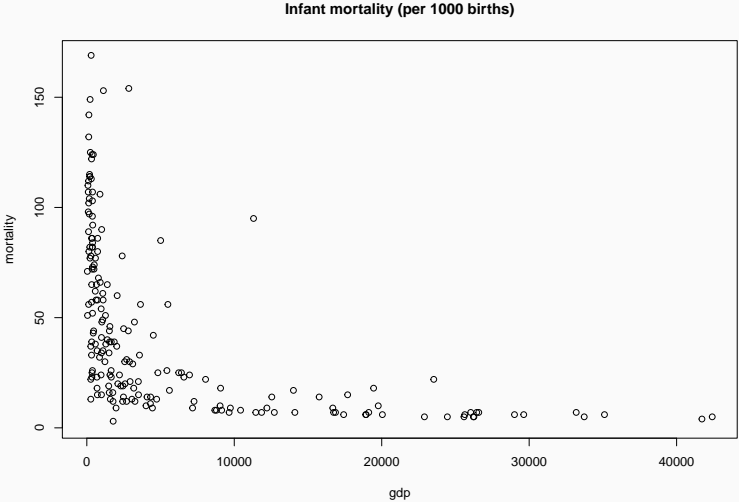
Example dataset: [GDP and infant mortality](#)

Read UN_GDP_infantmortality.csv.

country	mortality	gdp
Length:207	Min. : 2.00	Min. : 36
Class :character	1st Qu.: 12.00	1st Qu.: 442
Mode :character	Median : 30.00	Median : 1779
	Mean : 43.48	Mean : 6262
	3rd Qu.: 66.00	3rd Qu.: 7272
	Max. :169.00	Max. :42416
	NA's :6	NA's :10

Q: Is infant mortality related to GDP?

<https://pollev.com/franciscorod726>



Fit model

```
gdp.glm <- glm(cbind(mortality, 1000 - mortality) ~ gdp,  
              data = gdp, family = binomial)
```

Call:

```
glm(formula = cbind(mortality, 1000 - mortality) ~ gdp, family = binomial,  
    data = gdp)
```

Deviance Residuals:

Min	1Q	Median	3Q	Max
-9.2230	-3.5163	-0.5697	2.4284	13.5849

Coefficients:

	Estimate	Std. Error	z value	Pr(> z)
(Intercept)	-2.657e+00	1.311e-02	-202.76	<2e-16 ***
gdp	-1.279e-04	3.458e-06	-36.98	<2e-16 ***

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

(Dispersion parameter for binomial family taken to be 1)

Null deviance: 6430.2 on 192 degrees of freedom

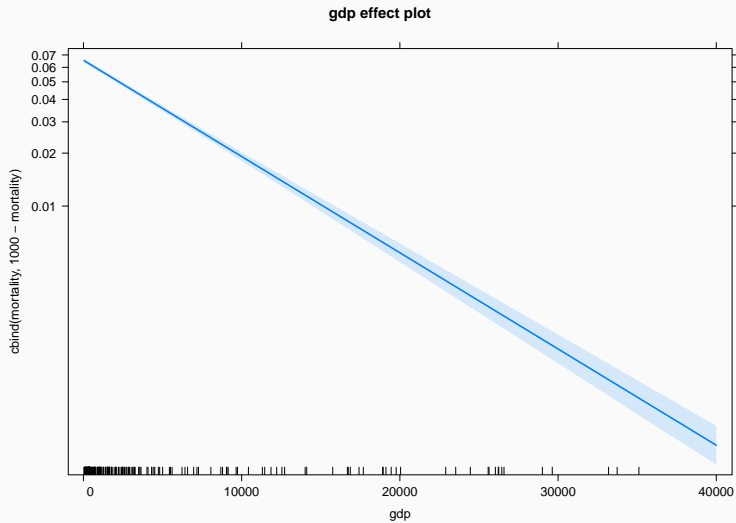
```
allEffects(gdp.glm)
```

```
model: cbind(mortality, 1000 - mortality) ~ gdp
```

```
gdp effect
```

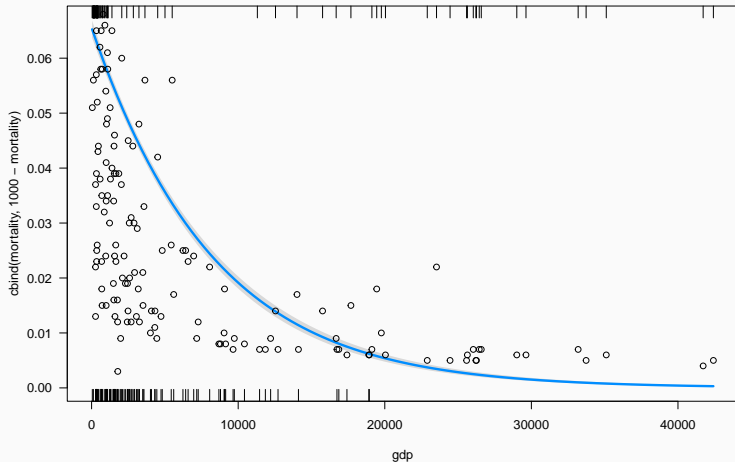
```
gdp
```

	40	10000	20000	30000	40000
	0.0652177296	0.0191438829	0.0054028095	0.0015096074	0.0004206154



Plot model using visreg:

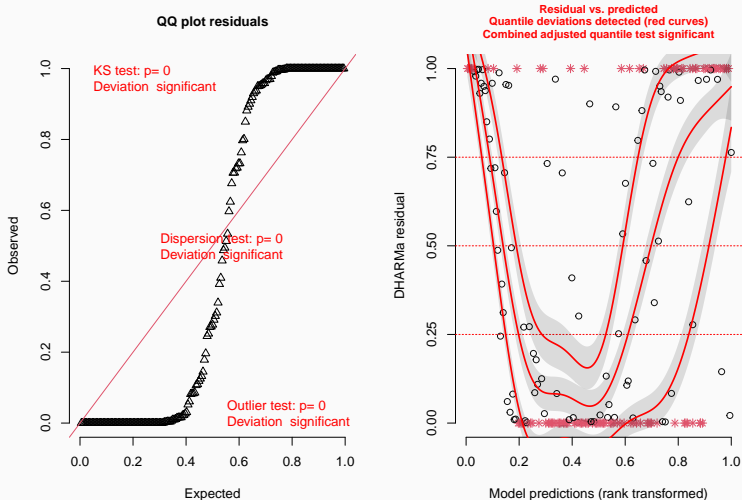
```
visreg(gdp.glm, scale = "response")  
points(mortality/1000 ~ gdp, data = gdp)
```



Residuals diagnostics with DHARMA

```
simulateResiduals(gdp.glm, plot = TRUE)
```

DHARMA residual



Overdispersion

Overdispersion:

more variation in the data than assumed by statistical model

$$\text{Var}(y) = np(1 - p)$$

Testing for overdispersion (DHARMA)

```
simres <- simulateResiduals(gdp.glm, refit = TRUE)  
testDispersion(simres, plot = FALSE)
```

DHARMA nonparametric dispersion test via mean deviance residuals
vs. simulated-refitted

```
data: simres  
dispersion = 21, p-value < 2.2e-16  
alternative hypothesis: two.sided
```

`quasibinomial` allows us to model overdispersed binomial data

Overdispersion in logistic regression with proportion data

```
gdp.overdisp <- glm(cbind(mortality, 1000 - mortality) ~ gdp,  
                    data = gdp, family = quasibinomial)
```

Call:

```
glm(formula = cbind(mortality, 1000 - mortality) ~ gdp, family = quasibinomial,  
     data = gdp)
```

Deviance Residuals:

Min	1Q	Median	3Q	Max
-9.2230	-3.5163	-0.5697	2.4284	13.5849

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	-2.657e+00	5.977e-02	-44.465	< 2e-16 ***
gdp	-1.279e-04	1.577e-05	-8.111	5.96e-14 ***

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

(Dispersion parameter for quasibinomial family taken to be 20.7947)

Null deviance: 6430.2 on 192 degrees of freedom

Mean estimates do not change after accounting for overdispersion

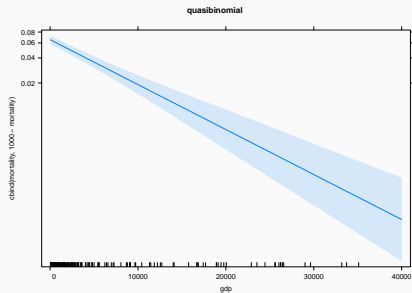
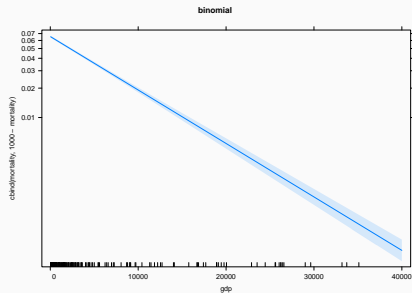
```
coef(gdp.overdisp)
```

```
(Intercept)          gdp  
-2.6574663734 -0.0001278976
```

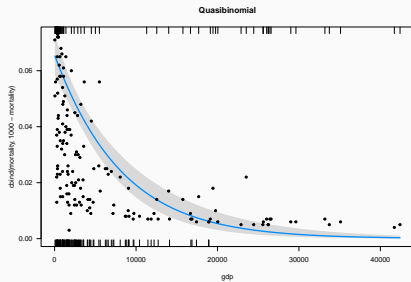
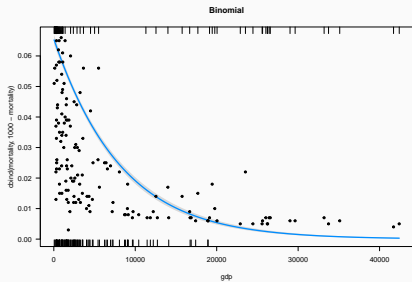
```
coef(gdp.glm)
```

```
(Intercept)          gdp  
-2.6574663734 -0.0001278976
```

But standard errors (uncertainty) do!



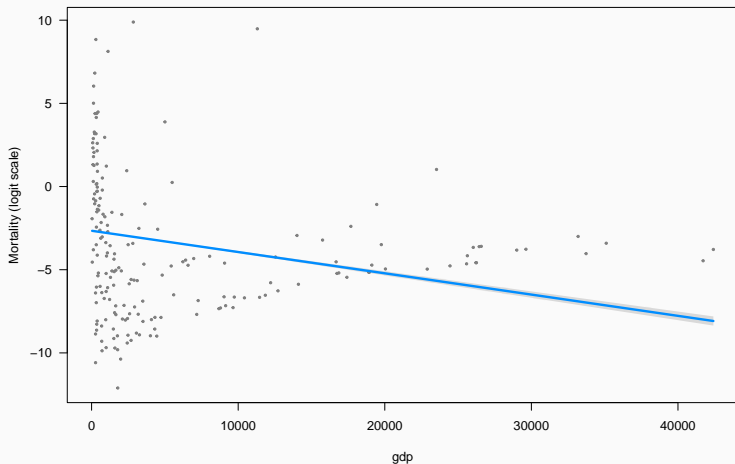
Plot model and data



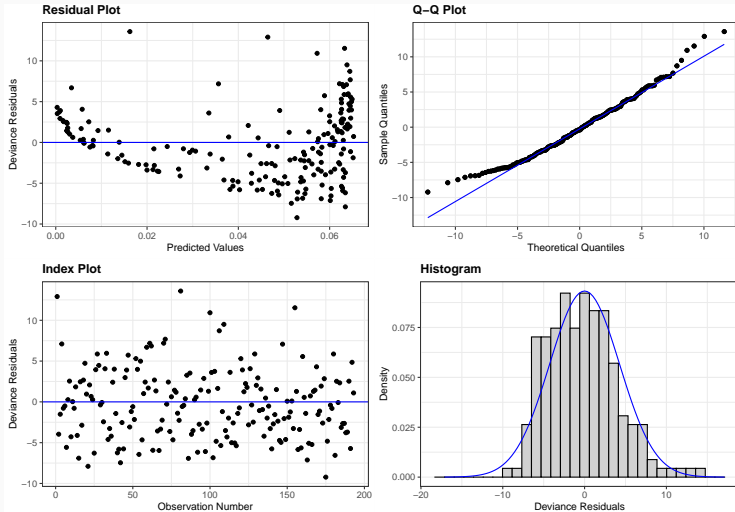
Think about the shape of
relationships

Think about the shape of relationships

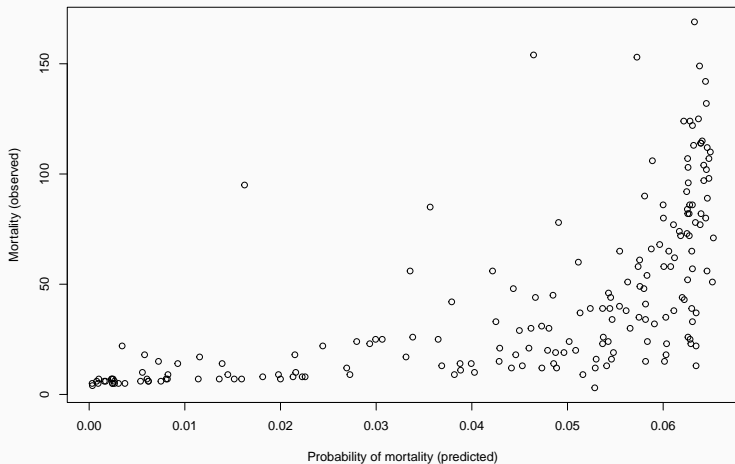
Not everything has to be linear...



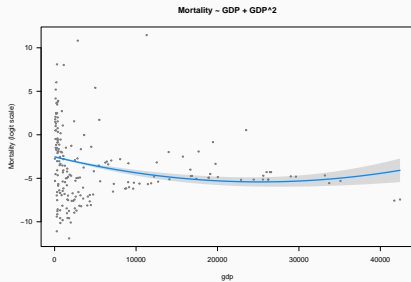
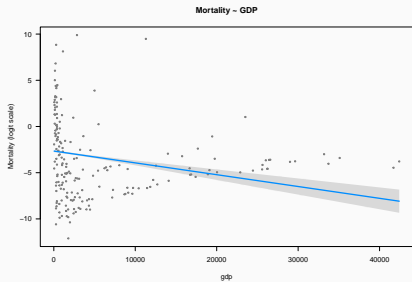
Residuals show non-linear pattern



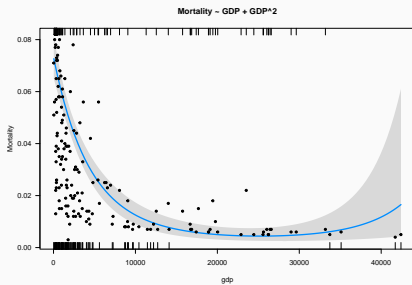
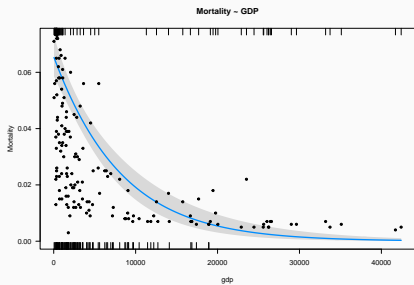
Calibration plot shows non-linear pattern



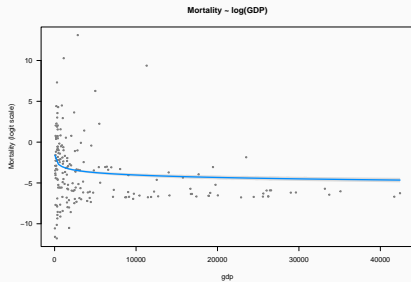
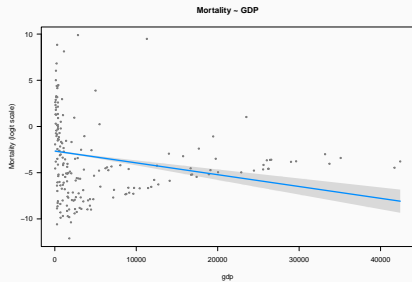
Trying polynomial predictor (GDP + GDP²)



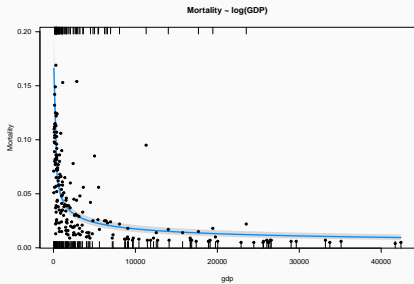
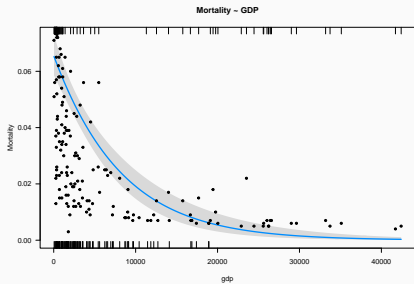
Think about the shape of relationships



Trying log(GDP)



Trying log(GDP)



- `seedset.csv`: Comparing seed set among plants (Data from [Harder et al. 2011](#))

- `seedset.csv`: Comparing seed set among plants (Data from [Harder et al. 2011](#))
- `moth.csv`: Probability of moth predation on trunk trees depending on morph (light/dark) and distance to Liverpool ([Bishop 1972](#))

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- `moth.csv`: Probability of moth predation on trunk trees depending on morph (light/dark) and distance to Liverpool ([Bishop 1972](#))
- `soccer.csv`: Probability of scoring penalty depending on goalkeeper's team being ahead, behind or tied ([Roskes et al 2011](#))

Probability of scoring penalty

Data on penalty shots

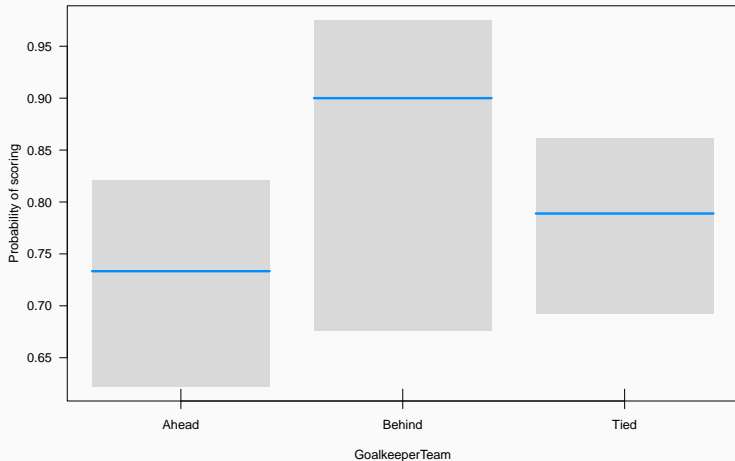
```
soccer <- read.csv("data/soccer.csv")  
soccer
```

	GoalkeeperTeam	Nshots	Scored
1	Behind	20	18
2	Tied	90	71
3	Ahead	75	55

Does probability of scoring penalty depends on match situation?

<https://pollev.com/franciscorod726>

Probability of scoring depending on match situation



Seed set among plants

Seed set among plants



Seed set among plants

```
# A tibble: 6 x 6
```

```
  species    plant pcmass fertilized seeds ovulecnt  
  <chr>      <dbl> <dbl>         <dbl> <dbl>    <dbl>  
1 ferruginea    2  0           70     52     330  
2 ferruginea    2  0.2         321    188     461  
3 ferruginea    2  0.485       351    278     435  
4 ferruginea    2  0.737       386    301     430  
5 ferruginea    2  1           367    342     419  
6 ferruginea    3  0           185     39     470
```

Questions:

<https://pollev.com/franciscorod726>

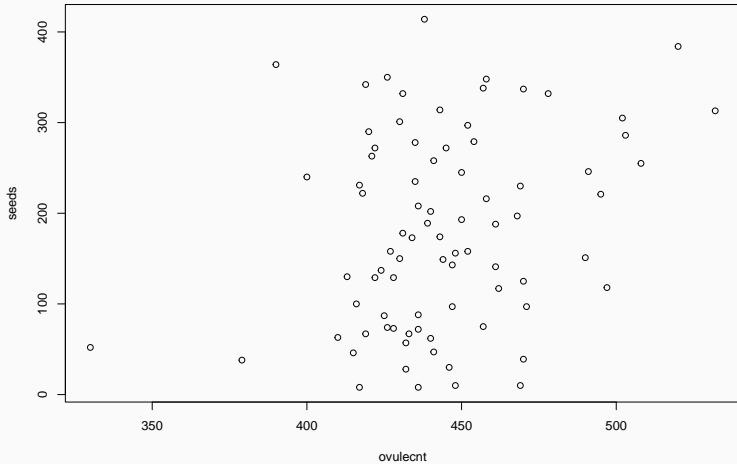
- Is seed set related to proportion of outcross pollen (p_{cmass})?

Questions:

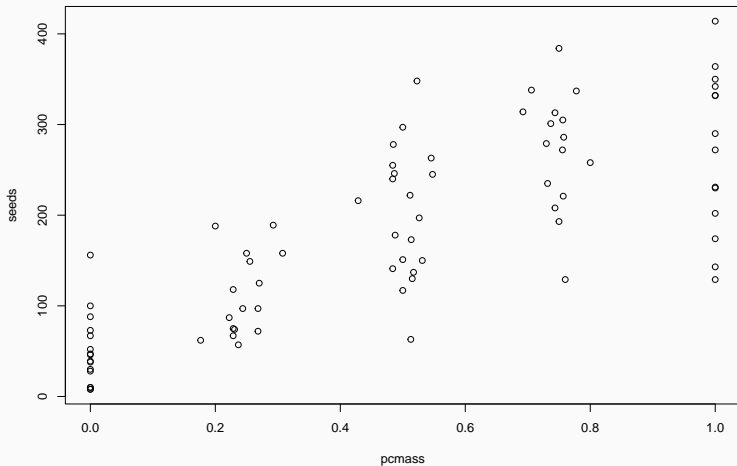
<https://pollev.com/franciscorod726>

- Is seed set related to proportion of outcross pollen (pcmass)?
- Which plant had lower seed set?

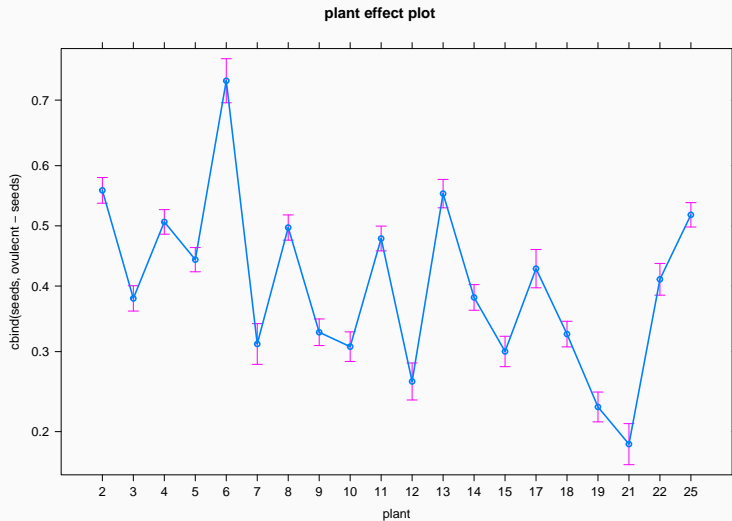
Number of seeds vs Number of ovules



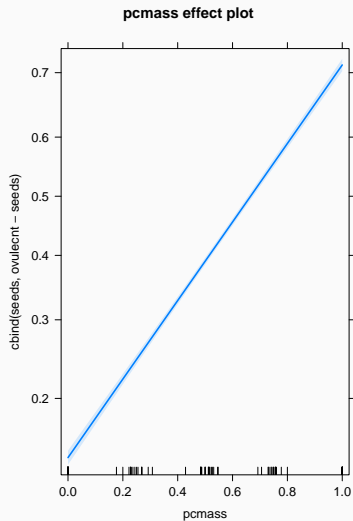
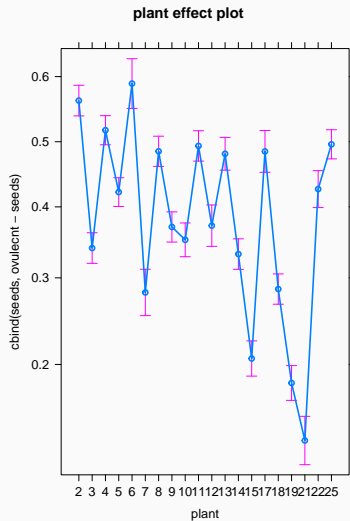
Number of seeds vs Proportion outcross pollen



Seed set across plants



Seed set ~ outcross pollen



GLM for count data: Poisson regression

Francisco Rodríguez-Sánchez

<https://frodriguezsanchez.net>

- Gaussian: lm

- Gaussian: `lm`
- Binary: `glm (family binomial / quasibinomial)`

- Gaussian: `lm`
- Binary: `glm (family binomial / quasibinomial)`
- Counts: `glm (family poisson / quasipoisson)`

Poisson regression

- Response variable: Counts (0, 1, 2, 3...) - discrete
- Link function: \log

Then

$$\log(N) = a + bx$$

$$N = e^{a+bx}$$

Example dataset: Seedling counts in quadrats

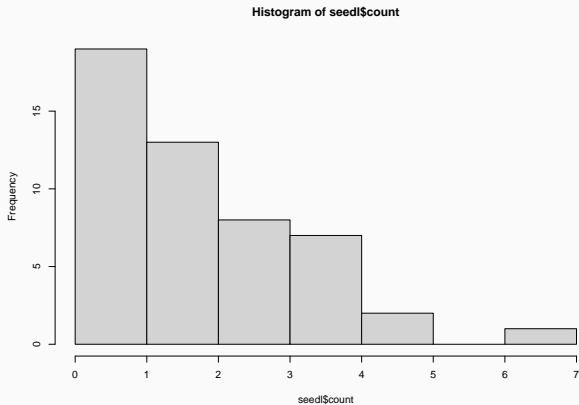
```
seedl <- read.csv("data/seedlings.csv")
```

sample	count	light	area
Min. : 1.00	Min. :0.00	Min. : 2.571	Min. :0.25
1st Qu.:13.25	1st Qu.:1.00	1st Qu.:26.879	1st Qu.:0.25
Median :25.50	Median :2.00	Median :47.493	Median :0.50
Mean :25.50	Mean :2.14	Mean :47.959	Mean :0.62
3rd Qu.:37.75	3rd Qu.:3.00	3rd Qu.:67.522	3rd Qu.:1.00
Max. :50.00	Max. :7.00	Max. :99.135	Max. :1.00

Exploring the data

```
table(seed$count)
```

```
0  1  2  3  4  5  7  
7 12 13  8  7  2  1
```




```
seedl.glm <- glm(count ~ light,  
                 data = seedl,  
                 family = poisson)
```

which corresponds to

```
equatiomatic::extract_eq(seedl.glm)
```

$$\log(E(\text{count})) = \alpha + \beta_1(\text{light}) \quad (1)$$

Interpreting Poisson GLM

Call:

```
glm(formula = count ~ light, family = poisson, data = seed1)
```

Deviance Residuals:

Min	1Q	Median	3Q	Max
-2.1906	-0.8466	-0.1110	0.5220	2.4577

Coefficients:

	Estimate	Std. Error	z value	Pr(> z)
(Intercept)	0.881805	0.188892	4.668	3.04e-06 ***
light	-0.002576	0.003528	-0.730	0.465

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

(Dispersion parameter for poisson family taken to be 1)

Null deviance: 63.029 on 49 degrees of freedom
Residual deviance: 62.492 on 48 degrees of freedom
AIC: 182.03

Number of Fisher Scoring iterations: 5

Parameter estimates are in log scale!

Parameter estimates (log scale):

```
coef(seedl.glm)[1]
```

(Intercept)

0.881805

We need to back-transform: apply the inverse of the logarithm

```
exp(coef(seedl.glm)[1])
```

(Intercept)

2.415255


```
allEffects(seedl.glm)
```

```
model: count ~ light
```

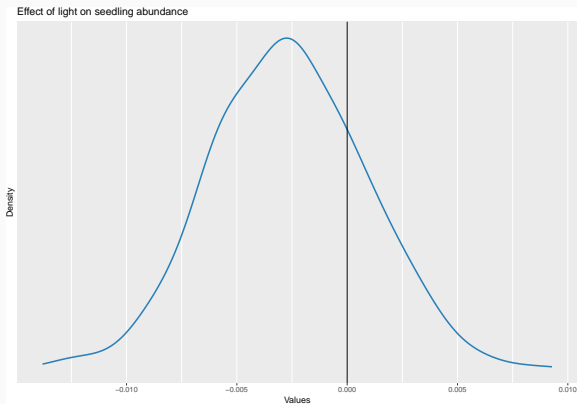
```
light effect
```

```
light
```

	3	30	50	70	100
	2.396665	2.235657	2.123408	2.016794	1.866826

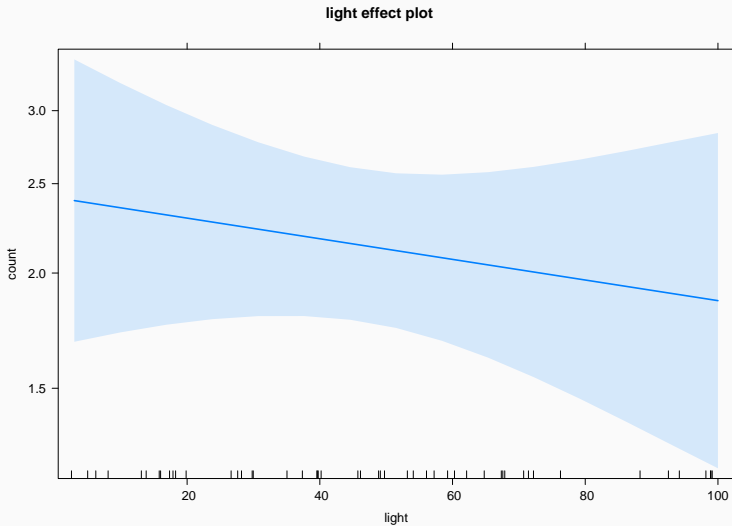
Estimated distribution of the slope parameter

```
library("parameters")  
plot(simulate_parameters(seedl.glm)) +  
  geom_vline(xintercept = 0) +  
  ggtitle("Effect of light on seedling abundance")
```



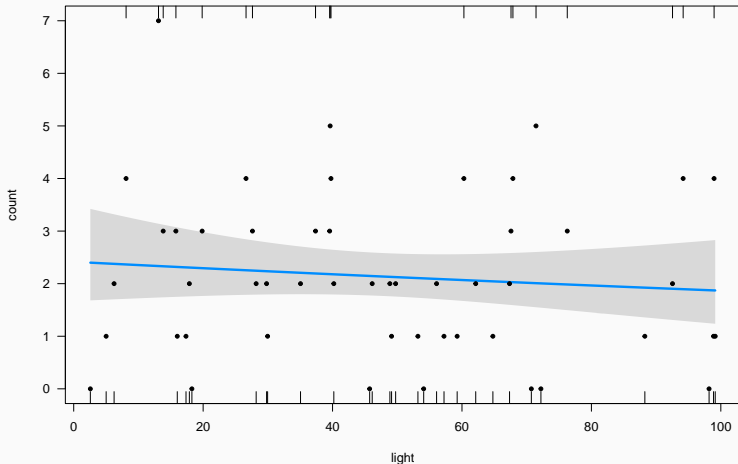
So what's the relationship between Nseedlings and light?

```
plot(allEffects(seedl.glm))
```



Using visreg

```
visreg(seedl.glm, scale = "response", ylim = c(0, 7))  
points(count ~ light, data = seedl, pch = 20)
```



```
library("performance")  
r2(seedl.glm)
```

```
# R2 for Generalized Linear Regression  
Nagelkerke's R2: 0.015
```

Describing the model results

```
library("report")  
report(seedl.glm)
```

We fitted a poisson model (estimated using ML) to predict count with light (formula: count ~ light). The model's explanatory power is very weak (Nagelkerke's $R^2 = 0.01$). The model's intercept, corresponding to light = 0, is at 0.88 (95% CI [0.50, 1.24], $p < .001$). Within this model:

- The effect of light is statistically non-significant and negative (beta = $-2.58e-03$, 95% CI [$-9.57e-03$, $4.28e-03$], $p = 0.465$; Std. beta = -0.07 , 95% CI [-0.27 , 0.12])

Standardized parameters were obtained by fitting the model on a standardized version of the dataset. 95% Confidence Intervals (CIs) and p-values were computed using a Wald z-distribution approximation.

Model checking

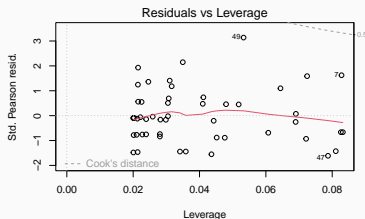
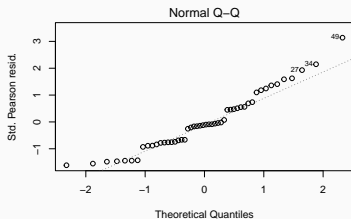
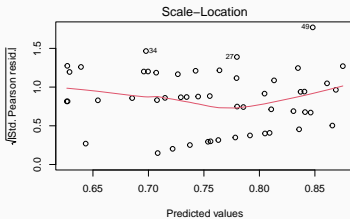
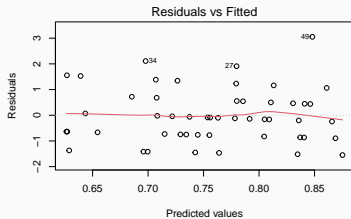
- Linearity (log response \sim predictors)

- Linearity (log response \sim predictors)
- Observations are independent

- Linearity (log response \sim predictors)
- Observations are independent
- Mean = Variance

Checking Poisson GLM

```
plot(seedl.glm)
```



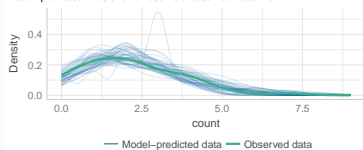
```
null device
```

Checking Poisson GLM

```
check_model(seedl.glm)
```

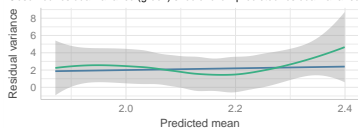
Posterior Predictive Check

Model-predicted lines should resemble observed data line



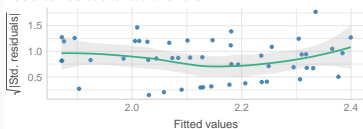
Overdispersion and zero-inflation

Observed residual variance (green) should follow predicted residual variance (blue)



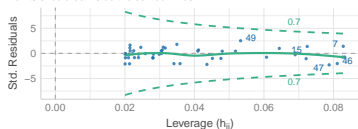
Homogeneity of Variance

Reference line should be flat and horizontal



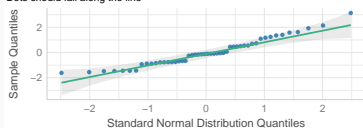
Influential Observations

Points should be inside the contour lines



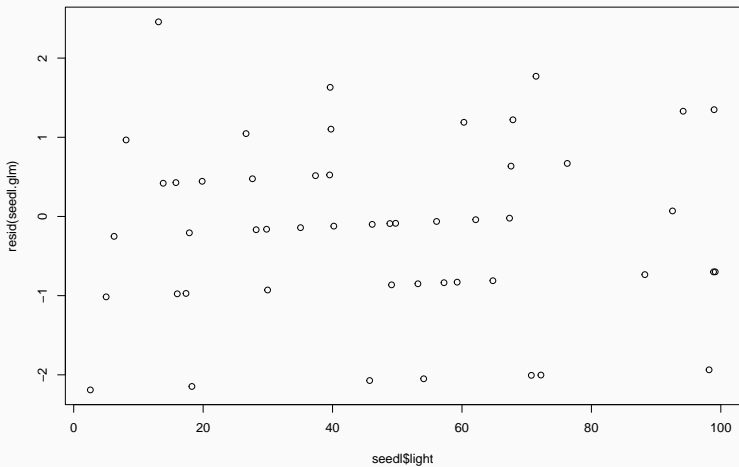
Normality of Residuals

Dots should fall along the line



Is there pattern of residuals along predictor?

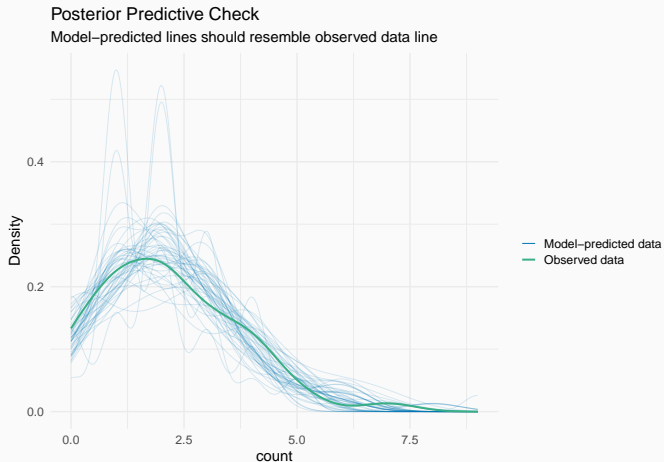
```
plot(seedl$light, resid(seedl.glm))
```



Posterior predictive checking

Simulate data from fitted model (`yrep`) and compare with observed data (`y`)

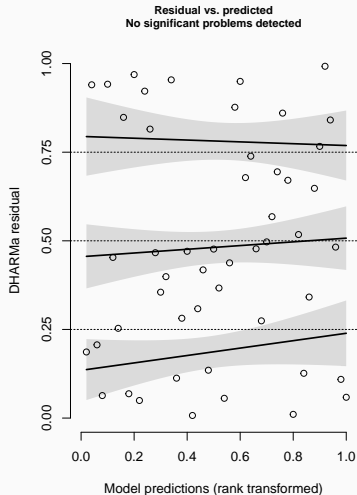
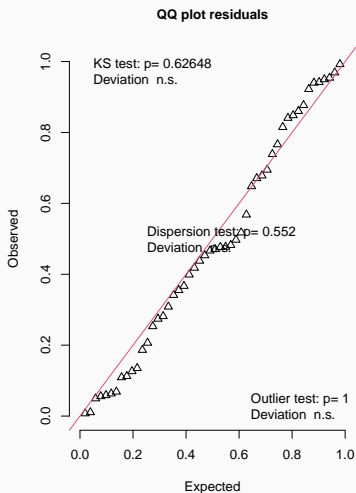
```
check_predictions(seed1.glm)
```



Residuals diagnostics with DHARMA

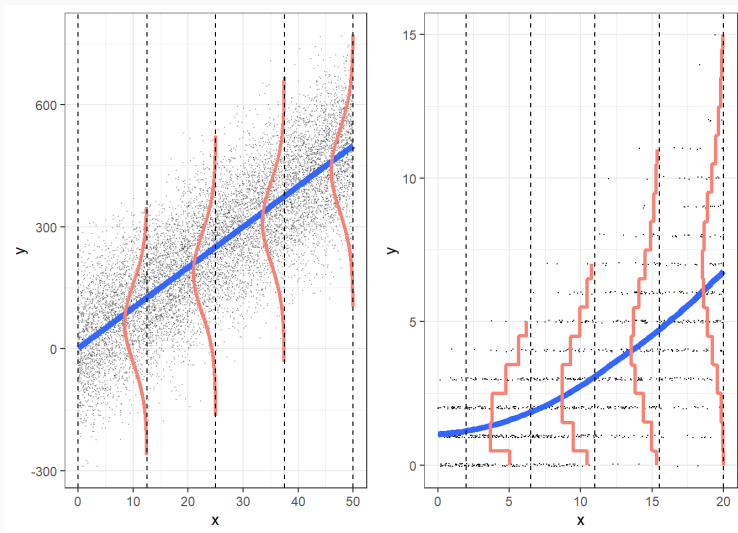
```
simulateResiduals(seedl.glm, plot = TRUE)
```

DHARMA residual



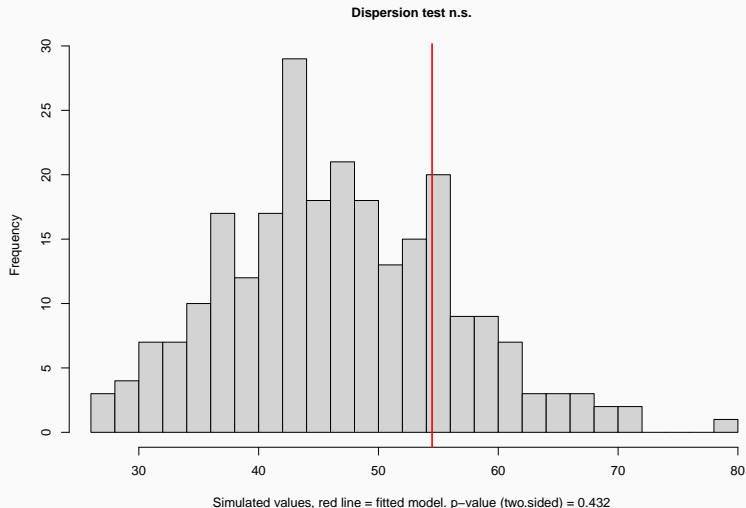
Overdispersion

Poisson GLM assumes mean = variance



Always check overdispersion with count data

```
simres <- simulateResiduals(seedl.glm, refit = TRUE)  
testDispersion(simres)
```



- Use family quasipoisson

- Use family `quasipoisson`
- Use negative binomial distribution (`MASS::glm.nb`)

- Use family `quasipoisson`
- Use negative binomial distribution (`MASS::glm.nb`)
- Include observation-level random effect (e.g. see [Harrison 2014](#))

Accounting for overdispersion with family quasipoisson

Call:

```
glm(formula = count ~ light, family = quasipoisson, data = seedl)
```

Deviance Residuals:

Min	1Q	Median	3Q	Max
-2.1906	-0.8466	-0.1110	0.5220	2.4577

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	0.881805	0.201230	4.382	6.37e-05 ***
light	-0.002576	0.003758	-0.685	0.496

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

(Dispersion parameter for quasipoisson family taken to be 1.134907)

Null deviance: 63.029 on 49 degrees of freedom
Residual deviance: 62.492 on 48 degrees of freedom
AIC: NA

Number of Fisher Scoring iterations: 5

Mean estimates do not change after accounting for overdispersion

```
allEffects(seedl.overdisp)
```

```
model: count ~ light
```

```
light effect
```

```
light
```

	3	30	50	70	100
	2.396665	2.235657	2.123408	2.016794	1.866826

```
allEffects(seedl.glm)
```

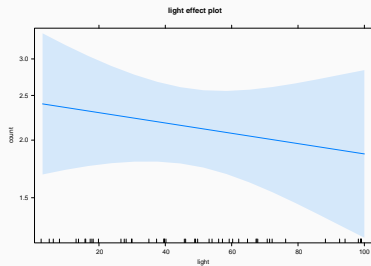
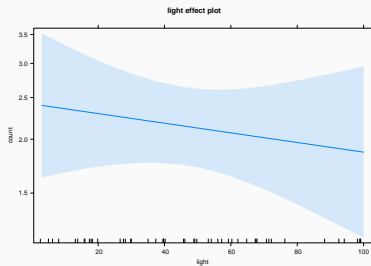
```
model: count ~ light
```

```
light effect
```

```
light
```

	3	30	50	70	100
	2.396665	2.235657	2.123408	2.016794	1.866826

But standard errors may change



Accounting for overdispersion using negative binomial

```
library("MASS")  
seedl.nb <- glm.nb(count ~ light, data = seedl)
```

Call:

```
glm.nb(formula = count ~ light, data = seedl, init.theta = 22.23419419,  
       link = log)
```

Deviance Residuals:

Min	1Q	Median	3Q	Max
-2.1349	-0.8162	-0.1061	0.4954	2.2814

Coefficients:

	Estimate	Std. Error	z value	Pr(> z)
(Intercept)	0.881996	0.198213	4.450	8.6e-06 ***
light	-0.002580	0.003691	-0.699	0.485

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

(Dispersion parameter for Negative Binomial(22.2342) family taken to be 1)

Null deviance: 58.247 on 49 degrees of freedom

Comparing Poisson and Negative Binomial

```
compare_models(seedl.glm, seedl.nb)
```

Parameter	seedl.glm	seedl.nb
(Intercept)	0.88 (0.51, 1.25)	0.88 (0.49, 1.27)
light	-2.58e-03 (-0.01, 0.00)	-2.58e-03 (-0.01, 0.00)
Observations	50	50

```
compare_performance(seedl.glm, seedl.nb)
```

```
# Comparison of Model Performance Indices
```

Name	Model	AIC	AIC weights	BIC	BIC weights	Nagelkerke's R2	RMSE
seedl.glm	glm	182.034	0.710	185.858	0.864	0.015	1.529
seedl.nb	negbin	183.827	0.290	189.563	0.136	0.014	1.529

What if survey plots have
different area?

Shall we *standardise* counts dividing by sampling plot area?

Model would be: $\text{count/area} \sim \text{light}$

	sample	count	light	area
1	1	0	70.71854	0.50
2	2	1	88.26021	0.25
3	3	2	67.35133	0.50
4	4	3	67.57850	1.00
5	5	4	26.63098	0.25
6	6	3	15.79433	1.00

J. R. Statist. Soc. A (1993)
156, Part 3, pp. 379–392

Spurious Correlation and the Fallacy of the Ratio Standard Revisited

By RICHARD A. KRONMAL†

<https://doi.org/10.2307/2983064>

Use offset to account for variable sampling effort

```
seedl.offset <- glm(count ~ light,  
                    offset = log(area),  
                    data = seedl,  
                    family = poisson)
```

Note estimates now referred to area units!

Call:

```
glm(formula = count ~ light, family = poisson, data = seedl,  
     offset = log(area))
```

Deviance Residuals:

Min	1Q	Median	3Q	Max
-2.9918	-1.0142	0.1673	0.8401	3.8230

Coefficients:

	Estimate	Std. Error	z value	Pr(> z)
(Intercept)	1.513185	0.183245	8.258	<2e-16 ***
light	-0.005674	0.003384	-1.677	0.0936 .

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

(Dispersion parameter for poisson family taken to be 1)

```
exp(coef(seedl.offset)[1])
```

(Intercept)

4.541173

Prediction

Predicting number of seedlings given light

```
new.lights <- data.frame(light = c(10, 90))  
predict(seedl.glm, newdata = new.lights, type = "response", se.fit
```

```
$fit
```

```
      1      2  
2.353841 1.915533
```

```
$se.fit
```

```
      1      2  
0.3756992 0.3502446
```

```
$residual.scale
```

```
[1] 1
```

- Infant mortality \sim GDP

- Infant mortality ~ GDP
- Number of cones consumed by squirrels ([data](#))

- Infant mortality ~ GDP
- Number of cones consumed by squirrels ([data](#))
- Elephant matings ([Poole 1989](#))

Mixed / Multilevel Models

Francisco Rodríguez-Sánchez

<https://frodriguezsanchez.net>

Example dataset: trees

- Data on 1000 trees from 10 sites.

```
head(trees)
```

	site	dbh	height	sex	dead
1	4	29.68	36.1	male	0
2	5	33.29	42.3	male	0
3	2	28.03	41.9	female	0
4	5	39.86	46.5	female	0
5	1	47.94	43.9	female	0
6	1	10.82	26.2	male	0

Example dataset: trees

- Data on 1000 trees from 10 sites.
- Trees per site: 4 - 392.

```
head(trees)
```

	site	dbh	height	sex	dead
1	4	29.68	36.1	male	0
2	5	33.29	42.3	male	0
3	2	28.03	41.9	female	0
4	5	39.86	46.5	female	0
5	1	47.94	43.9	female	0
6	1	10.82	26.2	male	0

Q: What's the relationship
between tree diameter and
height?

A simple linear model

```
lm.simple <- lm(height ~ dbh, data = trees)
```

Call:

```
lm(formula = height ~ dbh, data = trees)
```

Residuals:

Min	1Q	Median	3Q	Max
-13.3270	-2.8978	0.1057	2.7924	12.9511

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)	
(Intercept)	19.33920	0.31064	62.26	<2e-16	***
dbh	0.61570	0.01013	60.79	<2e-16	***

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Remember our model structure

$$y_i \sim N(\mu_i, \sigma^2)$$

$$\mu_i = \alpha + \beta x_i$$

In this case:

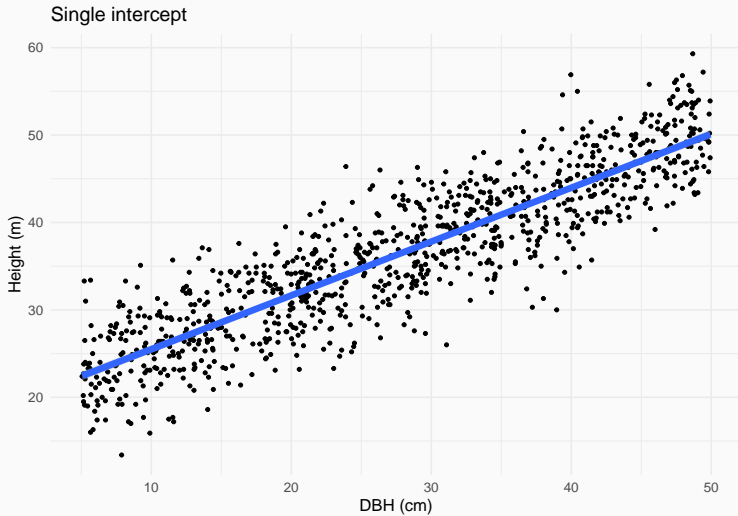
$$\text{Height}_i \sim N(\mu_i, \sigma^2)$$

$$\mu_i = \alpha + \beta \text{DBH}_i$$

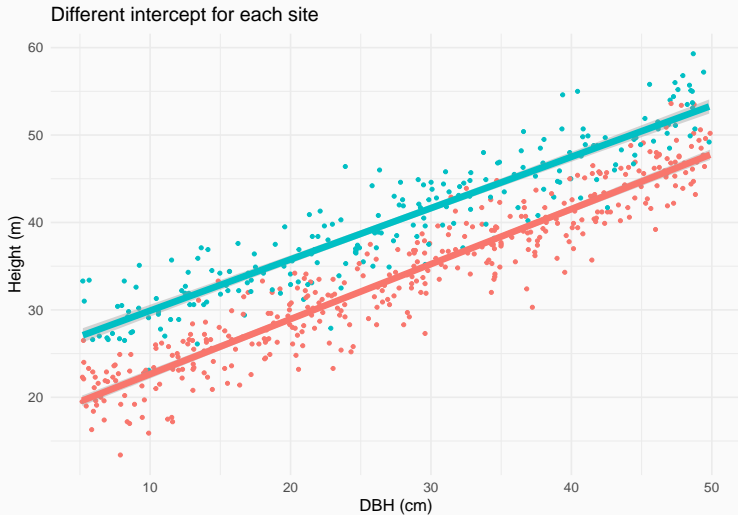
α : expected height when DBH = 0

β : how much height increases with every unit increase of DBH

There is only one intercept



What if allometry varies among sites?



Fitting a varying intercepts model with `lm`

Call:

```
lm(formula = height ~ factor(site) + dbh, data = trees)
```

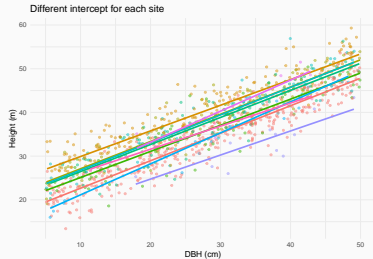
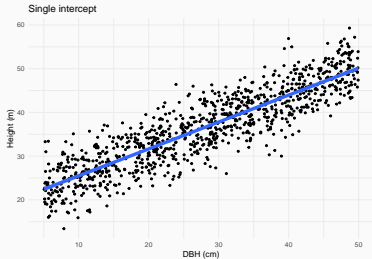
Residuals:

Min	1Q	Median	3Q	Max
-10.1130	-1.9885	0.0582	2.0314	11.3320

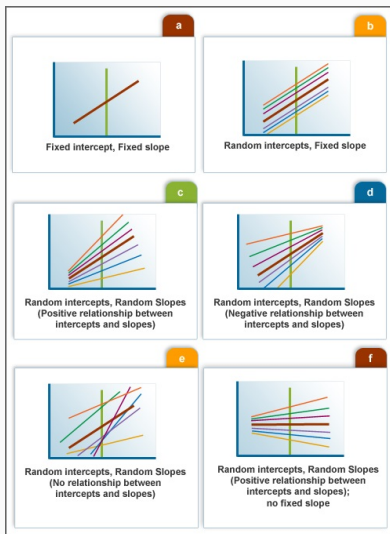
Coefficients:

	Estimate	Std. Error	t value	Pr(> t)	
(Intercept)	16.699037	0.260565	64.088	< 2e-16	***
factor(site)2	6.504303	0.256730	25.335	< 2e-16	***
factor(site)3	4.357457	0.354181	12.303	< 2e-16	***
factor(site)4	1.934650	0.356102	5.433	6.98e-08	***
factor(site)5	3.637432	0.339688	10.708	< 2e-16	***
factor(site)6	4.204511	0.421906	9.966	< 2e-16	***
factor(site)7	-0.176193	0.666772	-0.264	0.7916	
factor(site)8	-5.312648	0.893603	-5.945	3.82e-09	***
factor(site)9	5.437049	1.087766	4.998	6.84e-07	***
factor(site)10	2.263338	1.369986	1.652	0.0988	.
dbh	0.617075	0.007574	81.473	< 2e-16	***

Single vs varying intercept



Mixed models enable us to account for variability



www.esourceresearch.org/

$$y_i = a + \alpha_j + b \cdot x_i + \varepsilon_i$$

$$\alpha_j \sim N(0, \tau^2)$$

$$\varepsilon_i \sim N(0, \sigma^2)$$

In our example:

$$\text{Height}_i = a + \text{site}_j + b \cdot \text{DBH}_i + \varepsilon_i$$

$$\text{site}_j \sim N(0, \tau^2)$$

$$\varepsilon_i \sim N(0, \sigma^2)$$

Mixed models estimate **varying parameters**

(intercepts and/or slopes)

with pooling among levels

(rather than considering them fully independent)

Hence there's gradient between

- **complete pooling:** Single overall intercept.

Hence there's gradient between

- complete pooling: Single overall intercept.
 - `lm (height ~ dbh)`

Hence there's gradient between

- **complete pooling:** Single overall intercept.
 - `lm (height ~ dbh)`
- **no pooling:** One *independent* intercept for each site.

Hence there's gradient between

- **complete pooling:** Single overall intercept.
 - `lm (height ~ dbh)`
- **no pooling:** One *independent* intercept for each site.
 - `lm (height ~ dbh + site)`

Hence there's gradient between

- **complete pooling:** Single overall intercept.
 - `lm (height ~ dbh)`
- **no pooling:** One *independent* intercept for each site.
 - `lm (height ~ dbh + site)`
- **partial pooling:** Inter-related intercepts.

Hence there's gradient between

- **complete pooling:** Single overall intercept.
 - `lm (height ~ dbh)`
- **no pooling:** One *independent* intercept for each site.
 - `lm (height ~ dbh + site)`
- **partial pooling:** Inter-related intercepts.
 - `lmer(height ~ dbh + (1 | site))`

1. Fixed effects constant across individuals, random effects vary.

http://andrewgelman.com/2005/01/25/why_i_dont_use/

Random vs Fixed effects?

1. Fixed effects constant across individuals, random effects vary.
2. Effects are fixed if they are interesting in themselves; random if interest in the underlying population.

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Random vs Fixed effects?

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Random vs Fixed effects?

1. Fixed effects constant across individuals, random effects vary.
2. Effects are fixed if they are interesting in themselves; random if interest in the underlying population.
3. Fixed when sample exhausts the population; random when the sample is small part of the population.
4. Random effect if it's assumed to be a realized value of random variable.
5. Fixed effects estimated using least squares or maximum likelihood; random effects estimated with shrinkage.

http://andrewgelman.com/2005/01/25/why_i_dont_use/

What is a random effect, really?

- Varies by group

Random effects are estimated with *partial pooling*, while fixed effects are not (infinite variance).

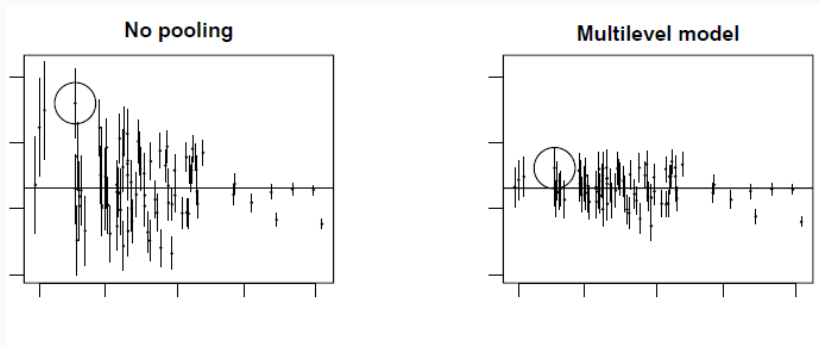
What is a random effect, really?

- Varies by group
- Variation estimated with **probability model**

Random effects are estimated with *partial pooling*, while fixed effects are not (infinite variance).

Shrinkage improves parameter estimation

Especially for groups with low sample size



From Gelman & Hill p. 253

Fitting mixed/multilevel models

```
library("lme4")  
mixed <- lmer(height ~ dbh + (1|site), data = trees)
```

Linear mixed model fit by REML [`'lmerMod'`]

Formula: `height ~ dbh + (1 | site)`

Data: `trees`

REML criterion at convergence: 5108.3

Scaled residuals:

Min	1Q	Median	3Q	Max
-3.3199	-0.6607	0.0227	0.6716	3.7328

Random effects:

Groups	Name	Variance	Std.Dev.
site	(Intercept)	11.195	3.346
	Residual	9.261	3.043

Number of obs: 1000, groups: site, 10

Fixed effects:

	Estimate	Std. Error	t value
(Intercept)	19.011468	1.100444	17.28
dbh	0.616927	0.007572	81.47

Correlation of Fixed Effects:

(Intr)

Retrieve model coefficients

```
coef(mixed)
```

```
$site
```

```
  (Intercept)      dbh
```

```
1      16.70800 0.6169271
2      23.19162 0.6169271
3      21.04229 0.6169271
4      18.64086 0.6169271
5      20.32995 0.6169271
6      20.88200 0.6169271
7      16.61686 0.6169271
8      11.88302 0.6169271
9      21.84779 0.6169271
10     18.97228 0.6169271
```

```
attr(,"class")
```

```
[1] "coef.mer"
```

Broom: model estimates in tidy form

```
library(broom.mixed)
tidy(mixed)
```

```
# A tibble: 4 x 6
```

	effect	group	term	estimate	std.error	statistic
	<chr>	<chr>	<chr>	<dbl>	<dbl>	<dbl>
1	fixed	<NA>	(Intercept)	19.0	1.10	17.3
2	fixed	<NA>	dbh	0.617	0.00757	81.5
3	ran_pars	site	sd__(Intercept)	3.35	NA	NA
4	ran_pars	Residual	sd__Observation	3.04	NA	NA

See also [broom.mixed](#)

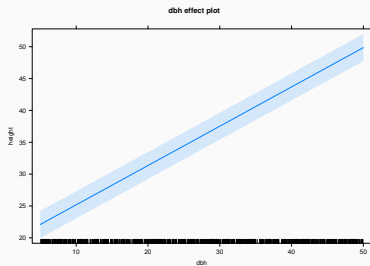
Visualising model: allEffects

```
allEffects(mixed)
```

```
model: height ~ dbh
```

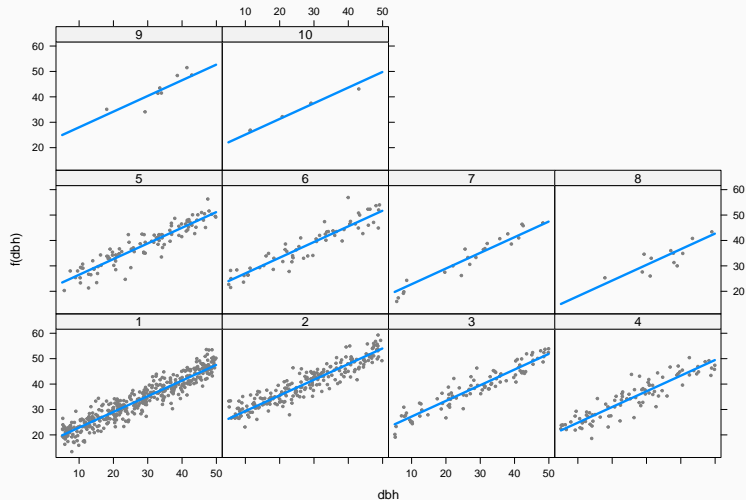
```
dbh effect
```

```
dbh  
      5      20      30      40      50  
22.09610 31.35001 37.51928 43.68855 49.85782
```



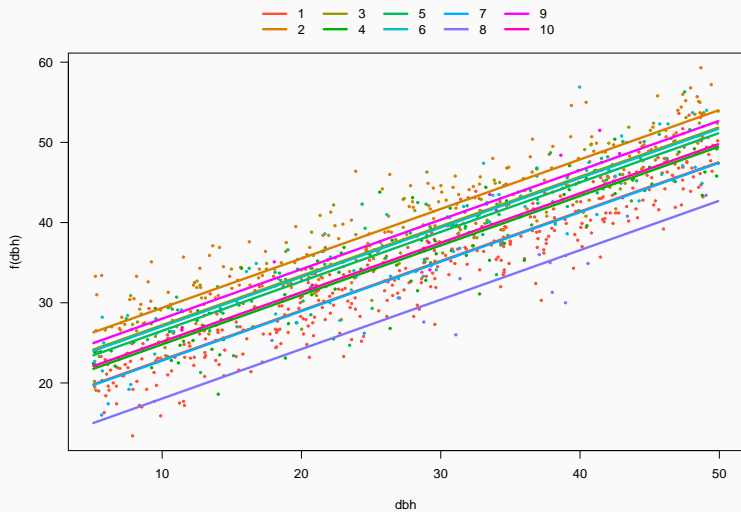
Visualising model: visreg

```
visreg(mixed, xvar = "dbh", by = "site", re.form = NULL)
```



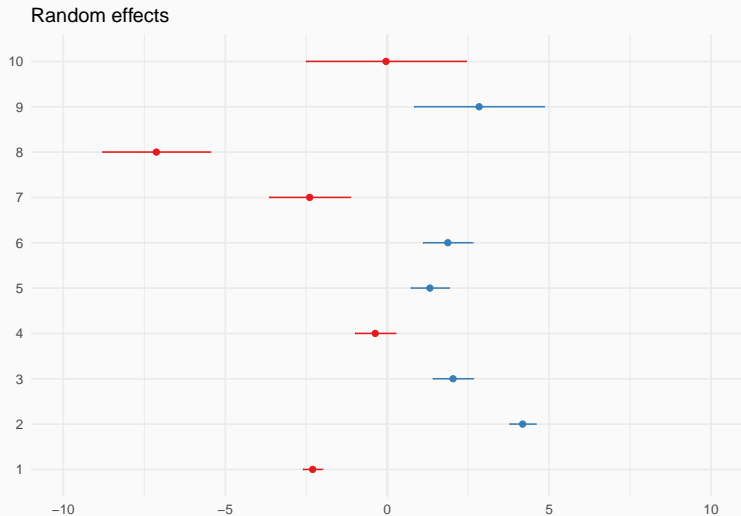
Visualising model

```
visreg(mixed, xvar = "dbh", by = "site", re.form = NULL, overlay = TRUE)
```



Visualising model: sjPlot

```
sjPlot::plot_model(mixed, type = "re")
```

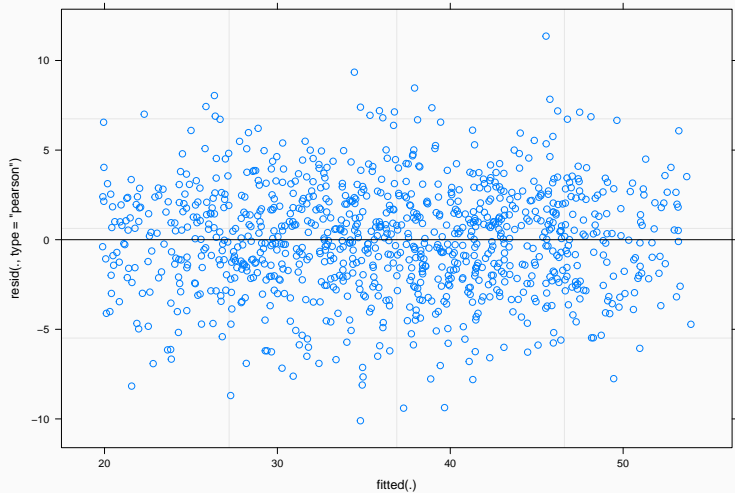


Using merTools to understand fitted model

```
library("merTools")  
shinyMer(mixed)
```


Checking residuals

```
plot(mixed)
```

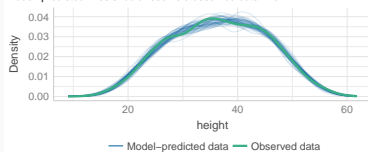


Checking residuals

```
library("performance")  
check_model(mixed)
```

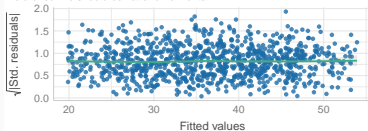
Posterior Predictive Check

Model-predicted lines should resemble observed data line



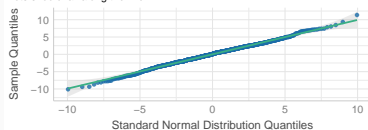
Homogeneity of Variance

Reference line should be flat and horizontal



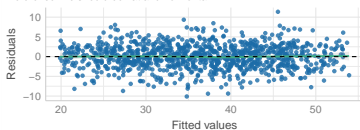
Normality of Residuals

Dots should fall along the line



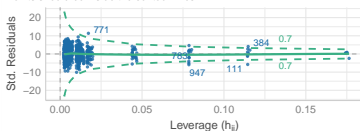
Linearity

Reference line should be flat and horizontal



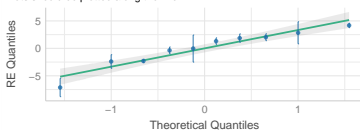
Influential Observations

Points should be inside the contour lines



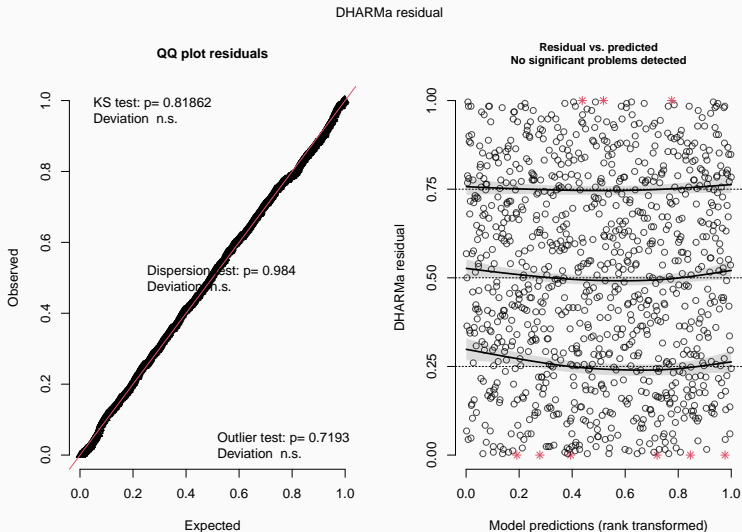
Normality of Random Effects (site)

Dots should be plotted along the line



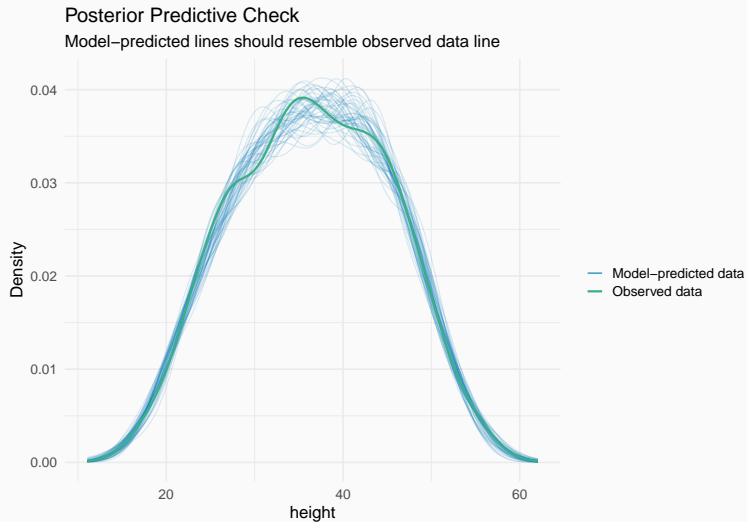
Checking residuals (DHARMA)

```
DHARMA::simulateResiduals(mixed, plot = TRUE, re.form = NULL)
```



Model checking with simulated data

```
check_predictions(mixed)
```



Many approaches! Somewhat polemic (e.g. see [this](#)).

Nakagawa & Schielzeth propose **marginal** (considering fixed effects only) and **conditional** R^2 (including random effects too):

```
r2(mixed)
```

```
# R2 for Mixed Models
```

```
Conditional R2: 0.888
```

```
  Marginal R2: 0.753
```

Growing the hierarchy: adding site-level predictors

Model with group-level predictors

We had:

$$y_i = a + \alpha_j + b \cdot x_i + \varepsilon_i$$

$$\alpha_j \sim N(0, \tau^2)$$

$$\varepsilon_i \sim N(0, \sigma^2)$$

Now

$$y_i = a + \alpha_j + b \cdot x_i + \varepsilon_i$$

$$\alpha_j \sim N(\mu_j, \tau^2)$$

$$\mu_j = \delta \cdot \text{Predictor}_j$$

$$\varepsilon_i \sim N(0, \sigma^2)$$

Are height differences among sites related to temperature?

$$\text{Height}_i = \text{site}_j + b \cdot \text{DBH}_i + \varepsilon_i$$

$$\text{site}_j \sim N(\mu_j, \tau^2)$$

$$\mu_j = a + \delta \cdot \text{Temperature}_j$$

$$\varepsilon_i \sim N(0, \sigma^2)$$

Are height differences among sites related to temperature?

```
sitedata <- read.csv("data/sitedata.csv")  
sitedata
```

	site	temp
1	1	15.1
2	2	22.0
3	3	20.1
4	4	20.4
5	5	20.0
6	6	20.1
7	7	17.5
8	8	14.6
9	9	19.2
10	10	16.0

Merging trees and site data

```
trees.full <- merge(trees, sitedata, by = "site")  
head(trees.full)
```

	site	dbh	height	sex	dead	temp
1	1	21.05	32.2	male	0	15.1
2	1	46.63	45.9	female	0	15.1
3	1	43.86	45.5	male	0	15.1
4	1	29.03	35.5	male	0	15.1
5	1	6.02	21.1	male	0	15.1
6	1	40.82	38.7	male	0	15.1

Fit multilevel model

```
group.pred <- lmer(height ~ dbh + (1 | site) + temp, data = trees.full)
```

Linear mixed model fit by REML [`'lmerMod'`]

Formula: `height ~ dbh + (1 | site) + temp`

Data: `trees.full`

REML criterion at convergence: 5098.2

Scaled residuals:

Min	1Q	Median	3Q	Max
-3.3247	-0.6517	0.0192	0.6663	3.7268

Random effects:

Groups	Name	Variance	Std.Dev.
site	(Intercept)	3.158	1.777
Residual		9.266	3.044

Number of obs: 1000, groups: site, 10

Fixed effects:

	Estimate	Std. Error	t value
(Intercept)	-1.730910	4.671330	-0.371
dbh	0.616894	0.007571	81.484
temp	1.115104	0.248000	4.496

Correlation of Fixed Effects:

(Intr) dbh

Centre (and scale) continuous variables

```
mean(sitedata$temp)
```

```
[1] 18.5
```

```
trees.full$temp.c <- trees.full$temp - 18
```

Temperatures now referred as deviations from 18 °C (close to average)

Fit multilevel model

```
group.pred <- lmer(height ~ dbh + (1 | site) + temp.c, data = trees.full)
```

```
Linear mixed model fit by REML ['lmerMod']  
Formula: height ~ dbh + (1 | site) + temp.c  
Data: trees.full
```

REML criterion at convergence: 5098.2

Scaled residuals:

Min	1Q	Median	3Q	Max
-3.3247	-0.6517	0.0192	0.6663	3.7268

Random effects:

Groups	Name	Variance	Std.Dev.
site	(Intercept)	3.158	1.777
Residual		9.266	3.044

Number of obs: 1000, groups: site, 10

Fixed effects:

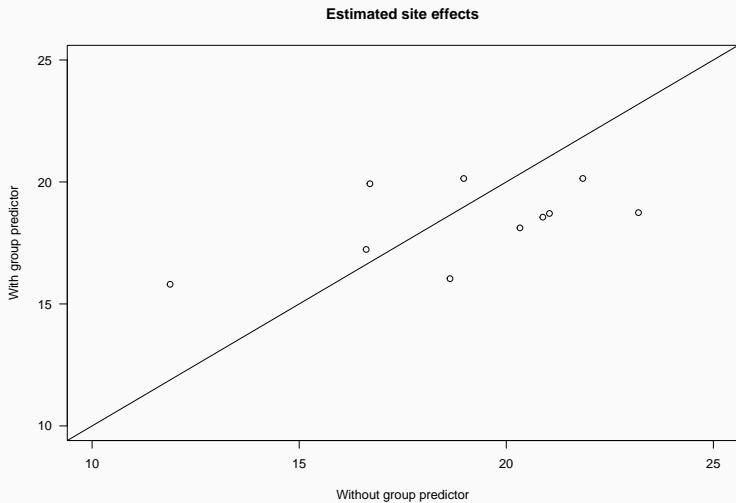
	Estimate	Std. Error	t value
(Intercept)	18.340954	0.655054	27.999
dbh	0.616894	0.007571	81.484
temp.c	1.115104	0.248000	4.496

Correlation of Fixed Effects:

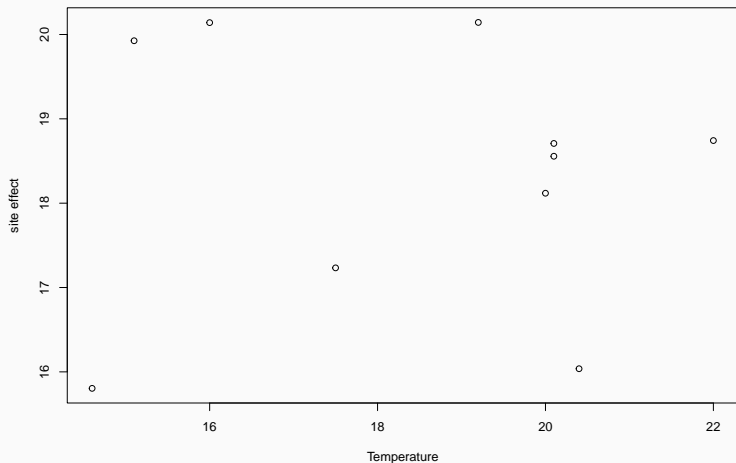
(Intr) dbh

```
shinyMer(group.pred)
```

Comparing site effects with and without group predictor



Are site effects related to temperature?



Varying intercepts and slopes

Varying intercepts and slopes

There is overall difference in height among sites (different intercepts)

AND

Relationship between DBH and Height varies among sites (different slopes)

```
mixed.slopes <- lmer(height ~ dbh + (1 + dbh | site), data=trees)
```

Varying intercepts and slopes

```
Linear mixed model fit by REML ['lmerMod']  
Formula: height ~ dbh + (1 + dbh | site)  
Data: trees
```

```
REML criterion at convergence: 5105.1
```

```
Scaled residuals:
```

Min	1Q	Median	3Q	Max
-3.3342	-0.6599	0.0375	0.6916	3.7756

```
Random effects:
```

Groups	Name	Variance	Std.Dev.	Corr
site	(Intercept)	1.566e+01	3.95671	
	dbh	3.087e-04	0.01757	-1.00
	Residual	9.226e+00	3.03744	

```
Number of obs: 1000, groups: site, 10
```

```
Fixed effects:
```

	Estimate	Std. Error	t value
(Intercept)	18.95272	1.29190	14.67
dbh	0.61837	0.00946	65.37

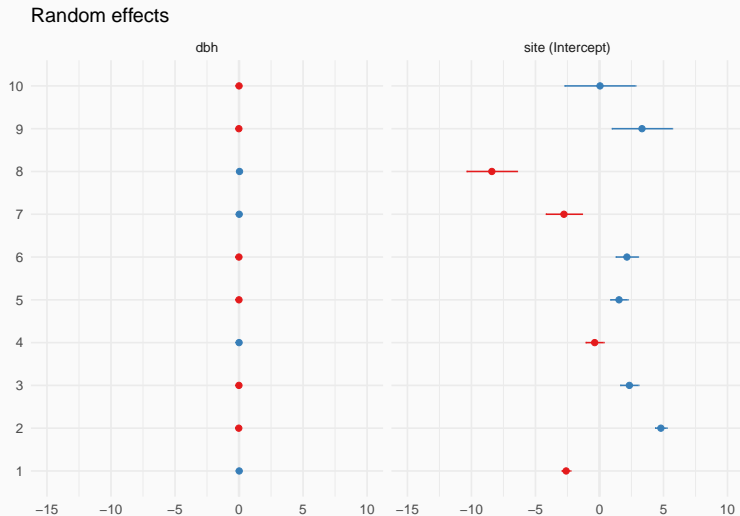
Varying intercepts and slopes

```
$site
  (Intercept)      dbh
1    16.34655 0.6299443
2    23.74733 0.5970814
3    21.28802 0.6080019
4    18.57844 0.6200337
5    20.47961 0.6115916
6    21.09608 0.6088542
7    16.17675 0.6306983
8    10.54681 0.6556978
9    22.27301 0.6036281
10   18.99463 0.6181856
```

```
attr(,"class")
[1] "coef.mer"
```

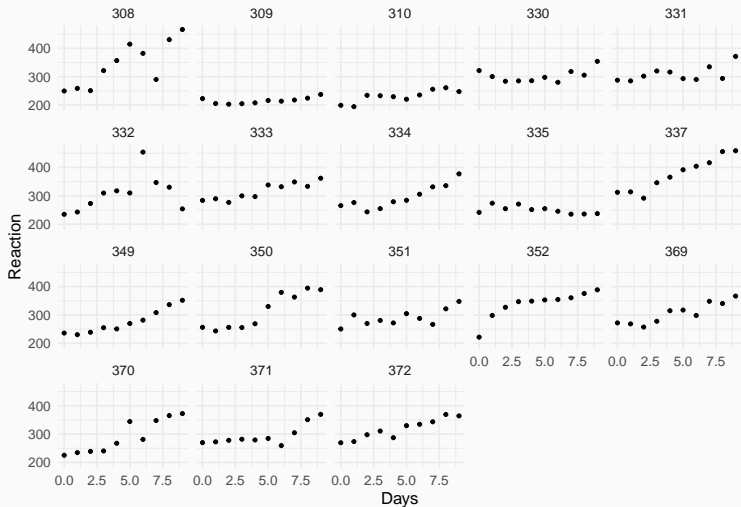
Visualising model: sjPlot

```
plot_model(mixed.slopes, type = "re")
```



More examples

sleepstudy (repeated measures)



Varying intercepts and slopes (lme4)

```
sleep <- lmer(Reaction ~ Days + (1+Days|Subject), data = sleepstudy)
```

Linear mixed model fit by REML ['lmerMod']

Formula: Reaction ~ Days + (1 + Days | Subject)

Data: sleepstudy

REML criterion at convergence: 1743.6

Scaled residuals:

Min	1Q	Median	3Q	Max
-3.9536	-0.4634	0.0231	0.4634	5.1793

Random effects:

Groups	Name	Variance	Std.Dev.	Corr
Subject	(Intercept)	612.10	24.741	
	Days	35.07	5.922	0.07
Residual		654.94	25.592	

Number of obs: 180, groups: Subject, 18

Fixed effects:

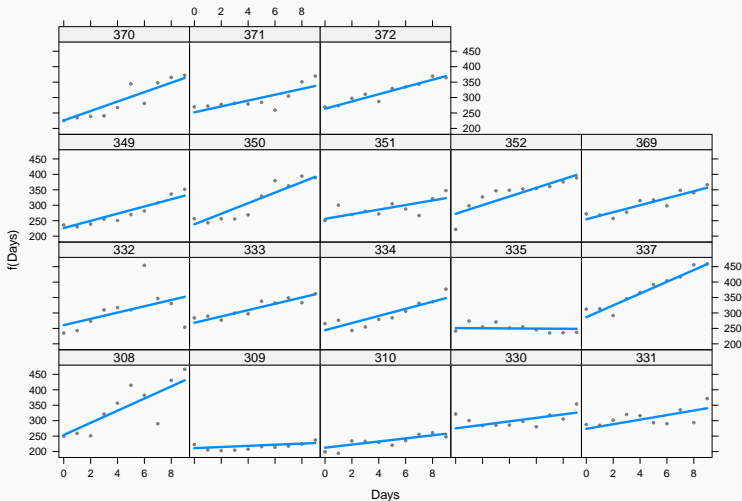
	Estimate	Std. Error	t value
(Intercept)	251.405	6.825	36.838
Days	10.467	1.546	6.771

Correlation of Fixed Effects:

(Intr)

Varying intercepts and slopes (lme4)

```
visreg(sleep, xvar = "Days", by = "Subject", re.form = NULL)
```



Fitting multilevel models (GAMM) with mgcv

```
sgamm <- mgcv::gam(Reaction ~ s(Days, Subject, k = 3, bs = "fs"),  
                  data = sleepstudy, method = "REML")
```

Family: gaussian

Link function: identity

Formula:

Reaction ~ s(Days, Subject, k = 3, bs = "fs")

Parametric coefficients:

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	298.51	9.05	32.98	<2e-16 ***

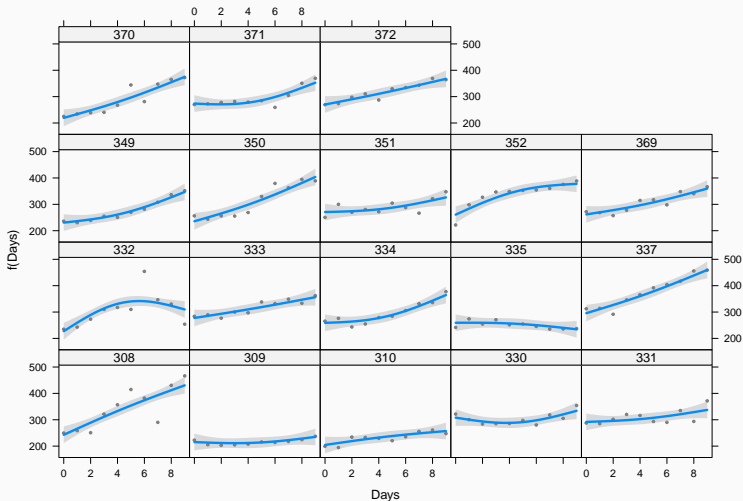
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Approximate significance of smooth terms:

	edf	Ref.df	F	p-value
s(Days,Subject)	45.67	53	17.11	<2e-16 ***

Fitting multilevel models (GAMM) with mgcv

```
visreg(sgamma, xvar = "Days", by = "Subject")
```



Hierarchical generalized additive models: an introduction with mgcv

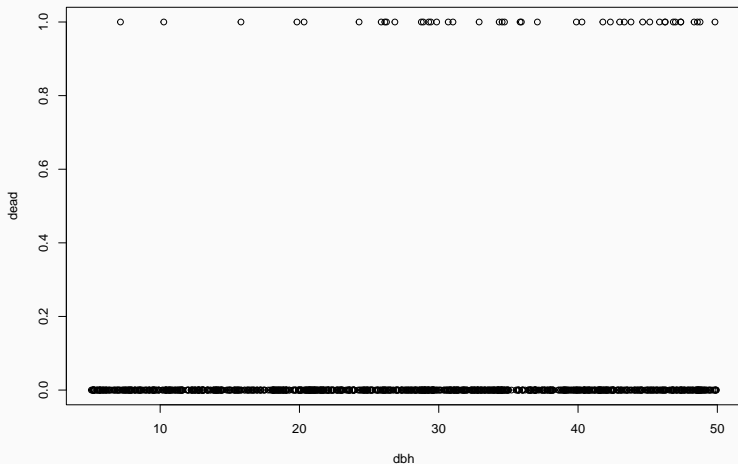
Eric J Pedersen ^{Corresp., 1,2}, David L. Miller ^{3,4}, Gavin L. Simpson ⁵, Noam Ross ⁶

<https://doi.org/10.7287/peerj.preprints.27320v1>

Multilevel logistic regression

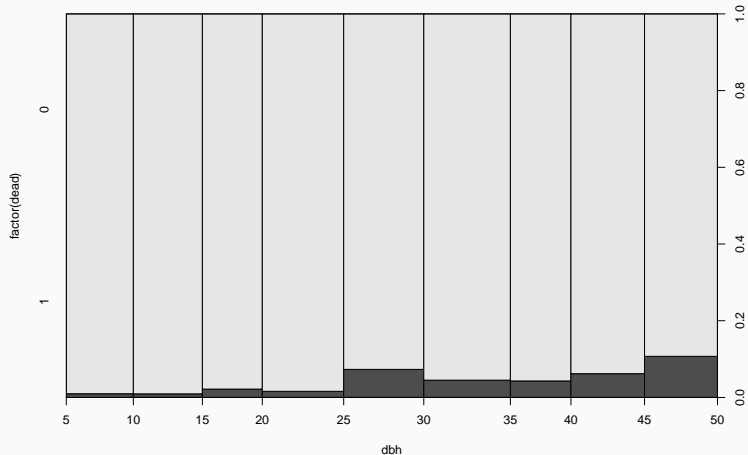
Q: Relationship between tree size and mortality

```
plot(dead ~ dbh, data = trees)
```



Q: Relationship between tree size and mortality

```
plot(factor(dead) ~ dbh, data = trees)
```



Fit simple logistic regression

```
simple.logis <- glm(dead ~ dbh, data = trees, family=binomial)
```

Call:

```
glm(formula = dead ~ dbh, family = binomial, data = trees)
```

Deviance Residuals:

Min	1Q	Median	3Q	Max
-0.4805	-0.3520	-0.2647	-0.1928	2.9690

Coefficients:

	Estimate	Std. Error	z value	Pr(> z)
(Intercept)	-4.77874	0.50902	-9.388	< 2e-16 ***
dbh	0.05365	0.01377	3.895	9.82e-05 ***

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

(Dispersion parameter for binomial family taken to be 1)

Null deviance: 360.91 on 999 degrees of freedom
Residual deviance: 343.69 on 998 degrees of freedom
AIC: 347.69

Number of Fisher Scoring iterations: 6

Logistic regression with *independent* site effects

```
logis2 <- glm(dead ~ dbh + factor(site), data = trees, family=binomial)
```

Call:

```
glm(formula = dead ~ dbh + factor(site), family = binomial, data = trees)
```

Deviance Residuals:

Min	1Q	Median	3Q	Max
-0.6359	-0.3449	-0.2561	-0.1852	2.9763

Coefficients:

	Estimate	Std. Error	z value	Pr(> z)
(Intercept)	-4.80123	0.54985	-8.732	<2e-16 ***
dbh	0.05371	0.01381	3.889	0.0001 ***
factor(site)2	-0.29692	0.46073	-0.644	0.5193
factor(site)3	0.21275	0.52799	0.403	0.6870
factor(site)4	0.39841	0.53025	0.751	0.4524
factor(site)5	-0.42557	0.64018	-0.665	0.5062
factor(site)6	0.66861	0.53656	1.246	0.2127
factor(site)7	0.11862	1.06211	0.112	0.9111
factor(site)8	0.43899	1.08058	0.406	0.6846
factor(site)9	-13.63389	840.90382	-0.016	0.9871
factor(site)10	-13.17148	1042.21823	-0.013	0.9899

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Fit multilevel logistic regression

```
mixed.logis <- glmer(dead ~ dbh + (1|site), data=trees, family = binomial)
```

Generalized linear mixed model fit by maximum likelihood (Laplace
Approximation) [glmerMod]

Family: binomial (logit)

Formula: dead ~ dbh + (1 | site)

Data: trees

AIC	BIC	logLik	deviance	df.resid
349.7	364.4	-171.8	343.7	997

Scaled residuals:

Min	1Q	Median	3Q	Max
-0.3498	-0.2528	-0.1888	-0.1370	9.0031

Random effects:

Groups Name	Variance	Std.Dev.
site (Intercept)	0	0

Number of obs: 1000, groups: site, 10

Fixed effects:

	Estimate	Std. Error	z value	Pr(> z)
(Intercept)	-4.77874	0.50904	-9.388	< 2e-16 ***
dbh	0.05365	0.01377	3.895	9.83e-05 ***

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Retrieve model coefficients

```
coef(mixed.logis)
```

```
$site
```

```
  (Intercept)      dbh
```

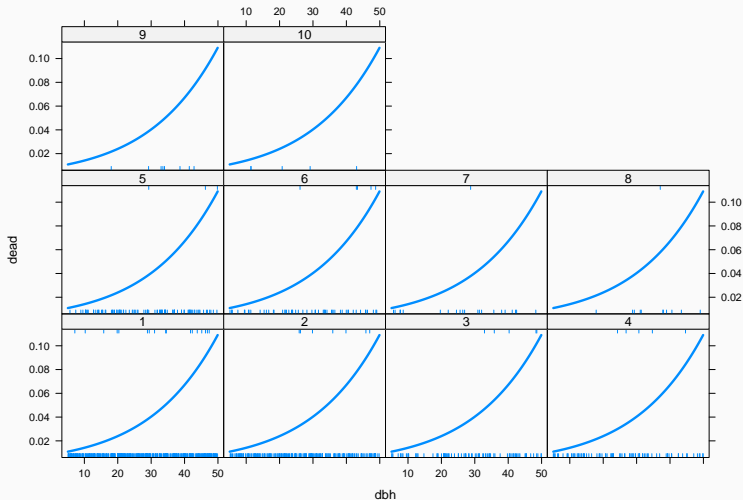
```
1   -4.778744  0.05364989
2   -4.778744  0.05364989
3   -4.778744  0.05364989
4   -4.778744  0.05364989
5   -4.778744  0.05364989
6   -4.778744  0.05364989
7   -4.778744  0.05364989
8   -4.778744  0.05364989
9   -4.778744  0.05364989
10  -4.778744  0.05364989
```

```
attr(,"class")
```

```
[1] "coef.mer"
```

Visualising model: visreg

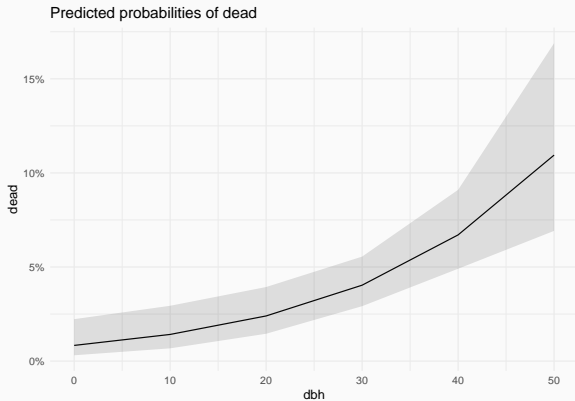
```
visreg(mixed.logis, xvar = "dbh", by = "site", scale = "response")
```



Visualising model: sjPlot

```
plot_model(mixed.logis, type = "eff", show.ci = TRUE)
```

\$dbh



- Perfect for **structured data** (space-time)

Advantages of multilevel models

- Perfect for **structured data** (space-time)
- Predictors enter at the appropriate level

Advantages of multilevel models

- Perfect for **structured data** (space-time)
- Predictors enter at the appropriate level
- Accommodate **variation** in treatment effects

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- Using all the data to perform inferences for groups with **small sample size**

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 - $y \sim x + (1 \mid \text{group})$

Formula syntax for different models

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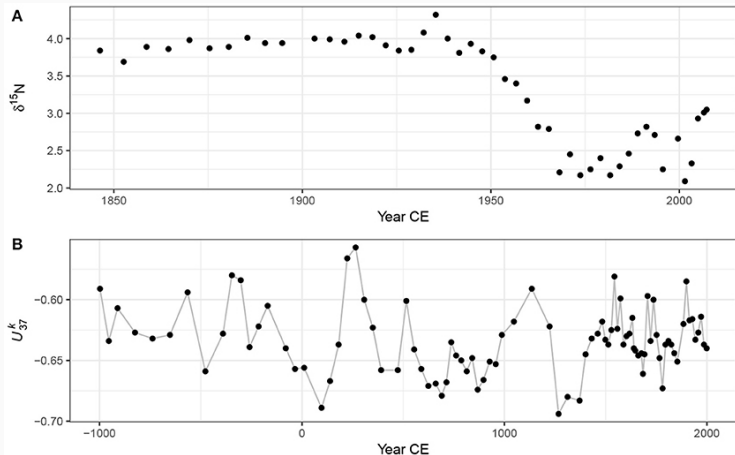
<https://bbolker.github.io/mixedmodels-misc/glmmFAQ.html>

Generalised Additive Models

Francisco Rodríguez-Sánchez

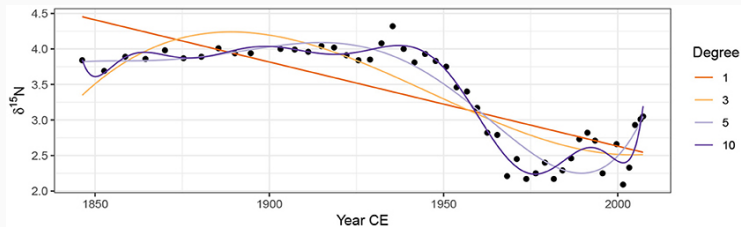
<https://frodriguezsanchez.net>

How do we model these time series?



Simpson 2018

How do we model these time series?



Simpson 2018

GAMs allow us to model non-linear relationships using smooths

Generalised Linear Model (GLM):

$$y = a + bx$$

Generalised Additive Model (GAM):

$$y = a + s(x)$$

Modelling non-linear time series with GAM

```
isotopes <- readRDS("data/isotope.rds")
```

	Depth	d13C	TotalC	d15N	TotalN	DryWeight	Year
1	0.2	-27.57	806.49	3.05	64.21	8.2	2007.254
2	0.4	-27.67	949.33	3.01	73.26	7.6	2006.510
3	0.8	-27.63	1305.52	2.93	93.25	11.6	2004.941
4	1.2	-27.62	1136.04	2.33	86.09	9.6	2003.269
5	1.6	-27.48	1028.27	2.09	93.80	10.9	2001.496
6	2.0	-27.39	809.91	2.66	79.98	9.9	1999.626

Modelling non-linear time series with GAM

```
library("mgcv")  
m <- gam(d15N ~ s(Year, k = 15), data = isotopes, method = "REML")
```

```
Family: gaussian  
Link function: identity
```

```
Formula:  
d15N ~ s(Year, k = 15)
```

```
Parametric coefficients:
```

```
      Estimate Std. Error t value Pr(>|t|)  
(Intercept) 3.30958    0.02622  126.2 <2e-16 ***
```

```
---
```

```
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
Approximate significance of smooth terms:
```

```
      edf Ref.df    F p-value  
s(Year) 9.282  11.07 61.33 <2e-16 ***
```

```
---
```

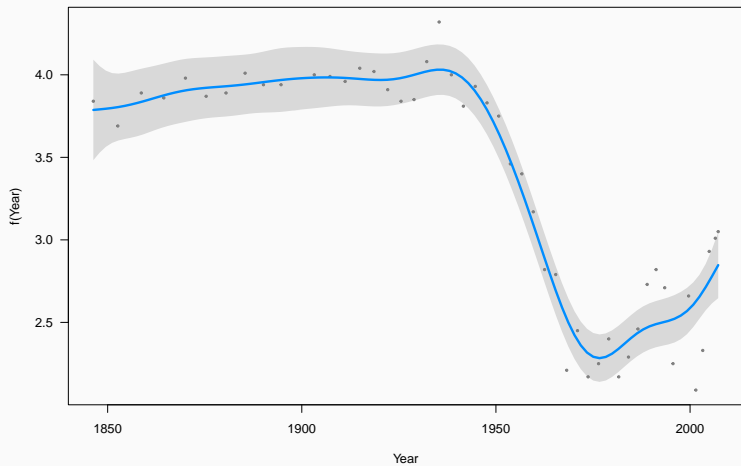
```
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
R-sq.(adj) = 0.935  Deviance explained = 94.8%
```

```
-REML = 3.9734  Scale est. = 0.03299  n = 48
```

Visualising fitted GAM

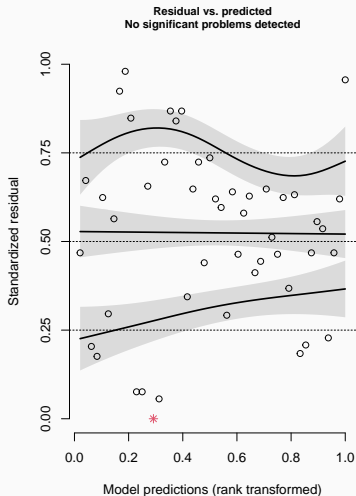
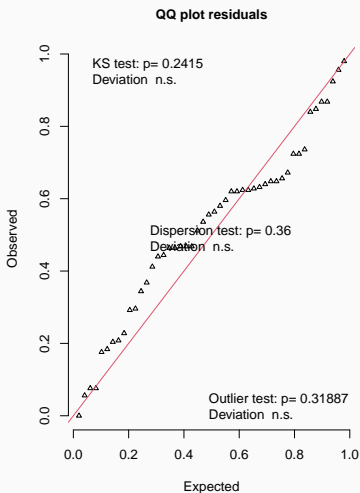
`visreg(m)`



Checking fitted GAM

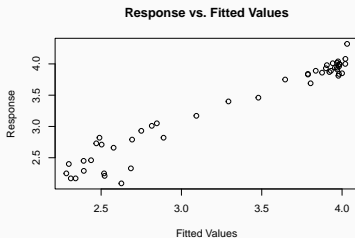
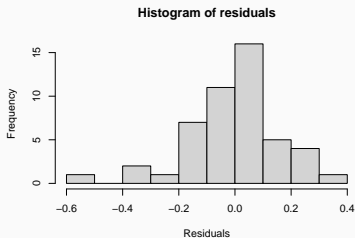
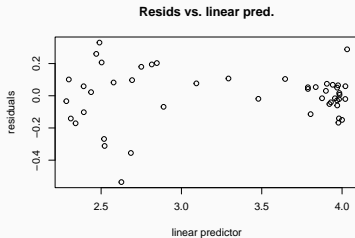
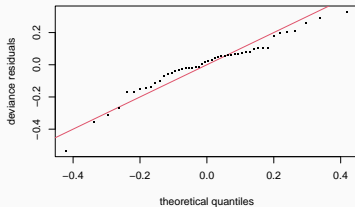
```
library("DHARMa")  
simulateResiduals(m, plot = TRUE)
```

DHARMa residual diagnostics



Checking fitted GAM

```
gam.check(m)
```



Method: REML Optimizer: outer newton

Including temporal autocorrelation

```
mod <- gamm(d15N ~ s(Year, k = 15), data = isotopes,  
            correlation = corCAR1(form = ~ Year), method = "REML")
```

Family: gaussian

Link function: identity

Formula:

```
d15N ~ s(Year, k = 15)
```

Parametric coefficients:

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	3.30909	0.03489	94.84	<2e-16 ***

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Approximate significance of smooth terms:

	edf	Ref.df	F	p-value
s(Year)	7.954	7.954	47.44	<2e-16 ***

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

R-sq.(adj) = 0.929

Modelling infant mortality

Modelling infant mortality

```
mort <- read.csv("data/UN_GDP_infantmortality.csv")
```

	country	infant.mortality	gdp
1	Afghanistan	154	2848
2	Albania	32	863
3	Algeria	44	1531
4	American.Samoa	11	NA
5	Andorra	NA	NA
6	Angola	124	355

Modelling infant mortality with a GLM

```
library("MASS")  
mort.glm <- glm.nb(infant.mortality ~ gdp, data = mort)
```

Call:

```
glm.nb(formula = infant.mortality ~ gdp, data = mort, init.theta = 2.460991808,  
       link = log)
```

Deviance Residuals:

Min	1Q	Median	3Q	Max
-2.8024	-1.0447	-0.3650	0.5232	2.9116

Coefficients:

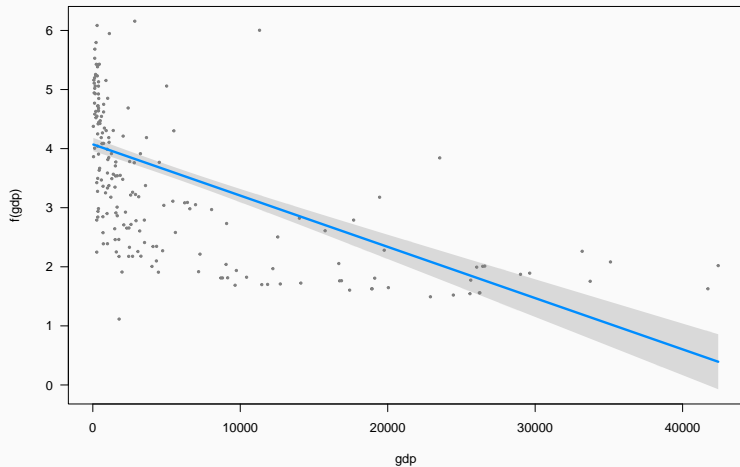
	Estimate	Std. Error	z value	Pr(> z)
(Intercept)	4.072e+00	5.727e-02	71.11	<2e-16 ***
gdp	-8.675e-05	6.221e-06	-13.95	<2e-16 ***

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

(Dispersion parameter for Negative Binomial(2.461) family taken to be 1)

Null deviance: 385.83 on 192 degrees of freedom
Residual deviance: 202.51 on 191 degrees of freedom

Modelling infant mortality with a GLM



Modelling infant mortality with a GLM (log.gdp)

```
mort$log.gdp <- log(mort$gdp)
mort.glm.log <- glm.nb(infant.mortality ~ log.gdp, data = mort)
```

Call:

```
glm.nb(formula = infant.mortality ~ log.gdp, data = mort, init.theta = 3.119314453,
       link = log)
```

Deviance Residuals:

Min	1Q	Median	3Q	Max
-2.7517	-0.8692	-0.3575	0.3090	4.5063

Coefficients:

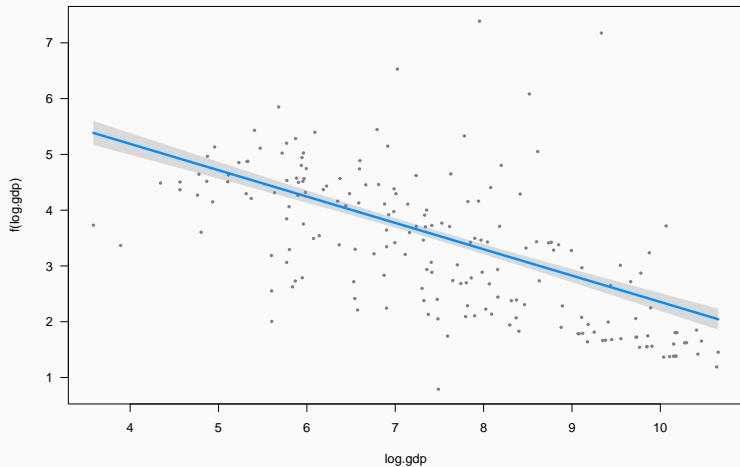
	Estimate	Std. Error	z value	Pr(> z)
(Intercept)	7.07818	0.20045	35.31	<2e-16 ***
log.gdp	-0.47238	0.02647	-17.85	<2e-16 ***

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

(Dispersion parameter for Negative Binomial(3.1193) family taken to be 1)

Null deviance: 478.54 on 192 degrees of freedom
Residual deviance: 198.03 on 191 degrees of freedom

Modelling infant mortality with a GLM (log.gdp)



Modelling infant mortality with a GAM

```
library("mgcv")  
mort.gam <- gam(infant.mortality ~ s(log.gdp), family = nb, data = mort)
```

Family: gaussian

Link function: identity

Formula:

d15N ~ s(Year, k = 15)

Parametric coefficients:

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	3.30958	0.02622	126.2	<2e-16 ***

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

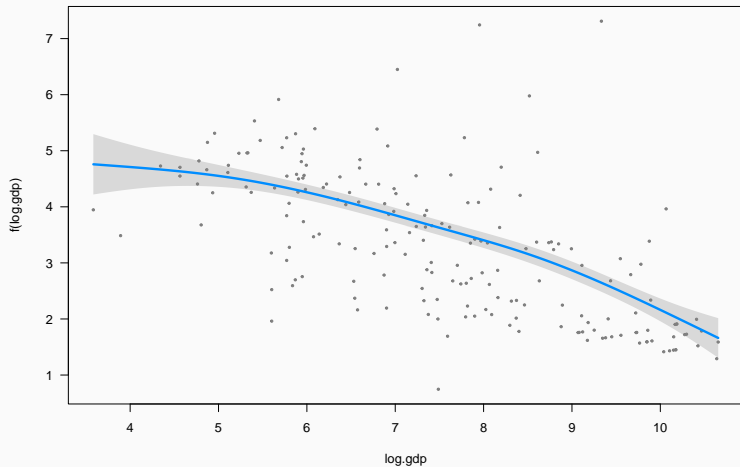
Approximate significance of smooth terms:

	edf	Ref.df	F	p-value
s(Year)	9.282	11.07	61.33	<2e-16 ***

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

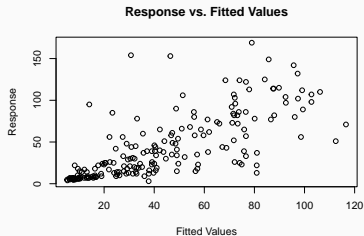
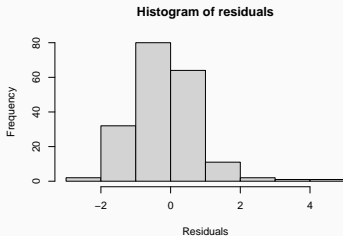
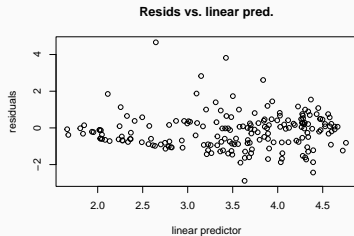
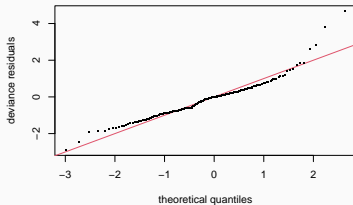
R-sq.(adj) = 0.935 Deviance explained = 94.8%

Modelling infant mortality with a GAM



Checking GAM

```
gam.check(mort.gam)
```



Method: REML Optimizer: outer newton

Comparing models

```
library("performance")  
compare_performance(mort.glm, mort.glm.log, mort.gam)
```

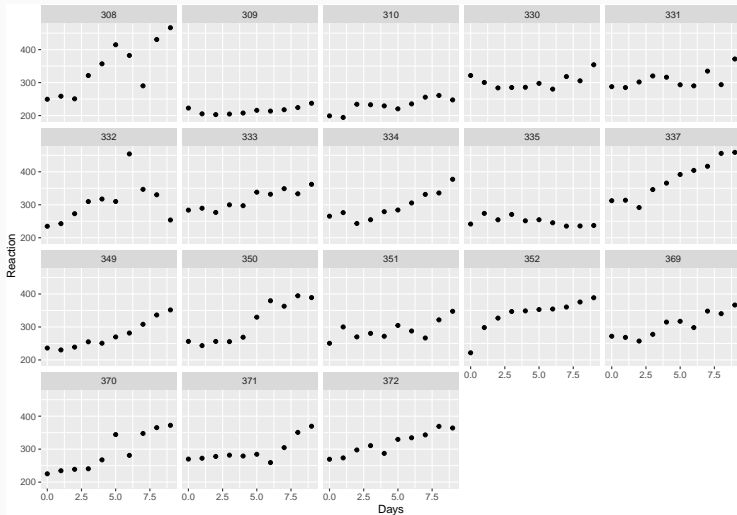
```
# Comparison of Model Performance Indices
```

Name	Model	AIC	BIC	RMSE	Sigma	Score
mort.glm	negbin	1714.957	1724.745	31.089	1.030	-
mort.glm.log	negbin	1667.750	1677.538	30.034	1.018	-
mort.gam	gam	1661.141	1680.512	26.249	1.027	-

Generalised Additive Mixed Models (GAMM)

Reaction time with sleep deprivation

```
library("lme4")  
data("sleepstudy")
```



Modelling reaction time with sleep deprivation (GAMM)

```
sgamm <- gam(Reaction ~ s(Days, Subject, k = 3, bs = "fs"),  
            data = sleepstudy, method = "REML")
```

Family: gaussian

Link function: identity

Formula:

```
Reaction ~ s(Days, Subject, k = 3, bs = "fs")
```

Parametric coefficients:

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	295.22	10.49	28.15	<2e-16 ***

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Approximate significance of smooth terms:

	edf	Ref.df	F	p-value
s(Days,Subject)	42.2	53	16.05	<2e-16 ***

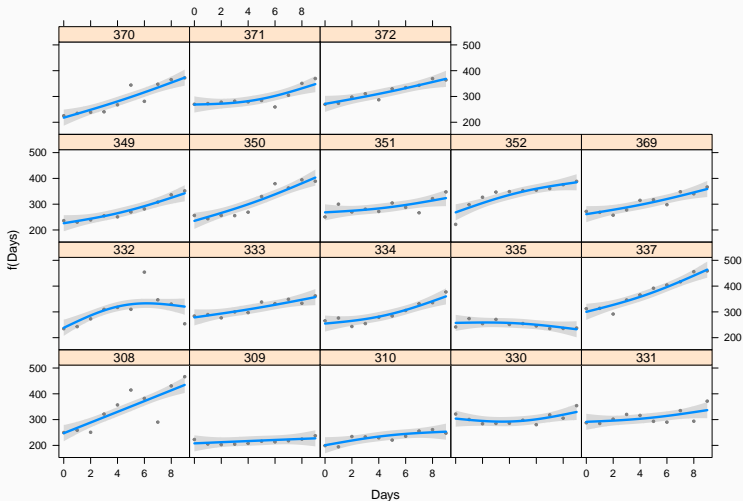
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

R-sq.(adj) = 0.826 Deviance explained = 86.7%

-REML = 886.71 Scale est. = 551.61 n = 180

Modelling reaction time with sleep deprivation (GAMM)

```
visreg(sgamma, xvar = "Days", by = "Subject")
```



An introduction to Bayesian modelling with brms and Stan

Francisco Rodríguez-Sánchez

<https://frodriguezsanchez.net>

Our dataset: tree heights and DBH

- One species

```
trees <- read.csv("data/trees.csv")
```

	site	dbh	height
Min.	: 1.0	Min. : 5.06	Min. :13.40
1st Qu.:	1.0	1st Qu.:17.69	1st Qu.:29.68
Median :	2.0	Median :28.62	Median :36.55
Mean :	2.7	Mean :27.88	Mean :36.51
3rd Qu.:	4.0	3rd Qu.:38.97	3rd Qu.:43.33
Max.	:10.0	Max. :49.92	Max. :59.30

Our dataset: tree heights and DBH

- One species
- 10 plots

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trees <- read.csv("data/trees.csv")
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Min.	: 1.0	Min. : 5.06	Min. :13.40
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Our dataset: tree heights and DBH

- One species
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- 1000 trees

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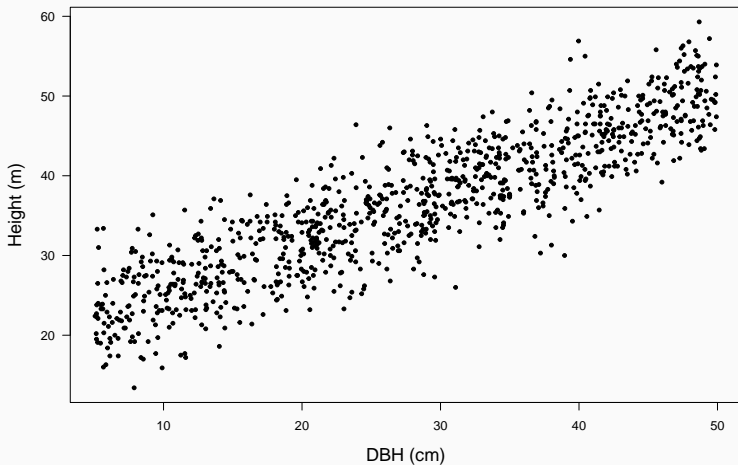
Our dataset: tree heights and DBH

- One species
- 10 plots
- 1000 trees
- Number of trees per plot ranging from 4 to 392

```
trees <- read.csv("data/trees.csv")
```

	site	dbh	height
Min.	: 1.0	Min. : 5.06	Min. :13.40
1st Qu.:	1.0	1st Qu.:17.69	1st Qu.:29.68
Median :	2.0	Median :28.62	Median :36.55
Mean :	2.7	Mean :27.88	Mean :36.51
3rd Qu.:	4.0	3rd Qu.:38.97	3rd Qu.:43.33
Max.	:10.0	Max. :49.92	Max. :59.30

What's the relationship between DBH and height?



First step: linear regression (lm)

```
simple.lm <- lm(height ~ dbh, data = trees)
```

Call:

```
lm(formula = height ~ dbh, data = trees)
```

Residuals:

Min	1Q	Median	3Q	Max
-13.3270	-2.8978	0.1057	2.7924	12.9511

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	19.33920	0.31064	62.26	<2e-16 ***
dbh	0.61570	0.01013	60.79	<2e-16 ***

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Residual standard error: 4.093 on 998 degrees of freedom

Multiple R-squared: 0.7874, Adjusted R-squared: 0.7871

F-statistic: 3695 on 1 and 998 DF, p-value: < 2.2e-16

Center continuous variables

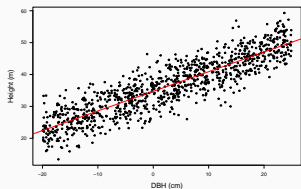
```
summary(trees$dbh)
```

Min.	1st Qu.	Median	Mean	3rd Qu.	Max.
5.06	17.69	28.62	27.88	38.97	49.92

```
trees$dbh.c <- trees$dbh - 25
```

So, all parameters will be referred to a 25 cm DBH tree.

Linear regression with centred DBH



```
lm(formula = height ~ dbh.c, data = trees
```

```
      coef.est coef.se
```

```
(Intercept) 34.73    0.13
```

```
dbh.c        0.62    0.01
```

```
---
```

```
n = 1000, k = 2
```

```
residual sd = 4.09, R-Squared = 0.79
```

Let's make it Bayesian

Remember our model structure

$$y_i \sim N(\mu_i, \sigma^2)$$

$$\mu_i = \alpha + \beta x_i$$

In this case:

$$\text{Height}_i \sim N(\mu_i, \sigma^2)$$

$$\mu_i = \alpha + \beta \text{DBH}_i$$

α : expected height when DBH = 25 cm

β : how much height increases with every unit increase of DBH

Defining model formula

```
library("brms")  
  
height.formu <- brmsformula(height ~ dbh.c)
```

We must define **prior distributions** for every parameter

brms default priors

```
get_prior(height.formu, data = trees)
```

```
          prior      class  coef group resp dpar nlpar bound
          (flat)         b
          (flat)         b dbh.c
student_t(3, 36.5, 10.2) Intercept
  student_t(3, 0, 10.2)      sigma
  source
  default
(vectorized)
  default
  default
```

Choosing priors

Avoid 'non-informative' priors

Use *weakly informative* (e.g. relatively wide Normal or t-student distributions) or *strongly informative* priors based on previous knowledge and common sense.

Some tips for setting priors:

- <https://github.com/stan-dev/stan/wiki/Prior-Choice-Recommendations>

Run **prior predictive checks** (just priors, no data)

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- http://www.mrc-bsu.cam.ac.uk/wp-content/uploads/bugsbook_chapter5.pdf

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Choosing priors

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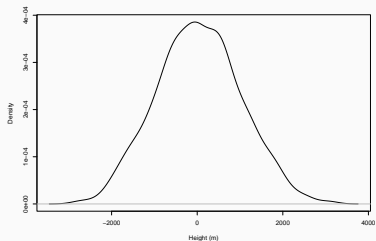
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- http://www.mrc-bsu.cam.ac.uk/wp-content/uploads/bugsbook_chapter5.pdf
- <https://doi.org/10.1111/oik.05985>

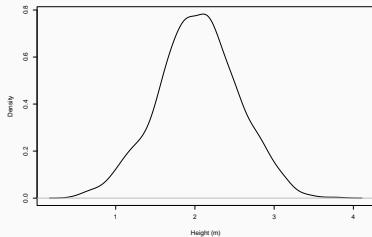
Run **prior predictive checks** (just priors, no data)

Example: estimating people height across countries

Unreasonable prior



Reasonable prior

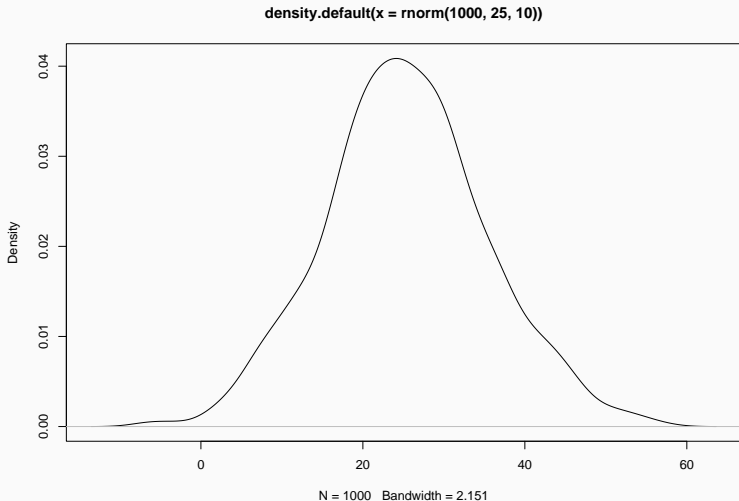


Defining priors for our trees example

```
priors <- c(  
  set_prior("normal(30, 10)", class = "Intercept"),  
  set_prior("normal(0.5, 0.4)", class = "b"),  
  set_prior("normal(0, 5)", class = "sigma")  
)
```

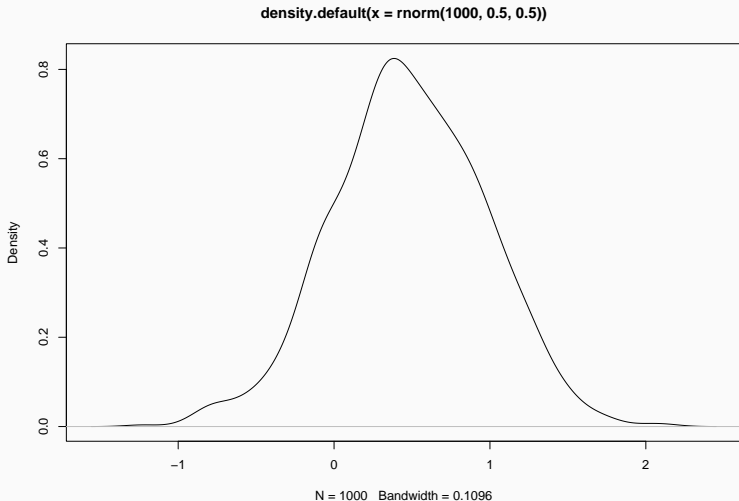
Prior for intercept (average height of 25-cm diameter tree)

```
plot(density(rnorm(1000, 25, 10)))
```



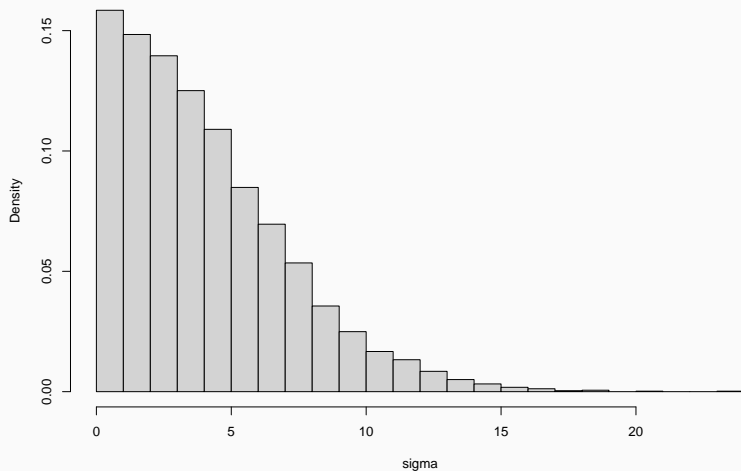
Prior for slope

```
plot(density(rnorm(1000, 0.5, 0.5)))
```



Prior for sigma (residual sd)

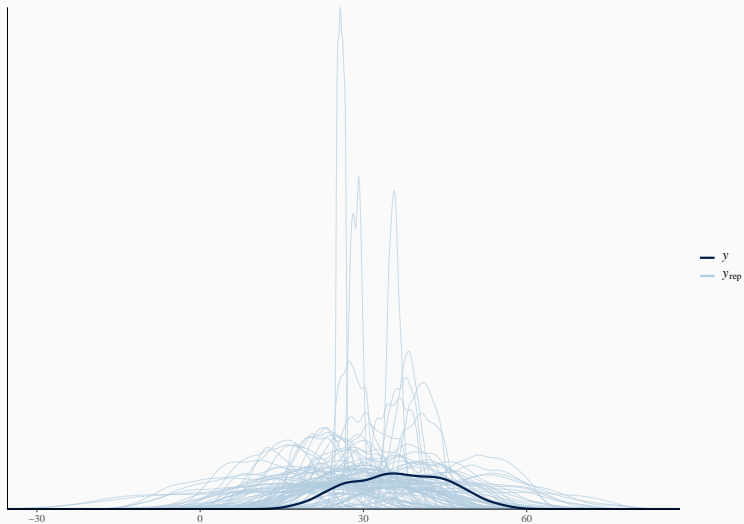
Histogram of sigma



```
height.mod <- brm(height.formu,  
  data = trees,  
  prior = priors,  
  sample_prior = "only")
```

Prior predictive check

```
pp_check(height.mod, ndraws = 100)
```



Fit model (now with data)

```
height.mod <- brm(height.formu,  
  data = trees,  
  prior = priors)
```

Model summary

```
summary(height.mod)
```

Family: gaussian

Links: mu = identity; sigma = identity

Formula: height ~ dbh.c

Data: trees (Number of observations: 1000)

Draws: 4 chains, each with iter = 2000; warmup = 1000; thin = 1;
total post-warmup draws = 4000

Population-Level Effects:

	Estimate	Est.Error	l-95% CI	u-95% CI	Rhat	Bulk_ESS	Tail_ESS
Intercept	34.73	0.13	34.47	34.98	1.00	3620	2251
dbh.c	0.62	0.01	0.60	0.64	1.00	4523	3133

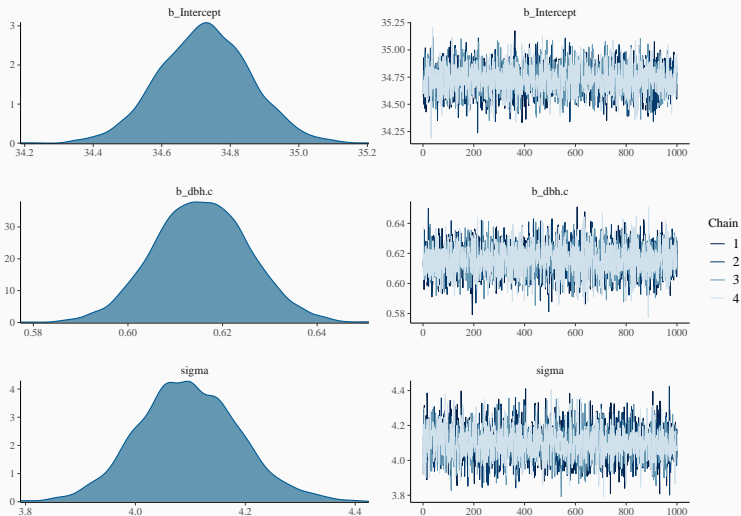
Family Specific Parameters:

	Estimate	Est.Error	l-95% CI	u-95% CI	Rhat	Bulk_ESS	Tail_ESS
sigma	4.10	0.09	3.92	4.29	1.00	4176	2913

Draws were sampled using sampling(NUTS). For each parameter, Bulk_ESS and Tail_ESS are effective sample size measures, and Rhat is the potential scale reduction factor on split chains (at convergence, Rhat = 1).

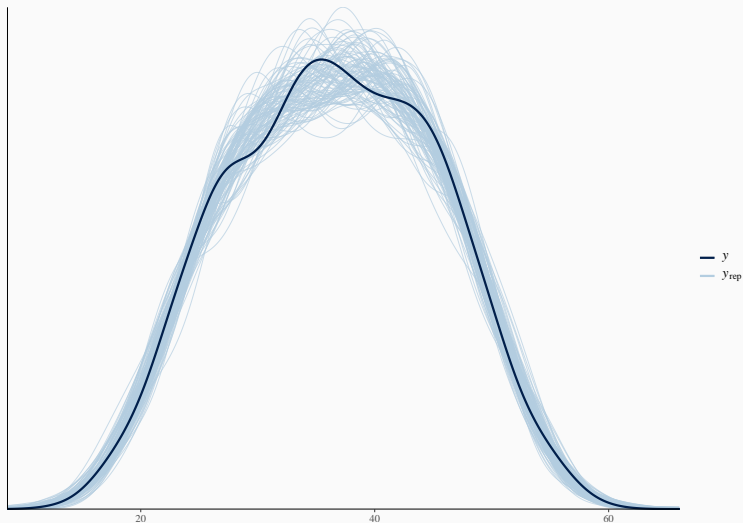
Model visualisation

```
plot(height.mod)
```



Posterior predictive checking

```
pp_check(height.mod, ndraws = 100)
```



```
library("shinytan")  
launch_shinytan(height.mod)
```

height ~ sex